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(54) Title: METHODS AND COMPOSITIONS FOR MAKING EMAMECTIN

(57) Abstract: Disclosed is a family of P450 monooxygenases, each member of which regioselectively oxidizes avermectin to 4"-keto-avermectin. The P450 monooxygenases find use in methods and formulations for making emamectin from avermectin. Also disclosed are methods for purifying the P450 monooxygenases of the invention, binding agents that specifically bind to the P450 monooxygenases of the invention, and genetically engineered cells that express the P450 monooxygenases of the invention. Also disclosed are ferrodoxins and ferredoxin reductases that are active with the P450 monooxygenases of the invention.

METHODS AND COMPOSITIONS FOR MAKING EMAMECTIN

The invention relates to the field of agrochemicals, and in particular, to insecticides. More specifically, this invention relates to the derivatization of avermectin, particularly for the synthesis of emamectin.

Emamectin is a potent insecticide and controls many pests such as thrips, leafminers, and worm pests including alfalfa caterpillar, beet armyworm, cabbage looper, corn earworm, cutworms, diamondback moth, tobacco budworm, tomato fruitworm, and tomato pinworm. Emamectin (4"-deoxy-4"-epi-N-methylamino avermectin Bla/Blb) is described in U.S. Patent No. 4,874,749 and in Cvetovich, R.J. *et al.*, *J. Organic Chem.* **59**:7704-7708, 1994 (as MK-244).

U.S. Patent No. 5,288,710 describes salts of emamectin that are especially valuable agrochemically. These salts of emamectin are valuable pesticides, especially for combating insects and representatives of the order Acarina. Some pests for which emamectin is useful in combating are listed in European Patent Application EP-A 736,252.

One drawback to the use of emamectin is the difficulty of its synthesis from avermectin. This is due to the first step of the process, which is the most costly and time-consuming step of producing emamectin, in which the 4"-carbinol group of avermectin must be oxidized to a ketone. The oxidation of the 4"-carbinol group is problematic due to the presence of two other hydroxyl groups on the molecule that must be chemically protected before oxidation and deprotected after oxidation. Thus, this first step, significantly increases the overall cost and time of producing emamectin from avermectin.

Because of the efficacy and potency of emamectin as an insecticide, there is a need to develop a cost and time effective method and/or reagent for regioselectively oxidizing the 4"-carbinol group of avermectin to produce 4"-keto-avermectin, which is a necessary intermediate for producing emamectin from avermectin.

The invention provides a novel family of P450 monooxygenases, each member of which is able to regioselectively oxidize the 4"-carbinol group of unprotected avermectin, thereby resulting in a cheap, effective method to produce 4"-keto-avermectin, a necessary intermediate in the production of emamectin. The invention allows elimination of the costly, time-consuming steps of (1) chemically protecting the two other hydroxyl groups on the avermectin

molecule prior to oxidation of the 4"-carbinol group that must be chemically protected before oxidation; and (2) chemically deprotecting these two other hydroxyl groups after oxidation. The invention thus provides reagents and methods for significantly reducing the overall cost of producing emamectin from avermectin.

- Accordingly, in one aspect, the invention provides a purified nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In a specific embodiment , the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin, which polypeptide is substantially similar, and preferably has between at least 50%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2, with each individual number within this range of between 50% and 99% also being part of the invention.
- In a further specific embodiment , the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO:2.
- The invention further provides a purified nucleic acid molecule comprising a nucleotide sequence
 - a) as given in SEQ ID NO:1;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
 - e) complementary to (a), (b) or (c);
 - f) which is the reverse complement of (a), (b) or (c), or

- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In a specific embodiment , the invention relates to a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin, which polypeptide is substantially similar, and preferably has at least between 60%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95, with each individual number within this range of between 60% and 99% also being part of the invention.
- In a further specific embodiment , the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- The invention further provides a purified nucleic acid molecule comprising a nucleotide sequence
 - a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID

- NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;
- e) complementary to (a), (b) or (c);
 - f) which is the reverse complement of (a), (b) or (c); or
 - g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.

In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least between 66%, and 99% identical to SEQ ID NO:1, with each individual number within this range of between 66%, and 99% also being part of the invention..

- In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least between 70%, and 99% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94, with each individual number within this range of between 70%, and 99% also being part of the invention..
- In some embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 80% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.
- In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 90% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.

- In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 95% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.
- In some embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, and SEQ ID NO:94.
- In particular embodiments, the nucleic acid molecule is isolated from a *Streptomyces* strain. In certain embodiments, the *Streptomyces* strain is selected from the group consisting of *Streptomyces tubercidicus*, *Streptomyces lydicus*, *Streptomyces platensis*, *Streptomyces chattanoogensis*, *Streptomyces kasugaensis*, and *Streptomyces rimosus* and *Streptomyces albofaciens*.
- In some embodiments of this aspect, the nucleic acid molecule further comprises a nucleic acid sequence encoding a tag which is linked to the P450 monooxygenase via a covalent bond. In certain embodiments, the tag is selected from the group consisting of a His tag, a GST tag, an HA tag, a HSV tag, a Myc-tag, and VSV-G-Tag.
- In another aspect, the invention provides a purified polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In some embodiments, the polypeptide comprises or consists essentially of an amino acid sequence that is encoded by a nucleic acid molecule
 - a) as given in SEQ ID NO:1 or the complement thereof;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
 - e) complementary to (a), (b) or (c);

- f) which is the reverse complement of (a), (b) or (c); or;
- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.

In some embodiments, the polypeptide comprises or consists essentially of an amino acid sequence that is between at least 50%, and 99% identical to SEQ ID NO:2, with each individual number within this range of between 50% and 99% also being part of the invention..

- In some embodiments, the polypeptide comprises or consists essentially of an amino acid sequence that is encoded by a nucleic acid molecule
 - a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof, or the complement thereof;
 - e) complementary to (a), (b) or (c);
 - f) which is the reverse complement of (a), (b) or (c); or
 - g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In some embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is between at least 60%, and 99% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24,

SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95, with each individual number within this range of between 60% and 99% also being part of the invention..

- In certain embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 70% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 90% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In certain embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 95% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments of this aspect of the invention, the P450 monooxygenase comprises or consists essentially of an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, and SEQ ID NO:95.
- In certain embodiments, the polypeptide according to the invention exhibiting an enzymatic activity of a P450 monooxygenase further comprises a tag. In some

embodiments, the tag is selected from the group consisting of a His tag, a GST tag, an HA tag, a HSV tag, a Myc-tag, and VSV-G-Tag.

- In another aspect, the invention provides a binding agent that specifically binds to a polypeptide according to the invention exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In some embodiments, the binding agent is an antibody. In certain embodiments, the antibody is a polyclonal antibody or a monoclonal antibody.
- In yet another aspect, the invention provides a family of P450 monooxygenase polypeptides, wherein each member of the family regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is between at least 50%, and 99% identical to SEQ ID NO:2, with each individual number within this range of between 50% and 99% also being part of the invention..
- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is between at least 60%, and 99% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95, with each individual number within this range of between 60% and 99% also being part of the invention..
- In some embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 70% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95. In

some embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 90% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 95% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments of this aspect of the invention, each member of the family comprises or consists essentially of an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, and SEQ ID NO:95.
- In still another aspect, the invention provides a cell genetically engineered to comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In some embodiments, the nucleic acid molecule is positioned for expression in the cell. In certain embodiments, the cell further comprises a nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin protein.
- In some embodiments, the cell further comprises a nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase protein.
- In certain embodiments, the cell is a genetically engineered *Streptomyces* strain. In certain embodiments, the cell is a genetically engineered *Streptomyces lividans* strain. In particular embodiments, the genetically engineered *Streptomyces lividans* strain has NRRL Designation No. B-30478. In some embodiments, the cell is a genetically engineered *Pseudomonas* strain. In some embodiments, the cell is a genetically engineered

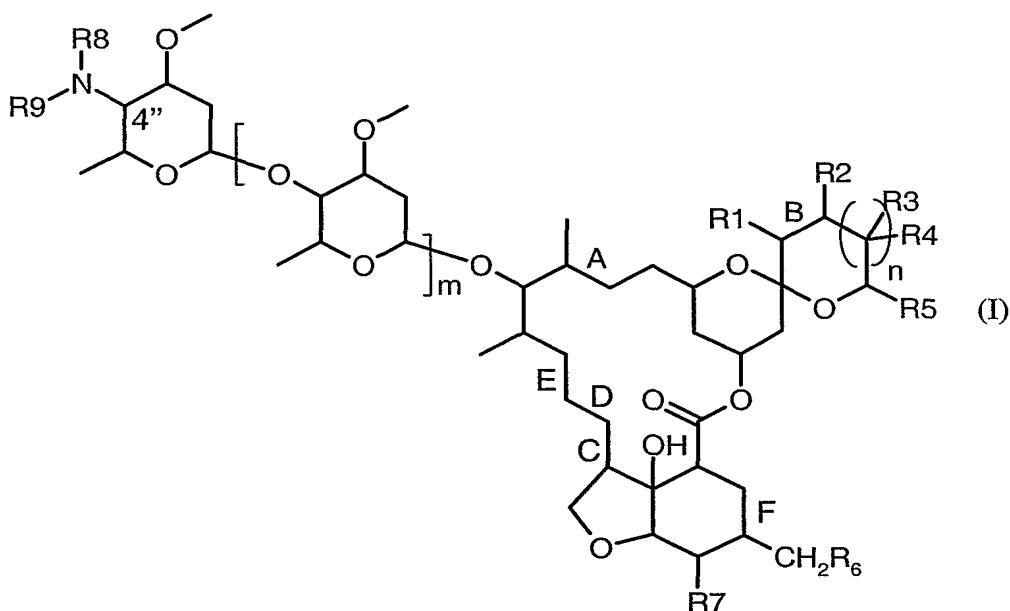
Pseudomonas putida strain. In certain embodiments, the genetically engineered *Pseudomonas putida* strain has NRRL Designation No. B-30479. In some embodiments, the cell is a genetically engineered *Escherichia coli* strain.

- In another aspect, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In a specific embodiment , the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits the enzymatic activity of a ferredoxin, which polypeptide is substantially similar, and preferably has between at least 80%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:36 or SEQ ID NO: 38, with each individual number within this range of between 80% and 99% also being part of the invention.
- In still a further specific embodiment , the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits the enzymatic activity of a ferredoxin, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO: 36 or SEQ ID NO: 38.
- The invention further provides a purified nucleic acid molecule comprising a nucleotide sequence
 - a) as given in SEQ ID NO:35 or SEQ ID NO: 37;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO: 35 or SEQ ID NO: 37, or the complement thereof;
 - e) complementary to (a), (b) or (c);
 - f) which is the reverse complement of (a), (b) or (c); or
 - g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a ferredoxin and regioselectively oxidizes avermectin to 4"-keto-avermectin.

- In certain embodiments, the nucleic acid molecule encoding a ferredoxin of the invention comprises or consists essentially of a nucleic acid sequence that is at least 81% identical to SEQ ID NO:35 or SEQ ID NO:37. In some embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:35 or SEQ ID NO:37. In certain embodiments, the nucleic acid molecule encoding a ferredoxin of the invention comprises or consists essentially of the nucleic acid sequence of SEQ ID NO:35 or SEQ ID NO:37.
- In yet another aspect, the invention provides a purified ferredoxin protein, wherein the ferredoxin protein is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In certain embodiments, the ferredoxin of the invention comprises or consists essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:36 or SEQ ID NO:38. In some embodiments, the nucleic acid molecule comprises or consists essentially of an amino acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:36 or SEQ ID NO:38.
- In particular embodiments, the ferredoxin of the invention comprises or consists essentially of the amino acid sequence of SEQ ID NO:36 or SEQ ID NO:38.
- In another aspect, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In certain embodiments, the nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase comprises or consists essentially of the nucleic acid sequence of SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:104.
- In yet another aspect, the invention provides a purified polypeptide exhibiting an enzymatic activity of a ferredoxin reductase protein, wherein the said polypeptide is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In certain embodiments, the

polypeptide of the invention comprises or consists essentially of the amino acid sequence of SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, or SEQ ID NO:105.

- In another aspect, the invention provides a process for the preparation a compound of the formula



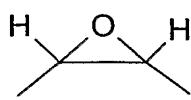
in which

R₁-R₉ represent, independently of each other hydrogen or a substituent;

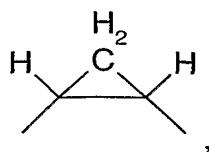
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula

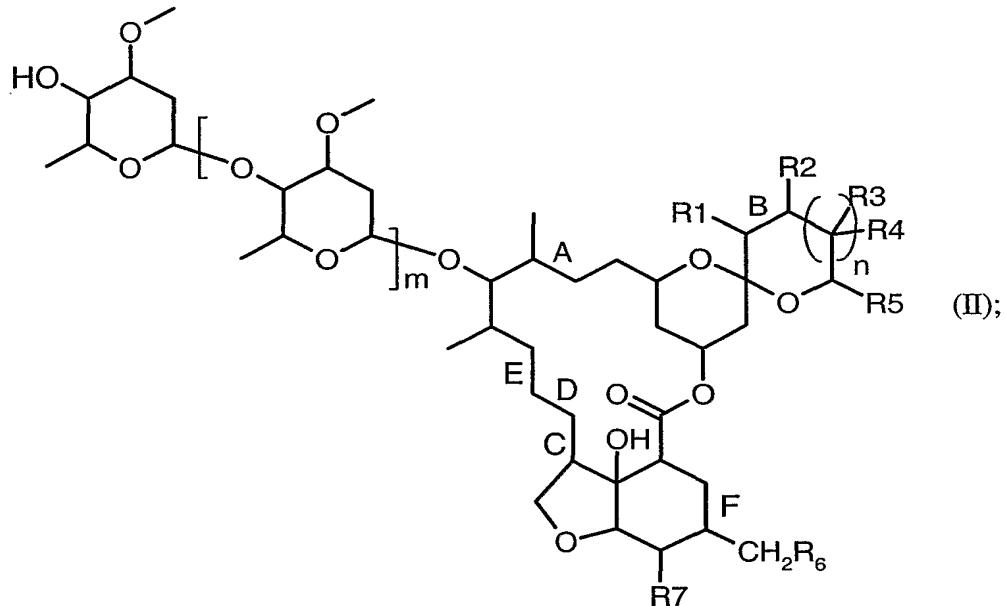


, or a single bond and a methylene bridge of the formula



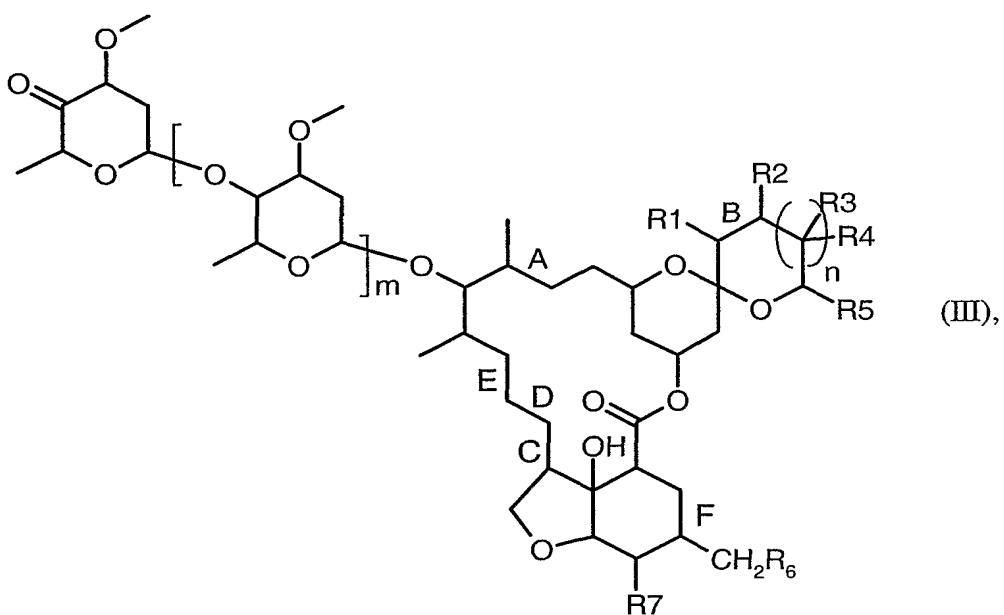
including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
 which process comprises

- 1) bringing a compound of the formula



wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above,
 into contact with a polypeptide according to the invention that is capable of
 regioselectively oxidising the alcohol at position 4" in order to form a compound of the
 formula



in which R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , m , n , A , B , C , D , E and F have the meanings given for formula (I); and

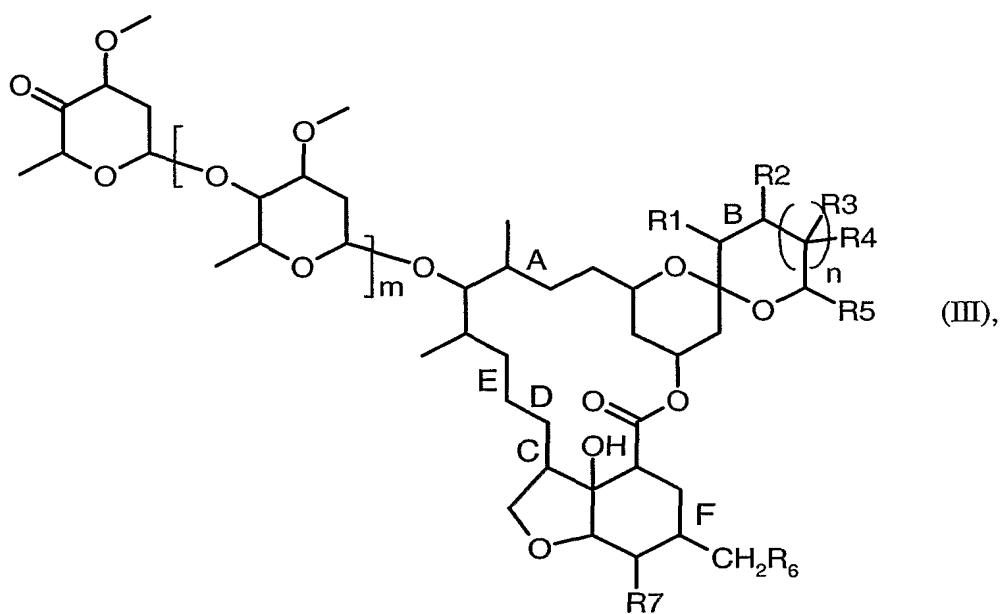
2) reacting the compound of the formula (III) with an amine of the formula $\text{HN}(\text{R}_8)\text{R}_9$, wherein R_8 and R_9 have the same meanings as given for formula (I), and which is known, in the presence of a reducing agent;

and, in each case, if desired, converting a compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, into a different compound of formula (I) or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, separating a mixture of E/Z isomers obtainable in accordance with the process and isolating the desired isomer, and/or converting a free compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, into a salt or converting a salt, obtainable in accordance with the process or by another method, of a compound of formula (I) or of an E/Z isomer or tautomer thereof into the free compound of formula (I) or an E/Z isomer or tautomer thereof or into a different salt.

- In some embodiments, the compound of formula (II) is further brought into contact with a polypeptide according to the invention exhibiting an enzymatic activity of a

ferredoxin. In certain embodiments, the compound of formula (II) is further brought into contact with a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase. In some embodiments, the compound of formula (II) is further brought into contact with a reducing agent (*e.g.*, NADH or NADPH).

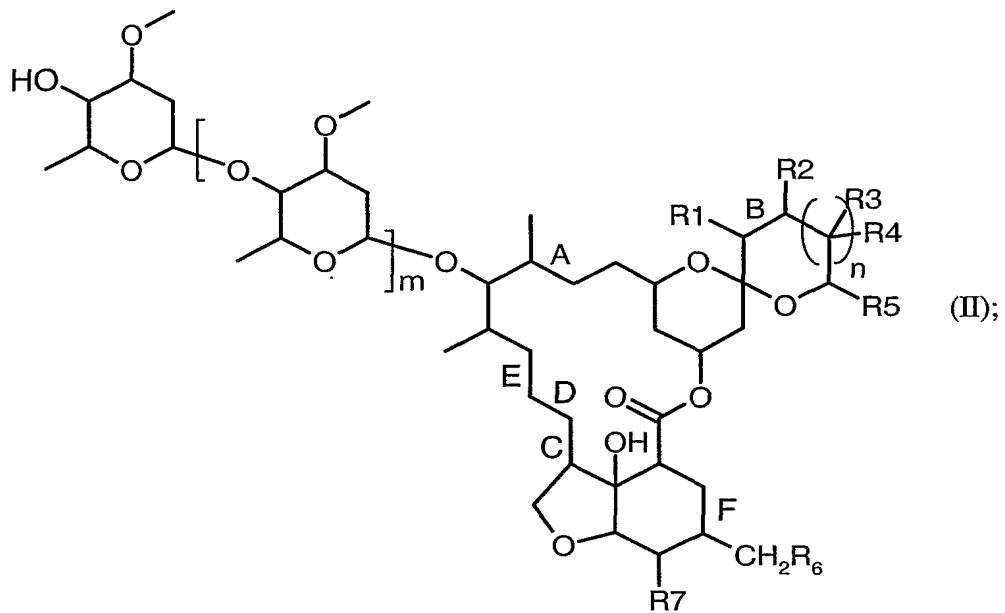
- In still a further embodiment, the invention provides a process for the preparation of a compound of the formula



in which R₁, R₂, R₃, R₄, R₅, R₆, R₇, m, n, A, B, C, D, E and F have the meanings given for formula (I) of claim 1,

which process comprises

- 1) bringing a compound of the formula



wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regioselectively oxidising the alcohol at position 4", maintaining said contact for a time sufficient for the oxidation reaction to occur and isolating and purifying the compound of formula (II).

- In yet another embodiment, the invention provides a process according to the invention for the preparation of a compound of the formula (I), in which
 - n is 1;
 - m is 1;
 - A is a double bond;
 - B is single bond or a double bond;
 - C is a double bond;
 - D is a single bond;
 - E is a double bond;
 - F is a double bond; or a single bond and a epoxy bridge; or a single bond and a methylene bridge;
 - R₁, R₂ and R₃ are H;

R₄ is methyl;

R₅ is C₁-C₁₀-alkyl, C₃-C₈-cycloalkyl or C₂-C₁₀-alkenyl;

R₆ is H;

R₇ is OH;

R₈ and R₉ are independently of each other H; C₁-C₁₀-alkyl or C₁-C₁₀-acyl; or together form -(CH₂)_q-; and

q is 4, 5 or 6.

- In still another embodiment, the invention provides a process according to the invention for the preparation of a compound of the formula (I), in which

n is 1;

m is 1;

A, B, C, E and F are double bonds;

D is a single bond;

R₁, R₂, and R₃ are H;

R₄ is methyl;

R₅ is s-butyl or isopropyl;

R₆ is H;

R₇ is OH;

R₈ is methyl

R₉ is H.

- In still another embodiment, the invention provides a process according to the invention for the preparation of 4"-deoxy-4"-N-methylamino avermectin B_{1a}/B_{1b} benzoate salt.

- In another aspect, the invention provides a method for making emamectin. The method comprises adding a polypeptide according to the invention exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin to a reaction mixture comprising avermectin and incubating the reaction mixture under conditions that allow the polypeptide to regioselectively oxidize avermectin to 4"-keto-avermectin. In some embodiments, the reaction mixture further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin. In

certain embodiments, the reaction mixture further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase. In some embodiments, the reaction mixture further comprises a reducing agent (*e.g.*, NADH or NADPH).

- In still another aspect, the invention provides a formulation for making a compound of formula (I) comprising a polypeptide according to the invention exhibiting a P450 monooxygenase activity that is capable of regioselectively oxidising the alcohol at position 4" in order to form a compound of formula (II). In some embodiments, the formulation further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin (*e.g.*, a ferredoxin from cell or strain from which the P450 monooxygenase was isolated or derived).
- In still another aspect, the invention provides a formulation for making emamectin comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In some embodiments, the formulation further comprises a ferredoxin (*e.g.*, a ferredoxin from cell or strain from which the P450 monooxygenase was isolated or derived).
- In certain embodiments, the formulation further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase (*e.g.*, a ferredoxin from cell or strain from which the P450 monooxygenase was isolated or derived). In some embodiments, the formulation further comprises a reducing agent (*e.g.*, NADH or NADPH).

Brief Description of the Drawings

Figure 1 is a diagrammatic representation showing a map of plasmid pTBBKA. Recognition sites by the indicated restriction endonucleases are shown, along with the location of the site in the nucleotide sequence of the plasmid. Also shown are genes (*e.g.*, kanamycin resistance "KanR"), and other functional aspects (*e.g.*, Tip promoter) contained in the plasmid.

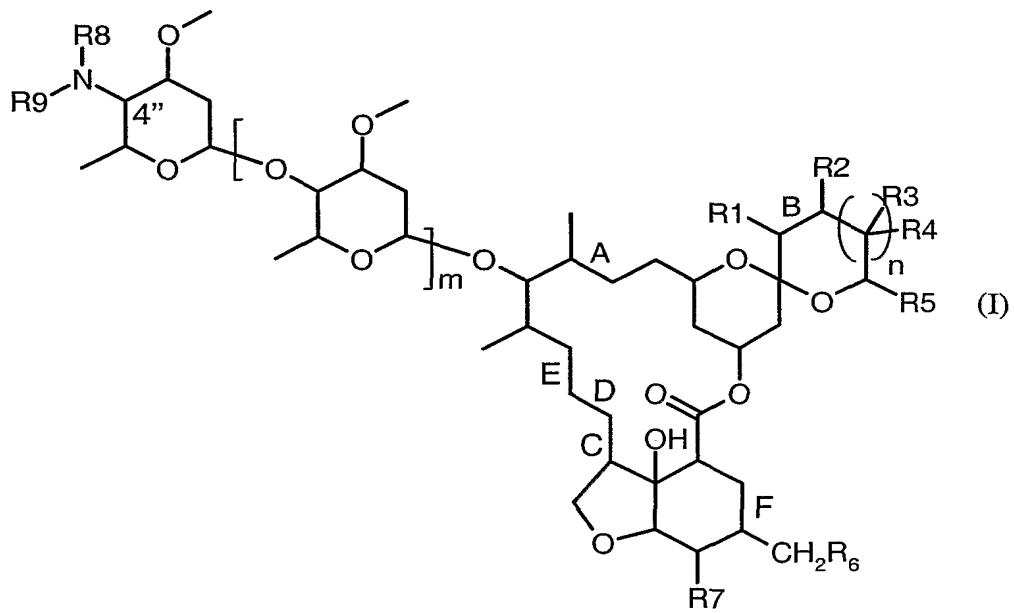
Figure 2 is a diagrammatic representation showing a map of plasmid pTUA1A. Recognition sites by the indicated restriction endonucleases are shown, along with the

location of the site in the nucleotide sequence of the plasmid. Also shown are genes (*e.g.*, ampicillin resistance “AmpR”) and other functional aspects (*e.g.*, Tip promoter) contained in the plasmid.

Figure 3 is a diagrammatic representation showing a map of plasmid pRK-*ema1/fd233*. This plasmid was derived by ligating a BglII fragment containing the *ema1* and *fd233* genes organized on a single transcriptional unit into the BglII site of the broad host-range plasmid pRK290. The *ema1/fd233* genes are expressed by the tac promoter (Ptac), and they are followed by the tac terminator (Ttac). Restriction endonuclease recognition sites shown are BglII “B”; EcoRI “E”; PacI “Pc”; PmeI “Pm”; and SalI “S.”

The present invention provides a family of polypeptides which exhibit an enzymatic activity of a P450 monooxygenases and are capable of regioselectively oxidizing the alcohol at position 4” of a compound of formula (II) such as avermectin in order to produce a compound of the formula (III), but especially 4”-keto-avermectin.

More particularly, the family of polypeptides according to the invention may be used in a process for the preparation a compound of the formula



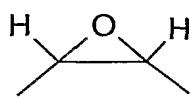
in which

R₁-R₉ represent, independently of each other hydrogen or a substituent;

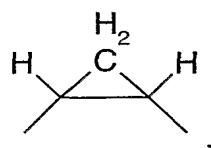
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula

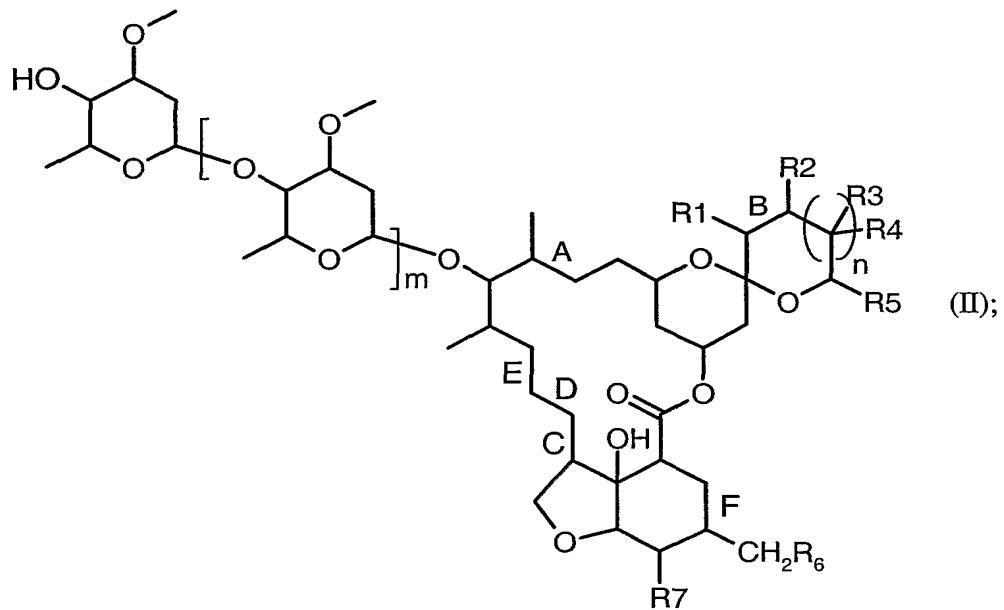


, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
which process comprises

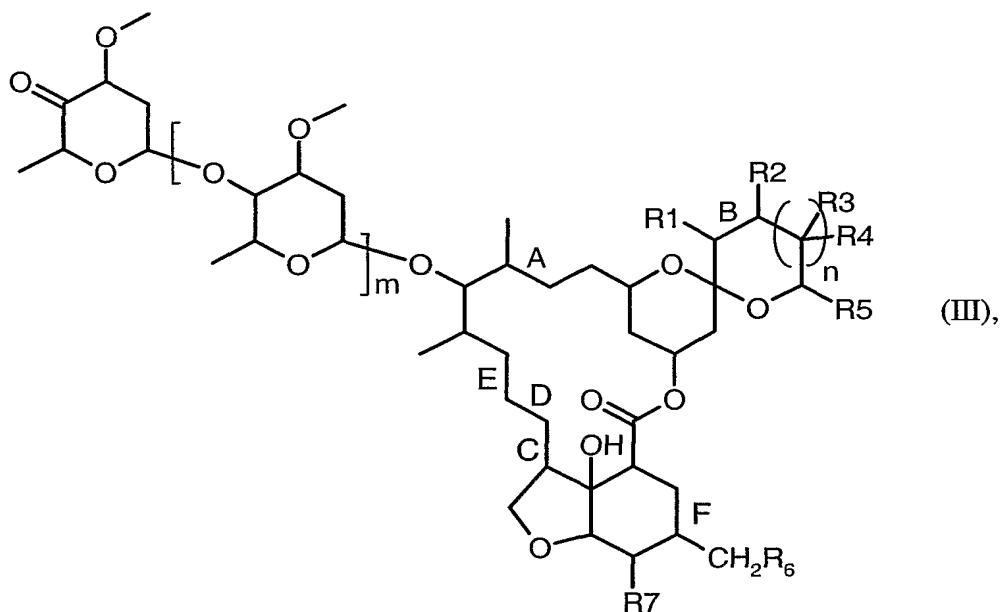
- 1) bringing a compound of the formula



wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above,

into contact with a polypeptide according to the invention which exhibits an enzymatic activity of a P450 monooxygenases and is capable of regioselectively oxidizing the alcohol at position 4" of formula (II) in order to produce a compound of the formula (III)



in which R₁, R₂, R₃, R₄, R₅, R₆, R₇, m, n, A, B, C, D, E and F have the meanings given for formula (I); and

2) reacting the compound of the formula (III) with an amine of the formula HN(R₈)R₉, wherein R₈ and R₉ have the same meanings as given for formula (I), and which is known, in the presence of a reducing agent;

and, in each case, if desired, converting a compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, into a different compound of formula (I) or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, separating a mixture of E/Z isomers obtainable in accordance with the process and isolating the desired isomer, and/or converting a free compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, into a salt or converting a salt, obtainable in accordance with the process or by another method, of a compound of formula (I) or of an E/Z isomer or tautomer thereof into the free compound of formula (I) or an E/Z isomer or tautomer thereof or into a different salt.

Methods of synthesis for the compounds of formula (I) are described in the literature. It has been found, however, that the processes known in the literature cause considerable problems during production basically on account of the low yields and the tedious procedures which have to be used. Accordingly, the known processes are not satisfactory in that respect, giving rise to the need to make available improved preparation processes for those compounds.

The compounds (I), (II) and (III) may be in the form of tautomers. Accordingly, hereinbefore and hereinafter, where appropriate the compounds (I), (II) and (III) are to be understood to include corresponding tautomers, even if the latter are not specifically mentioned in each case.

The compounds (I), (II) and (III) are capable of forming acid addition salts. Those salts are formed, for example, with strong inorganic acids, such as mineral acids, for example perchloric acid, sulfuric acid, nitric acid, nitrous acid, a phosphoric acid or a hydrohalic acid, with strong organic carboxylic acids, such as unsubstituted or substituted, for example halo-substituted, C₁-C₄alkanecarboxylic acids, for example acetic acid, saturated or unsaturated dicarboxylic acids, for example oxalic, malonic, succinic, maleic, fumaric or phthalic acid, hydroxycarboxylic acids, for example ascorbic, lactic, malic, tartaric or citric acid, or benzoic acid, or with organic sulfonic acids, such as unsubstituted or substituted, for example halo-substituted, C₁-C₄alkane- or aryl-sulfonic acids, for example methane- or p-toluene-sulfonic acid. Furthermore, compounds of formula (I), (II) and (III) having at least one acidic group are capable of forming salts with bases. Suitable salts with bases are, for example, metal salts, such as alkali metal or alkaline earth metal salts, for example sodium, potassium or magnesium salts, or salts with ammonia or an organic amine, such as morpholine, piperidine, pyrrolidine, a mono-, di- or tri-lower alkylamine, for example ethyl-, diethyl-, triethyl- or dimethyl-propyl-amine, or a mono-, di- or tri-hydroxy-lower alkylamine, for example mono-, di- or tri-ethanolamine. In addition, corresponding internal salts may also be formed. Preference is given within the scope of the invention to agrochemically advantageous salts. In view of the close relationship between the compounds of formula (I), (II) and (III) in free form and in the form of their salts, any reference hereinbefore or hereinafter to the free compounds of formula (I), (II) and (III) or to their respective salts is to be understood as including also the corresponding salts or the free compounds of formula (I), (II) and (III), where appropriate and

expedient. The same applies in the case of tautomers of compounds of formula (I), (II) and (III) and the salts thereof. The free form is generally preferred in each case.

Preferred within the scope of this invention is a process for the preparation of compounds of the formula (I), in which

n is 1;

m is 1;

A is a double bond;

B is single bond or a double bond,

C is a double bond,

D is a single bond,

E is a double bond,

F is a double bond; or a single bond and a epoxy bridge; or a single bond and a methylene bridge;

R₁, R₂ and R₃ are H;

R₄ is methyl;

R₅ is C₁-C₁₀-alkyl, C₃-C₈-cycloalkyl or C₂-C₁₀-alkenyl;

R₆ is H;

R₇ is OH;

R₈ and R₉ are independently of each other H; C₁-C₁₀-alkyl or C₁-C₁₀-acyl; or together form -(CH₂)_q-; and

q is 4, 5 or 6.

Especially preferred within the scope of this invention is a process for the preparation of a compound of the formula (I) in which

n is 1;

m is 1;

A, B, C, E and F are double bonds;

D is a single bond;

R₁, R₂, and R₃ are H;

R₄ is methyl;

R₅ is s-butyl or isopropyl;

R₆ is H;

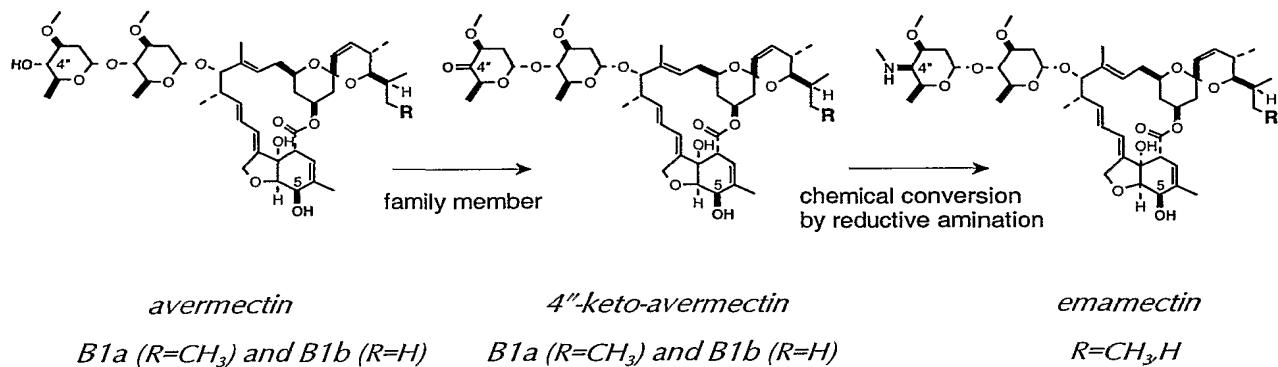
R₇ is OH;

R₈ is methyl

R₉ is H.

Very especially preferred is a process for the preparation of emamectin, more particularly the benzoate salt of emamectin. Emamectin is a mixture of 4"-deoxy-4"-N-methylamino avermectin B_{1a}/B_{1b} and is described in US-P-4,4874,749 and as MK-244 in Journal of Organic Chemistry, Vol. 59 (1994), 7704-7708. Salts of emamectin that are especially valuable agrochemically are described in US-P-5,288,710. Each member of this family of peptides exhibiting an enzymatic activity of a P450 monooxygenases as described hereinbefore is able to oxidize unprotected avermectin regioselectively at position 4", thus opening a new and more economical route for the production of emamectin.

The family members each catalyze the following reaction:



Accordingly, the invention provides a purified nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II) such as avermectin in order to produce a compound of formula (III), but especially 4"-keto-avermectin.

In particular, the invention provides a purified nucleic acid molecule encoding a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. A “nucleic

acid molecule” refers to single-stranded or double-stranded polynucleotides, such as deoxyribonucleic acid (DNA), ribonucleic acid (RNA), or analogs of either DNA or RNA.

The invention also provides a purified polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II) such as avermectin in order to produce a compound of formula (III), but especially 4"-keto-avermectin.

In particular, the invention also provides a purified P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin.

As used herein, by “purified” is meant a nucleic acid molecule or polypeptide (*e.g.*, an enzyme or antibody) that has been separated from components which naturally accompany it. An example of such a nucleotide sequence or segment of interest “purified” from a source, would be nucleotide sequence or segment that is excised or removed from said source by chemical means, *e.g.*, by the use of restriction endonucleases, so that it can be further manipulated, *e.g.*, amplified, for use in the invention, by the methodology of genetic engineering. Such a nucleotide sequence or segment is commonly referred to as “recombinant.” In one specific aspect, the purified nucleic acid molecule may be separated from nucleotide sequences, such as promoter or enhancer sequences, that flank the nucleic acid molecule as it naturally occurs in the chromosome.

In the case of a protein or a polypeptide, the purified protein and polypeptide, respectively, is separated from components, such as other proteins or fragments of cell membrane, that accompany it in the cell. Of course, those of ordinary skill in molecular biology will understand that water, buffers, and other small molecules may additionally be present in a purified nucleic acid molecule or purified protein preparation. A purified nucleic acid molecule or purified polypeptide (*e.g.*, enzyme) of the invention that is at least 95% by weight, or at least 98% by weight, or at least 99% by weight, or 100% by weight free of components which naturally accompany the nucleic acid molecule or polypeptide.

According to the invention, a purified nucleic acid molecule may be generated, for example, by excising the nucleic acid molecule from the chromosome. It may then be ligated into an expression plasmid. Other methods for generating a purified nucleic acid molecule encoding a P450 monooxygenase of the invention are available and include, without limitation, artificial synthesis of the nucleic acid molecule on a nucleic acid synthesizer.

Similarly, a purified P450 monooxygenase of the invention may be generated, for example, by recombinant expression of a nucleic acid molecule encoding the P450 monooxygenase in a cell in which the P450 monooxygenase does not naturally occur. Of course, other methods for obtaining a purified P450 monooxygenase of the invention include, without limitation, artificial synthesis of the P450 monooxygenase on a polypeptide synthesizer and isolation of the P450 monooxygenase from a cell in which it naturally occurs using, *e.g.*, an antibody that specifically binds the P450 monooxygenase.

Biotransformations of secondary alcohols to ketones by *Streptomyces* bacteria are known to be catalyzed by dehydrogenases or oxidases. However, prior to the present discovery of the cytochrome P450 monooxygenase from *Streptomyces tubercidicus* strain R-922 responsible for the regioselective oxidation of avermectin to 4"-keto-avermectin, no experimental data of another cytochrome P450 monooxygenase from *Streptomyces* to oxidize a secondary alcohol to a ketone had been reported.

According to some embodiments of the invention, the nucleic acid molecule and/or the polypeptide encoded by the nucleic acid molecule are isolated from a *Streptomyces* strain. Thus, the nucleic acid molecule (or polypeptide encoded thereby) may be isolated from, without limitation, *Streptomyces tubercidicus*, *Streptomyces lydicus*, *Streptomyces platensis*, *Streptomyces chattanoogensis*, *Streptomyces kasugaensis*, *Streptomyces rimosus*, and *Streptomyces albofaciens*.

As mentioned above and described in more detail below, an entire family of polypeptides exhibiting an enzymatic activity of P450 monooxygenases capable of regioselectively oxidizing avermectin to 4"-keto-avermectin are provided herein. All of these family members are related by at least 60% identity at the amino acid level. A useful nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase comprises or consists essentially of a nucleic acid sequence that is at least 70% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94. In certain embodiments, the nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase

comprises or consists essentially of a nucleic acid sequence that is at least 80% identical; or at least 85% identical; or at least 90% identical; or at least 95% identical; or at least 98% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.

Similarly, the invention provides a purified polypeptide exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin which, in some embodiments, comprises or consists essentially of an amino acid sequence that is at least 60% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95. In certain embodiments, the purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 70% identical; or at least 80% identical; or at least 90% identical; or at least 95% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

In some embodiments, the nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase comprises or consists essentially of the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94. Similarly, the purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase, in some embodiments, comprises or consists essentially of the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

To describe the sequence relationships between two or more nucleic acids or polynucleotides the following terms are used: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

(a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm. Preferred, non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller, 1988; the local homology algorithm of Smith et al. 1981; the homology alignment algorithm of Needleman and Wunsch 1970; the search-for-similarity-method of Pearson and Lipman 1988; the algorithm of Karlin and Altschul, 1990, modified as in Karlin and Altschul, 1993.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Version 8 (available from Genetics Computer Group (GCG), 575 Science Drive, Madison, Wisconsin, USA). Alignments using these programs can be performed using the default parameters. The

CLUSTAL program is well described by Higgins et al. 1988; Higgins et al. 1989; Corpet et al. 1988; Huang et al. 1992; and Pearson et al. 1994. The ALIGN program is based on the algorithm of Myers and Miller, *supra*. The BLAST programs of Altschul et al., 1990, are based on the algorithm of Karlin and Altschul *supra*.

Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., 1990). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached.

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul (1993). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al. 1997. Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships

between molecules. See Altschul et al., *supra*. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g. BLASTN for nucleotide sequences, BLASTX for proteins) can be used. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, 1989). See <http://www.ncbi.nlm.nih.gov>. Alignment may also be performed manually by inspection. For purposes of the present invention, comparison of nucleotide sequences for determination of percent sequence identity to the nucleotide sequences disclosed herein is preferably made using the BlastN program (version 1.4.7 or later) with its default parameters or any equivalent program. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by the preferred program.

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity." Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero

and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e)(i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, or 79%, preferably at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, and most preferably at least 95%, 96%, 97%, 98%, or 99% sequence identity, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 70%, more preferably at least 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions (see below). Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1°C to about 20°C, depending upon the desired degree of stringency as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two

nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e)(ii) The term "substantial identity" in the context of a polypeptide indicates that a polypeptide comprises a sequence with at least 50%, 60%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, or 79%, preferably 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, or even more preferably, 95%, 96%, 97%, 98% or 99%, sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch (1970). An indication that two polypeptide sequences are substantially identical is that one polypeptide is immunologically reactive with antibodies raised against the second polypeptide. Thus, a polypeptide is substantially identical to a second polypeptide, for example, where the two peptides differ only by a conservative substitution.

For sequence comparison, typically one sequence acts as a reference sequence to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

As noted above, another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions. The phrase "hybridizing specifically to" refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions when that sequence is present in a complex mixture (e.g., total cellular) DNA or RNA. "Bind(s) substantially" refers to complementary hybridization between a probe nucleic acid and a target nucleic acid and embraces minor mismatches that can be accommodated by reducing the stringency of the hybridization media to achieve the desired detection of the target nucleic acid sequence.

"Stringent hybridization conditions" and "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments such as Southern and Northern

hybridization are sequence dependent, and are different under different environmental parameters. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl, 1984; $T_m = 81.5^\circ\text{C} + 16.6 (\log M) + 0.41 (\% \text{GC}) - 0.61 (\% \text{ form}) - 500/L$; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. T_m is reduced by about 1°C for each 1% of mismatching; thus, T_m , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with >90% identity are sought, the T_m can be decreased 10°C . Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point I for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4°C lower than the thermal melting point I; moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10°C lower than the thermal melting point I; low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20°C lower than the thermal melting point I. Using the equation, hybridization and wash compositions, and desired T, those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T of less than 45°C (aqueous solution) or 32°C (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, 1993. Generally, highly stringent hybridization and wash conditions are selected to be about 5°C lower than the thermal melting point T_m for the specific sequence at a defined ionic strength and pH.

An example of highly stringent wash conditions is 0.15 M NaCl at 72°C for about 15 minutes. An example of stringent wash conditions is a 0.2X SSC wash at 65°C for 15 minutes (see, Sambrook, *infra*, for a description of SSC buffer). Often, a high stringency wash is preceded by a low stringency wash to remove background probe signal. An example medium

stringency wash for a duplex of, e.g., more than 100 nucleotides, is 1X SSC at 45°C for 15 minutes. An example low stringency wash for a duplex of, e.g., more than 100 nucleotides, is 4-6X SSC at 40°C for 15 minutes. For short probes (e.g., about 10 to 50 nucleotides), stringent conditions typically involve salt concentrations of less than about 1.5 M, more preferably about 0.01 to 1.0 M, Na ion concentration (or other salts) at pH 7.0 to 8.3, and the temperature is typically at least about 30°C and at least about 60°C for long robes (e.g., >50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. In general, a signal to noise ratio of 2X (or higher) than that observed for an unrelated probe in the particular hybridization assay indicates detection of a specific hybridization. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the proteins that they encode are substantially identical. This occurs, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

Very stringent conditions are selected to be equal to the T_m for a particular probe. An example of stringent conditions for hybridization of complementary nucleic acids which have more than 100 complementary residues on a filter in a Southern or Northern blot is 50% formamide, e.g., hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C.

The following are examples of sets of hybridization/wash conditions that may be used to clone orthologous nucleotide sequences that are substantially identical to reference nucleotide sequences of the present invention: a reference nucleotide sequence preferably hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium

dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C.

One non-limiting source of a purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin is the cell-free extract described in the examples below. Another method for purifying a polypeptide exhibiting a P450 monooxygenase activity in accordance with the invention is to attach a tag to the protein, thereby facilitating its purification. Accordingly, the invention provides a purified polypeptide exhibiting an enzymatic activity of a P450 monooxygenase which regioselectively oxidizes avermectin to 4"-keto-avermectin, wherein the polypeptide is covalently bound to a tag. The invention further provides a nucleic acid molecule encoding such a tagged polypeptide.

As used herein, a "tag" is meant a polypeptide or other molecule covalently bound to a polypeptide of the invention, whereby a binding agent (*e.g.*, a polypeptide or molecule) specifically binds the tag. In accordance with the invention, by "specifically binds" is meant that the binding agent (*e.g.*, an antibody or Ni²⁺ resin) recognizes and binds to a particular polypeptide or chemical but does not substantially recognize or bind to other molecules in the sample. In some embodiments, a binding agent that specifically binds a ligand forms an association with that ligand with an affinity of at least 10⁶ M⁻¹, or at least 10⁷ M⁻¹, or at least 10⁸ M⁻¹, or at least 10⁹ M⁻¹ either in water, under physiological conditions, or under conditions which approximate physiological conditions with respect to ionic strength, *e.g.*, 140 mM NaCl, 5 mM MgCl₂. For example, a His tag is specifically bound by nickel (*e.g.*, the Ni²⁺-charged column commercially available as His•Bind® Resin from Novagen Inc, Madison, WI). Likewise, a Myc tag is specifically bound by an antibody that specifically binds Myc.

As described below, a His tag is attached to the purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase by generating a nucleic acid molecule encoding the His-tagged polypeptide, and expressing the polypeptide in *E. coli*. These polypeptides, once expressed by *E. coli*, are readily purified by standard techniques (*e.g.*, using one of the His•Bind® Kits commercially available from Novagen or using the TALON™ Resin (and manufacturer's instructions) commercially available from Clontech Laboratories, Inc., Palo Alto, CA).

Additional tags may be attached to any or all of the polypeptides of the invention to facilitate purification. These tags include, without limitation, the HA-Tag (amino acid sequence: YPYDVPDYA (SEQ ID NO:39)), the Myc-tag (amino acid sequence: EQKLISEEDL (SEQ ID NO:40)), the HSV tag (amino acid sequence: QPELAPEDPED (SEQ ID NO:41)), and the VSV-G-Tag (amino acid sequence: YTIDIEMNRLGK (SEQ ID NO:42)). Covalent attachment (*e.g.*, via a polypeptide bond) of these tags to a polypeptide of the invention allows purification of the tagged polypeptide using, respectively, an anti-HA antibody, an anti-Myc antibody, an anti-HSV antibody, or an anti-VSV-G antibody, all of which are commercially available (for example, from MBL International Corp., Watertown, MA; Novagen Inc.; Research Diagnostics Inc., Flanders, NJ).

The tagged polypeptides of the invention exhibiting a P450 monooxygenase activity may also be tagged by a covalent bond to a chemical, such as biotin, which is specifically bound by streptavidin, and thus may be purified on a streptavidin column. Similarly, the tagged P450 monooxygenases of the invention may be covalently bound (*e.g.*, via a polypeptide bond) to the constant region of an antibody. Such a tagged P450 monooxygenase may be purified, for example, on protein A sepharose.

The tagged P450 monooxygenases of the invention may also be tagged to a GST (glutathione-S-transferase) or the constant region of an immunoglobulin. For example, a nucleic acid molecule of the invention (*e.g.*, comprising SEQ ID NO:1) can be cloned into one of the pGEX plasmids commercially available from Amersham Pharmacia Biotech, Inc. (Piscataway NJ), and the plasmid expressed in *E. coli*. The resulting P450 monooxygenase encoded by the nucleic acid molecule is covalently bound to a GST (glutathione-S-transferase). These GST fusion proteins can be purified on a glutathione agarose column (commercially available from, *e.g.*, Amersham Pharmacia Biotech), and thus purified. Many of the pGEX plasmids enable easy removal of the GST portion from the fusion protein. For example, the pGEX-2T plasmid contains a thrombin recognition site between the inserted nucleic acid molecule of interest and the GST-encoding nucleic acid sequence. Similarly, the pGES-3T plasmid contains a factor Xa site. By treating the fusion protein with the appropriate enzyme, and then separating the GST portion from the P450 monooxygenase of the invention using glutathione agarose (to which the GST specifically binds), the P450 monooxygenase of the invention can be purified.

Yet another method to obtain a purified polypeptide of the invention exhibiting a P450 monooxygenase activity is to use a binding agent that specifically binds to such a polypeptide. Accordingly, the invention provides a binding agent that specifically binds to a P450 monooxygenase of the invention. This binding agent of the invention may be a chemical compound (*e.g.*, a protein), a metal ion, or a small molecule.

In particular embodiments, the binding agent is an antibody. The term “antibody” encompasses, without limitation, polyclonal antibody, monoclonal antibody, antibody fragments (*e.g.*, Fab, Fv, or Fab’ fragments), single chain antibody, chimeric antibody, bi-specific antibody, antibody of any isotype (*e.g.*, IgG, IgA, and IgE), and antibody from any species (*e.g.*, rabbit, mouse, and human).

In one non-limiting example, the binding agent of the invention is a polyclonal antibody. In another non-limiting example, the binding agent of the invention is a monoclonal antibody. Methods for making both monoclonal and polyclonal antibodies are well known (*see, e.g.*, Current Protocols in Immunology, ed. John E. Coligan, John Wiley & Sons, Inc. 1993; Current Protocols in Molecular Biology, eds. Ausubel *et al.*, John Wiley & Sons, Inc. 2000).

The polypeptides described herein exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4”-keto-avermectin belong to a family of novel P450 monooxygenases. Accordingly, the invention also provides a family of P450 monooxygenase polypeptides, wherein each member of the family regioselectively oxidizes avermectin to 4”-keto-avermectin. In some embodiments, each member of the family comprises or consists of an amino acid sequence that is at least 50% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95. In particular embodiments, each member of the family is encoded by a nucleic acid molecule comprising or consisting of a nucleic acid sequence that is at least 66% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.

The present invention, which provides an entire family of P450 monooxygenases, each member of which is able to regioselectively oxidize avermectin to 4"-keto-avermectin, allowed for the generation of an improved P450 monooxygenase, which may not be naturally occurring, but which regioselectively oxidizes avermectin to 4"-keto-avermectin with efficiency and with reduced undesirable side product. For instance, one of the members of the P450 monooxygenase family of the invention, P450_{Ema1} enzyme catalyzes a further oxidation that is not desirable, since the formation of 3"-O-demethyl-4"-keto-avermectin has been detected in the reaction by *Streptomyces tubercidicus* strain R-922 and by *Streptomyces lividans* containing the *ema1* gene. The formation of 3"-O-demethyl-4"-keto-avermectin is brought about by the oxidation of the 3"-O-methyl group, whereby the hydrolytically labile 3"-O-hydroxymethyl group is formed which hydrolyzes to form formaldehyde and the 3"-hydroxyl group.

By providing a family of polypeptides exhibiting an enzymatic activity of P450 monooxygenases that regioselectively oxidize avermectin to 4"-keto-avermectin (see, e.g., Table 3 below), individual members of the family can be subjected to family gene shuffling efforts in order to produce new hybrid genes encoding optimized P450 monooxygenases of the invention. In one non-limiting example, a portion of the *ema1* gene encoding the O₂ binding site of the P450_{Ema1} protein can be swapped with the portion of the *ema2* gene encoding the O₂ binding site of the P450_{Ema2} protein. Such a chimeric *ema1/2* protein is within definition of a P450 monooxygenase of the invention.

Site-directed mutagenesis or directed evolution technologies may also be employed to generate derivatives of the *ema1* gene that encode enzymes with improved properties, including higher overall activity and/or reduced side product formation. One method for deriving such a mutant is to mutate the *Streptomyces* strain itself, in a manner similar to the UV mutation of *Streptomyces tubercidicus* strain R-922 described below.

Additional derivatives may be made by making conservative or non-conservative changes to the amino acid sequence of a P450 monooxygenase. Conservative and non-conservative amino acid substitutions are well known (see, e.g., Stryer, *Biochemistry*, 3rd Ed., W.H. Freeman and Co., NY 1988). Similarly, truncations of a P450 monooxygenase of the invention may be generated by truncating the protein at its N-terminus (e.g., see the *ema1A*

gene described below), at its C-terminus, or truncating (*i.e.*, removing amino acid residues) from the middle of the protein.

Such a mutant, derivative, or truncated P450 monooxygenase is a P450 monooxygenase of the invention as long as the mutant, derivative, or truncated P450 monooxygenase is able to regioselectively oxidize avermectin to 4"-keto-avermectin.

In another aspect, the invention provides a cell genetically engineered to comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. By "genetically engineered" is meant that the nucleic acid molecule is exogenous to the cell into which it is introduced. Introduction of the exogenous nucleic acid molecule into the genetically engineered cell may be accomplished by any means, including, without limitation, transfection, transduction, and transformation.

In certain embodiments, the nucleic acid molecule is positioned for expression in the genetically engineered cell. By "positioned for expression" is meant that the exogenous nucleic acid molecule encoding the polypeptide is linked to a regulatory sequence in such a way as to permit expression of the nucleic acid molecule when introduced into a cell. By "regulatory sequence" is meant nucleic acid sequences, such as initiation signals, polyadenylation (polyA) signals, promoters, and enhancers, which control expression of protein coding sequences with which they are operably linked. By "expression" of a nucleic acid molecule encoding a protein or polypeptide fragment is meant expression of that nucleic acid molecule as protein and/or mRNA.

A genetically engineered cell of the invention may be a prokaryotic cell (*e.g.*, *E. coli*) or a eukaryotic cell (*e.g.*, *Saccharomyces cerevisiae* or mammalian cell (*e.g.*, HeLa)). According to some embodiments of the invention, the genetically engineered cell is a cell wherein the wild-type (*i.e.*, not genetically engineered) cell does not naturally contain the inserted nucleic acid molecule and does not naturally express the protein encoded by the inserted nucleic acid molecule. Accordingly, the cell may be a genetically engineered *Streptomyces* strain, such as a *Streptomyces lividans* or a *Streptomyces avermitilis* strain. Alternatively, the cell may be a genetically engineered *Pseudomonas* strain, such as a *Pseudomonas putida* strain or a *Pseudomonas fluorescens* strain. In another alternative, the cell may be a genetically engineered *Escherichia coli* strain.

Note that in some types of cells genetically engineered to comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin, the actual genetically engineered cell, itself, may not be able to convert avermectin into 4"-keto-avermectin. Rather, the P450 monooxygenase heterogeneously expressed by such a genetically engineered cell may be purified from that cell, where the purified P450 monooxygenase of the invention can be used to regioselectively oxidize avermectin to 4"-keto-avermectin. Thus, the genetically engineered cell of the invention need not, itself, be able to regioselectively convert avermectin to 4"-keto-avermectin; rather, the genetically engineered cell of the invention need only comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin, regardless of whether the polypeptide is active inside that cell.

In addition, a cell (*e.g.*, *E. coli*) genetically engineered to comprise a nucleic acid molecule encoding a polypeptide of the invention which exhibits an enzymatic activity of a P450 monooxygenase may not be able to regioselectively oxidize avermectin to 4"-keto-avermectin, although the P450 monooxygenase purified from the genetically engineered cell is able to regioselectively oxidize avermectin to 4"-keto-avermectin. However, if the same cell were genetically engineered to comprise a polypeptide of the invention which exhibits an enzymatic activity of a P450 monooxygenase, a ferredoxin of the invention, and/or a ferredoxin reductase of the invention, then the P450 monooxygenase together with the ferredoxin and the ferredoxin reductase, all purified from that cell, and in the presence of a reducing agent (*e.g.*, NADH or NADPH), would be able to regioselectively oxidize avermectin to 4"-keto-avermectin. Furthermore the genetically engineered cell comprising a P450 monooxygenase of the invention, a ferredoxin of the invention, and a ferredoxin reductase of the invention, itself, might be able to carry out this oxidation.

Moreover, in a non-limiting example where a cell (*e.g.*, *E. coli*) is genetically engineered to express P450 monooxygenase, a ferredoxin, and a ferredoxin reductase proteins of the invention, all three of these proteins, when purified from the genetically engineered *E. coli*, are together and in the presence of a reducing agent (*e.g.*, NADH or NADPH) would be active and able to regioselectively oxidize avermectin to 4"-keto-avermectin, and so are useful in a method for making emamectin.

In accordance with the present invention, the following material has been deposited with the Agricultural Research Service, Patent Culture Collection (NRRL), 1815 North University Street, Peoria, Illinois 61604, under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure: (1) *Streptomyces lividans* ZX7 (*ema1/fd233-TUA1A*) NRRL Designation No. B-30478; and (2) *Pseudomonas putida* NRRL B-4067 containing plasmid pRK290-*ema1/fd233*, NRRL Designation No.B-30479

In identifying the novel family of polypeptides exhibiting an enzymatic activity of P450 monooxygenases that regioselectively oxidize avermectin to 4"-keto-avermectin, novel ferredoxins and novel ferredoxin reductases were also identified in the same strains of bacteria in which the P450 monooxygenases were found. Accordingly, in a further aspect, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a polypeptide that regioselectively oxidizes avermectin to 4"-keto-avermectin. Similarly, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin reductase, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a polypeptide that regioselectively oxidizes avermectin to 4"-keto-avermectin. The invention also provides a purified protein that exhibits an enzymatic activity of a ferredoxin, as well as a purified protein that exhibits an enzymatic activity of a ferredoxin reductase, wherein the ferredoxin protein and the ferredoxin reductase protein are isolated from a *Streptomyces* strain comprising a polypeptide that regioselectively oxidizes avermectin to 4"-keto-avermectin.

A useful nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin comprises or consists essentially of a nucleic acid sequence that is at least 81% identical to SEQ ID NO:35 or SEQ ID NO:37. Alternatively, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:35 or SEQ ID NO:37. The nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin

may comprise or consist essentially of the nucleic acid sequence of SEQ ID NO:35 or SEQ ID NO:37.

The protein of the invention exhibiting a ferredoxin activity may comprise or consist essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:36 or SEQ ID NO:38. In some embodiments, the nucleic acid molecule comprises or consists essentially an amino acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:36 or SEQ ID NO:38. The ferredoxin of the invention may comprise or consist essentially of the amino acid sequence of SEQ ID NO:36 or SEQ ID NO:38.

A useful nucleic acid molecule comprising a nucleotide sequence encoding a protein of the invention exhibiting a ferredoxin reductase comprises or consists essentially of the nucleic acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:104. In a particular embodiment of the invention, the nucleic acid molecule encoding a ferredoxin reductase of the invention may comprise or consist essentially of the amino acid sequence of SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:104.

The ferredoxin reductase of the invention may comprise or consist essentially of the amino acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, or SEQ ID NO:105. In a particular embodiment of the invention, the ferredoxin reductase of the invention may comprise or consist essentially of the amino acid sequence of SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, or SEQ ID NO:105.

. Methods for purifying ferredoxin and ferredoxin reductase proteins and nucleic acid molecules encoding such ferredoxin and ferredoxin reductase proteins are known in the art and are the same as those described above for purifying P450 monooxygenases of the invention and nucleic acid molecules encoding P450 monooxygenases of the invention.

In one non-limiting example to obtain a purified P450 monooxygenase of the invention with a purified ferredoxin, a *S. lividans* strain (or *P. putida* strain, or any other cell in which the P450 monooxygenase of the invention does not naturally occur) may be genetically engineered to contain a first nucleic acid molecule encoding a P450 monooxygenase of the invention and a second nucleic acid molecule encoding a ferredoxin protein, where both the first and second nucleic acid molecules are positioned for expression in the genetically

engineered cell. The first and the second nucleic acid molecules can be on separate plasmids, or can be on the same plasmid. Thus, the same engineered cell or strain will produce both the P450 monooxygenase of the invention and the ferredoxin protein of the invention.

In a further non-limiting example to obtain a purified P450 monooxygenase of the invention with a purified ferredoxin and with a purified ferredoxin reductase of the invention, a *S. lividans* strain (or *P. putida* strain, or any other cell in which the P450 monooxygenase of the invention does not naturally occur) may be genetically engineered to contain a first nucleic acid molecule encoding a P450 monooxygenase of the invention and a second nucleic acid molecule encoding a ferredoxin protein of the invention and a third nucleic acid molecule encoding a ferredoxin reductase protein of the invention, where all the first and second and third nucleic acid molecules are positioned for expression in the genetically engineered cell. The first and the second and the third nucleic acid molecules may be provided on separate plasmids, or on the same plasmid. Thus, the same engineered cell or strain will produce all the P450 monooxygenase of the invention and the ferredoxin and the ferredoxin reductase proteins of the invention.

As described above for the P450 monooxygenases of the invention, the ferredoxin protein and/or the ferredoxin reductase protein may further comprise a tag. Moreover, the invention contemplates binding agents (*e.g.*, antibodies) that specifically bind to the ferredoxin protein, and binding agents that specifically bind to the ferredoxin reductase proteins of the invention. Methods for generating tagged ferredoxin protein, tagged ferredoxin reductase protein, and binding agents (*e.g.*, antibodies) that specifically bind to ferredoxin or ferredoxin reductase are the same as those as described above for generating tagged P450 monooxygenases of the invention and generating binding agents that specifically bind P450 monooxygenases of the invention.

The invention also provides a method for making emamectin. In this method, a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin is added to a reaction mixture containing avermectin. The reaction mixture is then incubated under conditions that allow the P450 monooxygenase to regioselectively oxidize avermectin to 4"-keto-avermectin. The reaction mixture may further comprise a ferredoxin, such as a ferredoxin of the present invention. In particular embodiments, the reaction mixture further

comprises a ferredoxin reductase such as a ferredoxin of the present invention. The reaction mixture may further comprise a reducing agent, such as NADH or NADPH.

Additionally, the invention provides a method for making 4"-keto-avermectin. The method comprises adding a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin to a reaction mixture comprising avermectin and incubating the reaction mixture under conditions that allow the P450 monooxygenase to regioselectively oxidize avermectin to 4"-keto-avermectin. In some embodiments, the reaction mixture further comprises a ferredoxin, such as a ferredoxin of the present invention. The reaction mixture may also further comprise a ferredoxin reductase such as a ferredoxin of the present invention. In particular embodiments, the reaction mixture further comprises a reducing agent, such as NADH or NADPH.

The invention also provides a formulation for making emamectin comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In some embodiments, the formulation further comprises a ferredoxin, such as a ferredoxin of the present invention. In particular embodiments, the ferredoxin is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. The formulation may further comprise a ferredoxin reductase, such as a ferredoxin reductase of the present invention. In particular embodiments, the ferredoxin reductase is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. . In some embodiments, the formulation further comprises a reducing agent, such as NADH or NADPH.

In addition, the invention provides a formulation for making 4"-keto-avermectin comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In some embodiments, the formulation further comprises a ferredoxin, such as a ferredoxin of the present invention. In particular embodiments, the ferredoxin is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. In some embodiments, the formulation further comprises a ferredoxin reductase, such as a ferredoxin reductase of the present invention. In particular embodiments, the ferredoxin reductase is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. The formulation may further comprise a reducing agent, such as NADH or NADPH.

The following examples are intended to further illustrate certain preferred embodiments of the invention and are not limiting in nature.

EXAMPLE I

Optimized Growth Conditions for *Streptomyces tubercidicus* Strain R-922

In one non-limiting example the fermentation conditions needed to provide a steady supply of cells of *Streptomyces tubercidicus* strain R-922 highly capable of regioselectively oxidizing avermectin to 4"-keto-avermectin were optimized.

First, the following solutions were made. For ISP-2 agar, 4 g of yeast extract (commercially available from Oxoid Ltd, Basingstoke, UK), 4 g of D(+)-glucose, 10 g of bacto malt extract (Difco No. 0186-17-7 (Difco products commercially available from, e.g., Voigt Global Distribution, Kansas City, MO)), and 20 g of agar (Difco No. 0140-01) were dissolved in one liter of demineralized water, and the pH is adjusted to 7.0. The solution was sterilized at 121°C for 20 min., cooled down, and kept at 55°C for the time needed for the immediate preparation of the agar plates.

For PHG medium, 10 g of peptone (Sigma 0521; commercially available from Sigma Chemical Co., St. Louis, MO), 10 g of yeast extract (commercially available from Difco), 10 g of D-(+)-glucose, 2 g of NaCl, 0.15 g of MgSO₄ x 7 H₂O, 1.3 g of NaH₂PO₄ x H₂O, and 4.4 g of K₂HPO₄ were dissolved in 1 liter of demineralized water, and the pH was adjusted to 7.0.

Streptomyces tubercidicus strain R-922 was grown in a Petri dish on ISP-2 agar at 28°C. This culture was used to inoculate four 500 ml shaker flasks with a baffle, each containing 100 ml PHG medium. These pre-cultures were grown on an orbital shaker at 120 rpm at 28°C for 72 hours and then used to inoculate a 10-liter fermenter equipped with a mechanical stirrer and containing 8 liters of PHG medium. This main culture was grown at 28°C with stirring at 500 rpm and with aeration of 1.75 vvm (14 l/min.) and a pressure of 0.7 bar. At the end of the exponential growth, after about 20 hours, the cells were harvested by centrifugation. The yield of wet cells was 70-80 g/l culture.

EXAMPLE II

Whole Cell Biocatalysis Assay

As determined in accordance with the present invention, the following whole cell biocatalysis assay was employed to determine that the activity from *Streptomyces* cells capable of regioselectively oxidizing avermectin to 4"-keto-avermectin is catalyzed by a P450 monooxygenase.

Streptomyces tubercidicus strain R-922 was grown in PHG medium, and *Streptomyces tubercidicus* strain I-1529 was grown in M-17 or PHG medium. PHG medium contains 10 g/l Peptone (Sigma, 0.521), 10 g/l Yeast Extract (Difco, 0127-17-9), 10 g/l D-Glucose, 2 g/l NaCl, 0.15 g/l MgSO₄ x 7 H₂O, 1.3 g/l NaH₂PO₄ x 1 H₂O, and 4.4 g/l K₂HPO₄ at pH 7.0. M-17 medium contains 10 g/l glycerol, 20 g/l Dextrin white, 10 g/l Soytone (Difco 0437-17), 3 g/l Yeast Extract (Difco 0127-17-9), 2 g/l (NH₄)₂SO₄, and 2 g/l CaCO₃ at pH 7.0

To grow the cells, an ISP2 agar plate (not older than 1-2 weeks) was inoculated and incubated for 3-7 days until good growth was achieved. Next, an overgrown agar piece was transferred (with an inoculation loop) to a 250ml Erlenmeyer flask with 1 baffle containing 50 ml PHG medium. This pre-culture is incubated at 28°C and 120 rpm for 2-3 days. Next, 5 ml of the pre-culture were transferred to a 500 ml Erlenmeyer flask with 1 baffle containing 100 ml PHG medium. The main culture was incubated at 28°C and 120 rpm for 2 days. Next, the culture was centrifuged for 10 min. at 8000 rpm on a Beckman Rotor JA-14. The cells were next washed once with 50 mM potassium phosphate buffer, pH 7.0.

To perform the whole cell biocatalysis assay, 500 mg wet cells were placed into a 25 ml Erlenmeyer flask, to which were added 10 ml of 50 mM potassium phosphate buffer, pH 7.0. The cells were stirred with a magnetic stir bar to distribute the cells. Next, 15 µl of a solution of avermectin B1a in isopropanol (30 mg/ml) were added, and the mixture shaken on an orbital shaker at 160 rpm and 28°C. Strain R-922 was reacted for 2 hours, and strain I-1529 was reacted for 30 hours.

To work up the cultures in the whole cell biocatalysis assay, 10 ml methyl-t-butyl-ether was added to an Erlenmeyer flask containing the resting cells and the entire cell mixture was transferred to a 30 ml-centrifuge tube, shaken vigorously, and then centrifuged at 16000 rpm for 10 min. The ether phase was pipetted into a 50 ml pear flask, and evaporated in vacuo by means of a rotary evaporator (\leq 0.1 mbar). The residue was re-dissolved in 1.2 ml acetonitrile

and transferred to an HPLC-sample vial. The conversion of avermectin B1a to 4"-hydroxy-avermectin B1a and 4"-keto-avermectin B1a (also called 4"-oxo-avermectin B1a) and the formation of a side product from the biocatalysis reaction could be observed by HPLC analysis using HPLC protocol I.

For HPLC protocol I, the following parameters were used:

Hardware

Pump:	L-6250 Merck-Hitachi
Autosampler:	AS-2000A Merck-Hitachi
Interface Module:	D-6000 Merck-Hitachi
Channel 1-Detector:	L-7450A UV-Diode Array Merck-Hitachi
Column Oven:	none
Column:	70mm x 4mm
Adsorbent:	Kromasil 100Å-3.5μ-C18

Gradient Mode: Low

Pressure Limit:	5-300bar
Column Temperature	ambient ($\approx 20^\circ C$)
Solvent A:	acetonitrile
Solvent B:	water
Flow:	1.5 ml/min
Detection:	243 nm
Pump Table:	0.0 min 75% A 25% B
<i>linear gradient</i>	7.0 min 100% A 0% B
	9.0 min 100% A 0% B
<i>jump</i>	9.1 min 75% A 25% B
	12.0 min 75% A 25% B
Stop time:	12 min
Sampling Period:	every 200 msec
Retention time table:	<i>time</i> <i>References</i>
	2.12 min 4"-hydroxy- avermectin B1a

3.27 min	avermectin B1a
3.77 min	3"-O-demethyl-4"-keto-avermectin B1a
4.83 min	4"-keto-avermectin B1a

EXAMPLE IIIBiotransformation With Cell-Free Extract From *Streptomyces* Strain R-922

To prepare an active cell-free extract from *Streptomyces tubercidicus* strain R-922 capable of regioselective oxidation of avermectin to 4"-keto-avermectin, the following solutions were made, stored at 4°C, and kept on ice when used.

Solution	Formula
PP-buffer	50 mM K ₂ HPO ₄ /KH ₂ PO ₄ (pH 7.0)
Disruption buffer	50 mM K ₂ HPO ₄ /KH ₂ PO ₄ (pH 7.0), 5 mM benzamidine, 2 mM dithiothreitol, and 0.5 mM Pefabloc (from Roche Diagnostics)
Substrate	10 mg avermectin were dissolved in 1 ml isopropanol

Six grams of wet cells from *Streptomyces* strain R-922 were washed in PP-buffer and then resuspended in 35 ml disruption buffer and disrupted in a French press at 4°C. The resulting suspension was centrifuged for 1 hour at 35000 x g. The supernatant of the cell free extract was collected. One µl substrate was added to 499µl of cleared cell free extract and incubated at 30°C for 1 hour. Then, 1 ml methyl-t-butyl ether was added to the reaction mixture and thoroughly mixed. The mixture was next centrifuged for 2 min. at 14000 rpm, and the methyl-t-butyl ether phase was transferred into a 10 ml flask and evaporated in vacuo

by means of a rotary evaporator. The residue was dissolved in 200 µl acetonitrile and transferred into an HPLC-sample vial.

For HPLC, the HPLC protocol I was used.

When 1 µl substrate was added to 499 µl of cleared cell free extract and incubated at 30°C, no conversion of avermectin to 4"-keto-avermectin was observed by HPLC analysis using HPLC protocol I.

However, the possibility of addition of spinach ferredoxin and spinach ferredoxin reductase and NADPH to the cell free extract to restore the biocatalytic activity was explored (see, generally, D.E. Cane and E.I. Graziani, *J. Amer. Chem. Soc.* **120**:2682, 1998).

Accordingly, the following solutions were made:

Solution	Formula
Substrate	10 mg avermectin were dissolved in 1 ml isopropanol
Ferredoxin	5 mg ferredoxin (from spinach), solution 1-3 mg/ml in Tris/HCl-buffer (from Fluka) or 5 mg ferredoxin (from <i>Clostridium pasteurianum</i>), solution of 1-3 mg/ml in Tris/HCl-buffer (from Fluka) or 5 mg ferredoxin (from <i>Porphyra umbilicalis</i>), solution of 1-3 mg/ml in Tris/HCl-buffer (from Fluka)
Ferredoxin Reductase	1 mg freeze-dried ferredoxin reductase (from spinach), solution of 3.9 U/mg in 1 ml H ₂ O (from Sigma)
NADPH	100 mM NADPH in H ₂ O (from Roche Diagnostics)

The substrate solution was stored at 4°C, the other solutions were stored at -20°C, and kept on ice when used.

Thus, to 475 µl of cleared cell free extract the following solutions were added: 10 µl ferredoxin, 10 µl ferredoxin reductase and 1 µl substrate. After the addition of substrate to the cells, the mixture was immediately and thoroughly mixed and aerated. Then, 5 µl of NADPH were added and the mixture incubated at 30°C for 30 min. Then, 1 ml methyl-t-butyl ether was added to the reaction mixture and thoroughly mixed. The mixture was next centrifuged for 2 min. at 14000 rpm, and the methyl-t-butyl ether phase was transferred into a 10 ml flask

and evaporated in vacuo by means of a rotary evaporator. The residue was dissolved in 200 µl acetonitrile and transferred into an HPLC-sample vial, and HPLC analysis performed using HPLC protocol I.

Formation of 4"-keto-avermectin was observable by HPLC analysis. Thus, addition of spinach ferredoxin and spinach ferredoxin reductase and NADPH to the cell free extract restored the biocatalytic activity.

Upon injection of a 30 µl sample, a peak appeared at 4.83 min., indicating the presence of 4"-keto-avermectin B1a. A mass of 870 D could be assigned to this peak by HPLC-mass spectrometry which corresponds to the molecular weight of 4"-keto-avermectin B1a.

Note that when analyzing product formation by HPLC and HPLC-mass spectrometry, in addition to the 4"-keto-avermectin, the corresponding ketohydrate 4"-hydroxy-avermectin was also found giving a peak at 2.12 min. This finding indicated that the P450 monooxygenase converts avermectin by hydroxylation to 4"-hydroxy-avermectin, from which 4"-keto-avermectin is formed by dehydration. Interestingly, when the spinach ferredoxin was replaced by ferredoxin from the bacterium *Clostridium pasteurianum* or from the red alga *Porphyra umbilicalis*, the biocatalytic conversion of avermectin to 4"-keto-avermectin still took place, indicating that the enzyme does not depend on a specific ferredoxin for receiving reduction equivalents.

EXAMPLE IV

Isolation of a Mutant *Streptomyces* Strain R-922 With Enhanced Activity

To obtain strains of *Streptomyces* strain R-922 that have an enhanced ability to regioselectively oxidize avermectin to 4"-keto-avermectin, UV mutants were generated. To do this, spores of *Streptomyces* strain R-922 were collected and stored in 15% glycerol at -20°C. This stock solution contained 2×10^9 spores.

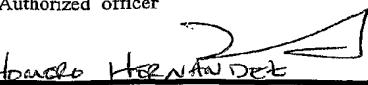
The spore stock solution was next diluted and transferred to petri plates containing 10ml of sterile water, and the suspension was exposed to UV light in a Stratalinker UV crosslinker 2400 (commercially available from Stratagene, La Jolla, CA). The Stratalinker UV crosslinker uses a 254-nm light source and the amount of energy used to irradiate a sample can be set in the "energy mode."

Applicant's or agent's file reference	PB/5-60016A	International application No.
		PCT/EP 02/05363

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B. IDENTIFICATION OF DEPOSIT	
Name of depositary institution Agricultural Research Service, Patent Culture Collection (NRRL)	
Address of depositary institution (<i>including postal code and country</i>) 1815 North University Street Peoria Illinois 61604 USA	
Date of deposit May 08, 2001	Accession Number NRRL B-30479
C. ADDITIONAL INDICATIONS (<i>leave blank if not applicable</i>) This information is continued on an additional sheet <input type="checkbox"/>	
Pseudomonas putida NRRL B-4067 containing plasmid pRK290-ema1/fd233	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (<i>if the indications are not for all designated States</i>)	
E. SEPARATE FURNISHING OF INDICATIONS (<i>leave blank if not applicable</i>)	
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Address of depositary institution (<i>including postal code and country</i>) 1815 North University Street Peoria Illinois 61604 USA	
Date of deposit May 08, 2001	Accession Number NRRL B-30478
C. ADDITIONAL INDICATIONS (<i>leave blank if not applicable</i>)	
Streptomyces lividans ZX7 (ema1/fd233-TUA1A)	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (<i>if the indications are not for all designated States</i>)	
E. SEPARATE FURNISHING OF INDICATIONS (<i>leave blank if not applicable</i>)	
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Through experimentation, it was determined that an exposure of 8000 microjoules of UV irradiation (254 nm) was required to kill 99.9% of the spores. This level of UV exposure was used in the mutagenesis.

Surviving UV-mutagenized spores were plated, cultured, and transferred to minimal media. Approximately 0.3-0.4% of the viable spores were determined to be auxotrophic, indicating a good level of mutagenesis in the population.

The mutagenized clones were screened for activity in the whole cell biocatalysis assay described in Example II. As shown in an HPLC chromatogram, one mutant ("R-922 UV mutant") showed a two to three fold increase in an ability to regioselectively oxidize avermectin to 4"-keto-avermectin as compared to wild-type strain R-922. Although the gene encoding the P450 monooxygenase responsible for the regioselectively oxidation activity, *ema1*, is not mutated in the R-922 UV mutant, this mutant nonetheless provides an excellent source for a cell-free extract containing *ema1* protein.

EXAMPLE V

Isolation of the P450 Monooxygenase from *Streptomyces* Strain R-922

To enrich the P450 enzyme, 35 ml of active cell free extract were filtered through a 45 µm filter and fractionated by anion exchange chromatography. Anion exchange chromatography conditions were as follows:

FPLC instrument: Äkta prime (from Pharmacia Biotech)

FPLC-column: HiTrapTMQ (5 ml) stacked onto Resource® Q (6 ml) (from Pharmacia Biotech)

eluents buffer A: 25 mM Tris/HCl (pH 7.5)

 buffer B: 25 mM Tris/HCl (pH 7.5) containing 1 M KCl

temperature eluent bottles and fractions in ice bath,

flow 3 ml/min

detection UV 280nm

Pump table: 0.0 min 100% A 0% B

linear gradient to 2.0 min 90% A 10% B

 5.0 min 90% A 10% B

<i>linear gradient to</i> 30.0 min	50% A	50% B
<i>linear gradient to</i> 40.0 min	0% A	100% B
50.0min	0% A	100% B

Enzyme activity eluted with 35%-40% buffer B. The active fractions were pooled and concentrated by centrifugal filtration through Biomax™ filters with an exclusion limit of 5kD (commercially available from Millipore Corp., Bedford, MA) at 5000 rpm and then rediluted in disruption buffer containing 20% glycerol to a volume of 5 ml containing 3-10 mg/ml protein. This enriched enzyme solution contained at least 25% of the original enzyme activity.

The enzyme was further purified by size exclusion chromatography. Size exclusion chromatography conditions were as follows:

FPLC instrument:	Äkta prime (from Pharmacia Biotech)
FPLC-column:	HiLoad 26/60 Superdex® 200 prep grade (from Pharmacia Biotech)
sample:	3-5 ml enriched enzyme solution from the anion chromatography step
sample preparation:	filtered through 45 µm filter
eluent buffer:	PP-buffer (pH 7.0) + 0.1 M KCl
temperature:	4°C
flow:	2 ml/min
detection:	UV 280nm

Enzyme activity eluted between 205-235 ml eluent buffer. The active fractions were pooled, concentrated by centrifugal filtration through Biomax™ filters with an exclusion limit of 5 kD (from Millipore) at 5000 rpm, and rediluted in disruption buffer containing 20% glycerol to form a solution of 0.5-1 ml containing 2-5mg/ml protein. This enriched enzyme solution contained 10% of the original enzyme activity. This enzyme preparation, when checked for purity by SDS page, (*see*, generally, Laemmli, U.K., *Nature* 227:680-685, 1970 and Current Protocols in Molecular Biology, *supra*) and stained with Coomassie blue, showed one dominant protein band with a molecular weight of 45-50 kD, according to reference proteins of known molecular weight.

EXAMPLE VI

Attempted Isolation of P450 Monooxygenase Genes
From *Streptomyces* Strains R-922 and I-1529

Based on results described above that suggested the enzyme from strain R-922 that is responsible for the regiospecific oxidation of avermectin to 4"-keto-avermectin is a P450 monooxygenase, a direct PCR-based approach to clone P450 monooxygenase genes from this strain was initiated (*see, generally, Hyun et al., J. Microbiol. Biotechnol. 8(3):295-299, 1998.*) This approach is based on the fact that all P450 monooxygenase enzymes contain highly conserved oxygen-binding and heme-binding domains that are also conserved at the nucleotide level. PCR primers were designed to prime to these conserved domains and to amplify the DNA fragment from P450 genes using R-922 or I-1529 genomic DNA as a template. The PCR primers used are shown in Table 1.

Table 1

O ₂ -Binding Domain Primers (5' to 3')*	Degeneracy	SEQ ID NOS
I A G H E T T ATC GCS GGS CAC GAG ACS AC	8	43 44
V A G H E T T GTS GCS GGS CAC GAG ACS AC	16	45 46
L A G H E T T CTS GCS GGS CAC GAG ACS AC	16	47 48
L L L I A G H E T TS CTS CTS ATC GCS GGS CAC GAG AC ^k	32	49 50
<hr/>		
Heme-Binding Domain Primers (3' to 5')*		
H Q C L G Q N L A GTG GTC ACG GAS CCS TGC TTG GAS CG ^k	8	51 52
F G H G V H Q C AAG CCS GTG CCS CAS GTG GTC ACG	8	53 54
F G F G V H Q C AAG GCS AAG CCS CAS GTG GTC ACG	8	55 56
F G H G I H Q C AAG CCS GTG CCS TAG GTG GTC ACG	4	57 58

F G H G V H F C		59
AAG CCS GTG CCS CAS GTG AAG ACG	8	60

* The amino acid sequence is shown on the top line and the corresponding nucleotide sequence is shown below on the second line; S=G or C.

& This primer was described by Hyun *et al.*, *supra*

PCR amplification using any of the primers specific to nucleotide sequences encoding the O₂-binding domain with any of the primers specific to the nucleotide sequences encoding the heme-binding domain and genomic DNA from *Streptomyces* strains R-922 or I-1529 resulted in the amplification of an approximately 350 bp DNA fragment. This is exactly the size that would be expected from this PCR amplification due to the approximately 350 bp separation in P450 genes of the gene segments encoding the O₂-binding and heme-binding sites.

The 350 bp PCR fragments were cloned into the pCR2.1-TOPO TA cloning plasmid (commercially available Invitrogen, Carlsbad, CA) and transformed into *E. coli* strain TOP10 (Invitrogen, Carlsbad, CA). Approximately 150 individual clones from strains R-922 and I-1529 were sequenced to determine how many unique P450 gene fragments were represented. Analysis of the sequences revealed that they included 8 unique P450 gene fragments from strain R-922 and 7 unique fragments from I-1529.

Blast analysis (alignment of the deduced amino acid sequences of P450 gene-specific PCR fragments derived from *Streptomyces tubercidicus* strain R-922 and *Streptomyces* strain I-1529, respectively, and the P450 monooxygenase from *S. thermotolerans* that is involved in the synthesis of carbomycin (Stol-ORFA) (GenBank Accession No. D30759) by the program Pretty from the University of Wisconsin Package version 10.1 (Altschul *et al.*, *Nucl. Acids Res.* **25**:3389-3402). demonstrated that all of the unique P450 gene fragments from both the R-922 and I-1529 strains were derived from P450 genes and encoded the region between the O₂-binding and heme-binding domains.

Next, in order to clone the full-length genes from which the PCR fragments were derived, the DNA fragments cloned by PCR were used as hybridization probes to gene libraries containing genomic DNA from strains R-922 and I-1529. To do this, genomic DNA from the R-922 and I-1529 strains was partially digested with Sau3A I, dephosphorylated with

calf intestinal alkaline phosphatase (CIP) and ligated into the cosmid pPEH215, a modified version of SuperCos 1 (commercially available from Stratagene, La Jolla, CA). Ligation products were packaged using the Gigapack III XL packaging extract and transfected into *E. coli* XL1 Blue MR host cells. Twelve cosmids that strongly hybridized to the PCR-generated P450 gene fragments were identified from the R-922 library, from which three unique P-450 genes were subcloned and sequenced. The hybridizations were performed at high stringency conditions according to the protocol of Church and Gilbert (Church and Gilbert, *Proc. Natl. Acad. Sci. USA* **81**:1991-1995, 1984). In brief, these high stringency conditions include Hybrid Buffer containing 500 mM Na-phosphate, 1 mM EDTA, 7% SDS, 1% BSA; Wash Buffer 1 containing 40 mM Na-phosphate, 1 mM EDTA, 5% SDS, 0.5% BSA; and Wash Buffer 2 containing 40 mM Na-phosphate, 1 mM EDTA, 1% SDS (Note that other high stringency hybridizations conditions are described, for example, in Current Protocols in Molecular Biology, *supra*.) Nineteen strongly hybridizing cosmids were identified from the I-1529 library, and from these, four unique P-450 genes were subcloned and sequenced.

In yet a further approach to isolate diverse P450 monooxygenase genes from strains R-922 and I-1529, a known P450 gene from another bacterium was used as a hybridization probe to identify cosmid clones containing homologous P450 genes from strains R-922 and I-1529. The *epoF* P450 gene from *Sorangium cellulosum* strain So ce90 that is involved in the synthesis of epothilones (Molnar *et al.*, *Chem Biol.* **7**(2):97-109, 2000) was used as a probe in this effort. Using the *epoF* P450 gene probe, one cosmid was identified from strain R-922 (clone LC), and three were identified from strain I-1529 (clones LA, LB, and EA). In each case, the homologous gene fragment was subcloned and sequenced, and found to code for P450 monooxygenase enzymes.

However, a comparison of the 17 polypeptide sequences identified in Example VII (below) failed to match any of these cloned genes. Two of the polypeptide sequences (namely, LVKDDPALLPR and AVHELMR) mapped to the region between the O₂ and heme binding domains, and so these should have identified any of the partial gene fragments derived by the PCR approach. Thus, the standard approaches based on the known PCR technique of Hyun *et al.*, *supra*, and using known P450 genes as hybridization probes failed to identify the gene that encodes the specific P450 monooxygenase responsible for the regioselective

oxidation of avermectin. Accordingly, it was determined that additional experimentation was required to isolate the gene encoding the P450 monooxygenase of the invention.

EXAMPLE VII

Partial Sequencing of the P450 Monooxygenase from *Streptomyces* Strain R-922

Partial amino acid sequencing of the P450 monooxygenase from *Streptomyces* strain R-922 was carried out by the Friedrich Miescher Institute, Basel Switzerland. The protein of the dominant band on the SDS page was tryptically digested and the formed peptides separated and sequenced by mass spectrometry and Edman degradation (*see, generally, Zerbe-Burkhardt et al., J. Biol. Chem. 273:6508, 1998*). The sequence of the following 17 peptides were found:

<u>Sequence</u>	<u>Sequence I.D. No.</u>
HPGEPNVMDPALITDPFTGYGALR	(SEQ ID NO:61)
FVNNPASPSSLNYAPEDNPLTR	(SEQ ID NO:62)
LLTHYPDISLGIAPEHLER	(SEQ ID NO:63)
VYLLGSILNYDAPDHTR	(SEQ ID NO:64)
TWGADLISMDPDR	(SEQ ID NO:65)
EALTDDLLSELIR	(SEQ ID NO:66)
FMDDSPVWLVTR	(SEQ ID NO:67)
LMEMLGLPEHLR	(SEQ ID NO:68)
VEQIADALLAR	(SEQ ID NO:69)
LVKDDPALLPR	(SEQ ID NO:70)
DDPALLPR	(SEQ ID NO:71)
TPLPGNWWR	(SEQ ID NO:72)
LNSLPVR	(SEQ ID NO:73)
ITDLRPR	(SEQ ID NO:74)
EQGPVVR	(SEQ ID NO:75)
AVHELMR	(SEQ ID NO:76)

AFTAR (SEQ ID NO:77)
FEEVR (SEQ ID NO:78)

Alignment of these peptides to a selection of actinomycete P450 monooxygenase sequences indicated that all the peptides were fragments of a single P450 mono-oxygenase.

EXAMPLE VIII

Cloning the P450 Monooxygenase Gene from Strain R-922 that Encodes the Enzyme Responsible for the Oxidation of Avermectin to 4"-Keto-Avermectin

PCR primers were designed by reverse translation from the amino acid sequences of several of the peptides derived from the P450 enzyme of strain R-922 (*see* Example VII and Table 2 below). Each of five forward primers (2aF, 2bF, 3F, 1F, and 7F) was paired with one reverse primer (5R) in PCR reactions with R-922 genomic DNA as a template. In each reaction, a DNA fragment of the expected size was produced.

Table 2

Primer	Primer sequence and the amino acid sequence to which they were designed*	Degeneracy	Expected size (bp) **	SEQ ID NO:
2aF	P G E D N V M 5'-CCS GGS GAR CCS AAY GTS ATG-3'	64	600 80	79
2bF	A L I T D P F 5'-GCS CTS ATY ACS GAC CCS TTC-3'	32	580 82	81
3F	F M D D S P V W 5'-TTC ATG GAC GAC WSS CCS GTS TGG-3'	32	549 84	83
1F	L N Y D A P D H 5'-CTS AAY TAY GAC GAC GCS CCS GAC CAC-3'	32	350 86	85
7F	V E Q I A D A L 5'-GTS GAR CAG ATY GCS GAC GCS CTS-3'	32	300 88	87
5R	D L I S M D P D 3'-CTG GAS TAR WSS TAC CTG GGS CTG-5'	64	---	89 90

* Ambiguity codes: Y=C or T; R=A or G; S=C or G; W=A or T

** Expected size of PCR product when the primer is when paired with primer 5R

The 580 and 600 bp PCR fragments generated by using primers (2bF and 5R) and (2aF and 5R), respectively, were cloned into the pCR-Blunt II-TOPO cloning plasmid (commercially available from Invitrogen, Carlsbad, CA) and transformed into *E. coli* strain TOP10 (Invitrogen, Carlsbad, CA). The inserted DNA fragments were then sequenced. Examination of the sequences revealed that the 600 and 580 bp fragments were identical in the 580 bp of sequence that they have in common. Also, there was a perfect match between the deduced amino acid sequence (SEQ ID NO:2) derived from the nucleotide sequence of the 600 bp and 580 bp fragments and the amino acid sequences of peptides isolated from the purified P450_{Ema1} enzyme that aligned in this region of the isolated gene. This result strongly suggested that the gene fragments isolated in these clones are derived from the gene that encodes the P450_{Ema1} enzyme that is responsible for the oxidation of avermectin to 4"-keto-avermectin.

The 600 bp PCR fragment produced using primers 2aF (SEQ ID No:80) and 5R (SEQ ID No:90) was used as a hybridization probe to a cosmid library of genomic DNA isolated from strain R-922 (cosmid library described in Example VI). Two cosmids named pPEH249 and pPEH250 were identified that hybridized strongly with the probe. The portion of each cosmid encoding the P450 enzyme was sequenced and the sequences were found to be identical between the two cosmids. The complete coding sequence of the *ema1* gene was identified (SEQ ID NO:1). The amino acid sequence of all polypeptide fragments from P450_{Ema1} matched perfectly with the deduced amino acid sequence from the *ema1* gene. Comparison of the deduced amino acid sequence of the protein encoded by the *ema1* gene using BLASTP (Altschul *et al.*, *supra*) determined that the closest match in the databases is to a P450 monooxygenase from *S. thermotolerans* that has a role in the biosynthesis of carbomycin (Arisawa *et al.*, *Biosci. Biotech. Biochem.* **59**(4):582-588, 1995) and whose identity with *ema1* is only 49% (Identities = 202/409 (49%), Positives = 271/409 (65%), Gaps = 2/409 (0%)). In the Blast analysis, the following settings were employed:

```

BLASTP 2.0.10
Lambda      K      H
 0.322     0.140   0.428
Gapped
Lambda      K      H
 0.270     0.0470  0.230
Matrix: BLOSUM62
Gap Penalties: Existence: 11, Extension: 1
Number of Hits to DB: 375001765
Number of Sequences: 1271323
Number of extensions: 16451653
Number of successful extensions: 46738
Number of sequences better than 10.0: 2211
Number of HSP's better than 10.0 without gapping: 628
Number of HSP's successfully gapped in prelim test: 1583
Number of HSP's that attempted gapping in prelim test: 43251
Number of HSP's gapped (non-prelim): 2577
length of query: 430
length of database: 409,691,007
effective HSP length: 55
effective length of query: 375
effective length of database: 339,768,242
effective search space: 127413090750
effective search space used: 127413090750

```

A similar comparison of the nucleotide sequences of these two genes demonstrated that they are 65% identical at the nucleotide level. These results demonstrate that P450_{Ema1} is a new enzyme.

EXAMPLE IX

Heterologous Expression of the *ema1* Gene in *Streptomyces lividans* Strain ZX7

The coding sequence of the *ema1* gene was fused to the thiostrepton-inducible promoter (*tipA*) (Murakami *et al.*, *J. Bacteriol.* **171**:1459-1466, 1989). The *tipA* promoter was derived from plasmid pSIT151 (Herron and Evans, *FEMS Microbiology Letters* **171**:215-221, 1999).

The fusion of the *tipA* promoter and the *ema1* coding sequence was achieved by first amplifying the *ema1* coding sequence with the following primers to introduce a PacI cloning site at the 5' end and a PmeI compatible end on the 3' end.

Forward Primer: The underlined sequence is a PacI recognition sequence; the sequence in bold-face type is the start of the coding sequence of *ema1*.

5'-AGATTAATTAA**TGT**CGGAATTAATGAACTGTCCGTT-3' (SEQ ID NO:91)

Reverse Primer: The underlined sequence is half of a PmeI recognition sequence; the bold-face type sequence is the reverse complement of the *ema1* translation stop codon followed by the 3' end of the *ema1* coding sequence.

5'-AAACTCACCCAACCGCACCGGAGCGAGTTC-3" (SEQ ID NO:92)

The PacI-digested PCR fragment containing the *ema1* coding sequence was cloned into plasmid pTBBKA (*see Figure 1*) that was restricted (*i.e.*, digested) with PacI and PmeI, and the ligated plasmid transformed into *E. coli*. Four clones were sequenced. Three of the four contained the complete and correct *ema1* coding sequence. The fourth *ema1* gene clone contained a truncated version of the *ema1* gene. The full-length *ema1* gene encodes a protein that begins with the amino acid sequence MSEL MNS (SEQ ID NO:93). The truncated gene encodes a protein that lacks the first 4 amino acids and begins with the second methionine residue. This gene has been named *ema1A*. The nucleotide and amino acid sequence of *ema1A* are provided as SEQ ID NO:33 and SEQ ID NO:34, respectively. The *ema1* and *ema1A* genes in these plasmids, pTBBKA-*ema1* and pTBBKA-*ema1A*, are in the correct juxtaposition with the *tipA* promoter to cause expression of the genes from this promoter.

Plasmid pTBBKA contains a gene from the *Streptomyces* insertion element IS117 that encodes an integrase that catalyzes site-specific integration of the plasmid into the chromosome of *Streptomyces* species (Henderson *et al.*, *Mol. Microbiol.* **3**:1307-1318, 1989 and Lydiate *et al.*, *Mol. Gen. Genet.* **203**:79-88, 1986). Since plasmid pTBBKA has only an *E. coli* replication origin and contains a mobilization site, it can be transferred from *E. coli* to *Streptomyces* strains by conjugation where it will not replicate. However, it is able to integrate into the chromosome due to the IS117 integrase and *Streptomyces* clones containing chromosomal integrations can be selected by resistance to kanamycin due to the plasmid-borne kanamycin resistance gene.

The *ema1* coding sequence was also cloned into other plasmids that are either replicative in *Streptomyces* or, like pTBBKA, integrate into the chromosome upon introduction into a *Streptomyces* host. For example, *ema1* was cloned into plasmid pEAA, which is similar to plasmid pTBBKA but the KpnI/PacI fragment containing the *tipA* promoter was replaced with the *ermE* gene promoter (Schmitt-John and Engels, *Appl*

Microbiol Biotechnol. **36**(4):493-498, 1992). In addition, pEAA does not contain the kanamycin resistance gene. The *ema1* gene was cloned into pEAA as a PacI/PmeI fragment to create plasmid pEAA-*ema1* in which the *ema1* gene is expressed from the constitutive *ermE* promoter.

Plasmid pTUA1A is a *Streptomyces-E.coli* shuttle plasmid (*see* Figure 2) that contains the *tipA* promoter. The *ema1* gene was also cloned into the PacI/PmeI site in plasmid pTUA1A to create plasmid pTUA-*ema1*.

The *ema1A* gene fragment was also ligated as a PacI/PmeI fragment into plasmids pTUA1A, and pEAA in the same way as the *ema1* gene fragment to create plasmids pTUA-*ema1A*, and pEAA-*ema1A*, respectively.

The pTBBKA, pTUA1A, and pEAA based plasmids containing the *ema1* or *ema1A* genes were introduced into *S. lividans* ZX7 and in each case transformants were obtained and verified (*S. lividans* strains ZX7::pTBBKA-*ema1* or *ema1A*, ZX7 (pTUA-*ema1* or -*ema1A*), and ZX7::pEAA-*ema1* or -*ema1A*, respectively).

Wild-type *Streptomyces lividans* strain ZX7 was tested and found to be incapable of the oxidation of avermectin to 4"-keto-avermectin. Transformed *S. lividans* strains ZX7::pTBBKA-*ema1*, ZX7::pTBBKA-*ema1A*, ZX7 (pTUA-*ema1*), ZX7 (pTUA-*ema1A*), ZX7::pEAA-*ema1*, and ZX7::pEAA-*ema1A* were each tested for the ability to oxidize avermectin to 4"-keto-avermectin using resting cells. To do this, the whole cell biocatalysis assay described above (including analysis method) was performed. Note that for the whole cell biocatalysis assay, transformed *Streptomyces lividans*, like strain R-922, was grown in PHG medium and, again like strain R-922, had a reaction time of 16 hours (*i.e.*, during which time the 500 mg transformed *Streptomyces lividans* wet cells in 10 ml of 50 mM potassium phosphate buffer, pH 7.0, were shaken at 160 rpm at 28°C in the presence of 15 µl of a solution of avermectin in isopropanol (30 mg/ml)).

In the presence of the inducer, thiostrepton (5 µg/ml), the *ema1*- or *ema1A*-containing strains ZX7::pTBBKA-*ema1*, ZX7::pTBBKA-*ema1A*, ZX7 (pTUA-*ema1*), ZX7 (pTUA-*ema1A*) were found to oxidize avermectin to 4"-keto-avermectin as evidenced by the appearance of the oxidized 4"-keto-avermectin compound (*see* Table 3).

Table 3

<u>Beispiel 1: Strain</u>	% Conversion of Avermectin	
	2 hour	16 hour
<i>Streptomyces lividans ZX7 + Plasmid</i> ¹		
None	0	0
pTBBKA- <i>ema1A</i>	0.5 ±0.059	1.17 ±0.112
pTBBKA- <i>ema1</i>	0.21 ±0.0.356	0.65 ±0.079
pTUA- <i>ema1</i>	20.96 ±1.044	42.0 ±2.5
pEAA- <i>ema1</i>	3.0 ±0.232	24.1 ±0.358
pTBBKA- <i>ema2</i>	4.79 ±0.096	9.57 ±0.423
pTUA- <i>ema2</i>	0.77 ±0.138	2.05 ±0.537
pEAA- <i>ema2</i>	0.0	1.73 ±3.00
pTBBKA- <i>ema1/fd233</i>	8.89 ±0.720	30.99 ±0.880
pTUA- <i>ema1/fd233</i>	23.29 ±0.854	61.2 ±3.548
pEAA- <i>ema1/fd233</i>	8.26 ±0.845	10.66 ±0.858
pTUA- <i>ema2/fd233</i>	1.85 ±0.861	6.40 ±1.918
<i>Pseudomonas putida S12 + Plasmid</i>		
None		0
pRK- <i>ema1</i>	ND ²	18
pRK- <i>ema1/fd233</i>	ND	32

¹pTBBKA= IS117 integrase, tipA promoter; pTUA= replicative plasmid, tipA promoter;

pEAA= IS117 integrase, ermE promoter

²Not Determined

These results conclusively demonstrate that the P450_{Ema1} enzyme encoded by the *ema1* gene is responsible for the oxidation of avermectin to 4"-keto-avermectin in *S. tubercidicus* strain R-922. Furthermore, the data demonstrates that the *ema1A* gene that is 4 amino acids shorter on the N-terminus than the native *ema1* gene also encodes an active P450_{Ema1} enzyme. As can be demonstrated by HPLC analysis, oxidation of avermectin to 4"-keto-avermectin by *S.*

lividans strain ZX7::pTBBKA-*ema1* following induction of *ema1* expression with 0, 0,5, or 5.0 µg/ml thiostrepton. is variable depending upon the amount of thiostrepton used to induce expression of *ema1*. Note that *S. lividans* strains ZX7::pEAA-*ema1* and ZX7::pEAA-*ema1A* (see Table 3) demonstrated this oxidation activity in the absence of thiostrepton since in these strains the *ema1* or *ema1A* genes are expressed from the *ermE* promoter that does not require induction.

EXAMPLE X

Isolation of an *ema1*-Homologous Gene From *Streptomyces tubercidicus* Strain I-1529

Streptomyces tubercidicus strain I-1529 was also found to be active in biocatalysis of avermectin to form the 4"-keto-avermectin derivative. The cosmid library from strain I-1529, described in Example VI, was probed at the high stringency conditions of Church and Gilbert (Church and Gilbert, *Proc. Natl. Acad. Sci. USA* 81:1991-1995, 1984) with the 600 bp *ema1* PCR fragment produced using primers 2aF (SEQ ID No:80) and 5R (SEQ ID No:90) described previously to identify clones containing the *ema1* homolog from strain I-1529. Three strongly hybridizing cosmids were identified. The P450 gene regions in two of the cosmids, pPEH252 and pPEH253, were sequenced and found to be identical. Analysis of the DNA sequence revealed the presence of a gene with high homology to the *ema1* gene of strain R-922. A comparison of the deduced amino acid sequence of Ema2 (*i.e.*, P450_{Ema2}), Ema1 (*i.e.*, P450_{Ema1}), and a P450 monooxygenase from *Streptomyces thermotolerans* that is involved in the biosynthesis of carbomycin (Carb-450) (GenBank Accession No. D30759). demonstrated that all of the unique P450 gene fragments from both the R-922 and I-1529 strains were derived from P450 genes and encoded the region between the O₂-binding and heme-binding domains.

The gene from *Streptomyces tubercidicus* strain I-1529, named *ema2*, encodes an enzyme with 90% identity at the amino acid level and 90.6% identity at the nucleotide level to the P450_{Ema1} enzyme. The nucleotide sequence of the *ema2* gene and the deduced amino acid sequence of P450_{Ema2} are provided in SEQ ID NO:3 and SEQ ID NO:4, respectively.

The *ema2* coding sequence was cloned in the same manner as the *ema1* and *ema1A* genes into plasmids pTBBKA, pTUA1A, and pEAA such that the coding sequence was

functionally fused to the *tipA* or *ermE** promoter in these plasmids. The resulting plasmids, pTBBKA-*ema2*, pTUA-*ema2*, and pEAA-*ema2* were transferred from *E. coli* to *S. lividans* ZX7 by conjugation to create strains ZX7::TBBKA-*ema2* and ZX7 (pTUA-*ema2*), and ZX7::pEAA-*ema2* containing the *ema2* gene integrated into the chromosome or maintained on a plasmid.

Strains ZX7::TBBKA-*ema2*, ZX7 (pTUA-*ema2*), and ZX7::pEAA-*ema2* were next tested for the ability to oxidize avermectin to 4"-keto-avermectin. The *ema2* gene was also shown to provide biocatalysis activity, although at a lower level compared to the *ema1* gene (see Table 3).

These results demonstrate that the *ema2* gene from *S. tubercidicus* strain I-1529 also encodes a P450 enzyme (P450_{Ema2}) capable of oxidizing avermectin to 4"-keto-avermectin.

EXAMPLE XI

Characterization of *ema1* Homologs From Other Biocatalysis Strains

Seventeen *Streptomyces* sp. strains, including strains R-922 and I-1529, were identified that are capable of catalyzing the regiospecific oxidation of the 4"-carbinol of avermectin to a ketone. Next, the isolation and characterization of the genes encoding the biocatalysis enzyme from all of these strains was accomplished.

To do this, genomic DNA was isolated from the strains and was evaluated by restriction with several restriction endonucleases and Southern hybridization with the *ema1* gene. A specific restriction endonuclease was identified for each DNA that would generate a single DNA fragment of a defined size to which the *ema1* gene hybridizes. For each strain, there was only one strongly hybridizing DNA fragment, thus suggesting that other P450 genes were not detected under the high stringency hybridization conditions used in these experiments. Each DNA was digested with the appropriate restriction endonuclease, and the DNA was subjected to agarose gel electrophoresis. DNA in a narrow size range that included the size of the *ema1*-hybridizing fragment was excised from the gel. The size selected DNA was ligated into an appropriate cloning plasmid and this ligated plasmid was used to transform *E. coli*. The *E. coli* clones from each experiment were screened by colony hybridization with the *ema1* gene fragment to identify clones containing the *ema1*-homologous DNA fragment.

The nucleotide sequence of the cloned DNA in each *ema1*-homologous clone was determined and examined for the presence of a gene encoding a P450 enzyme with homology to *ema1*. In this way, *ema1*-homologous genes were isolated from 14 of the 15 other active strains. The nucleotide and deduced amino acid sequences of these are referenced in Table 4 as SEQ ID NOS:5-32 and 94-95. The relationship of these enzymes can be shown in the form of a phylogenetic tree. Such a phylogenetic tree can be generated using the commercially available GCG Wisconsin software program version 1.0 (Madison, WI).

Table 4

Strain Number		Classification	SEQ ID NO (nucleotide and amino acid, respectively)
R-0922	<i>ema1</i>	<i>Streptomyces tubercidicus</i>	2. 1 and 2
I-1529	<i>ema2</i>	<i>Streptomyces tubercidicus</i>	3 and 4
1053	<i>ema3</i>	<i>Streptomyces rimosus</i>	5 and 6
R-0401	<i>ema4</i>	<i>Streptomyces lydicus</i>	7 and 8
I-1525	<i>ema5</i>	<i>Streptomyces</i> sp.	9 and 10
DSM-40241	<i>ema6</i>	<i>Streptomyces chattanoogensis</i> *	3. 11 and 12
IHS-0435	<i>ema7</i>	<i>Streptomyces</i> sp.	13 and 14
C-00083	<i>ema8</i>	<i>Streptomyces albofaciens</i>	15 and 16
MAAG-7479	<i>ema9</i>	<i>Streptomyces platensis</i>	17 and 18
A/96-1208710	<i>ema10</i>	<i>Streptomyces kasugaensis</i>	4. 19 and 20
R-2374	<i>ema11</i>	<i>Streptomyces rimosus</i>	21 and 22
MAAG-7027	<i>ema12</i>	<i>Streptomyces tubercidicus</i>	5. 23 and 24
Tue-3077	<i>ema13</i>	<i>Streptomyces platensis</i>	25 and 26
I-1548	<i>ema14</i>	<i>Streptomyces platensis</i>	27 and 28
NRRL-2433	<i>ema15</i>	<i>Streptomyces lydicus</i>	6. 29 and 30
MAAG-0114	<i>ema16</i>	<i>Streptomyces lydicus</i>	31 and 32
DSM-40261	<i>ema17</i>	<i>Streptomyces tubercidicus</i>	94 and 95

* This strain was shown to be in the *chattanoogensis* species by 16s rDNA analysis; however, classical taxonomic methods used by the German culture collection (DSMZ) showed it to be *saracepticus*.

EXAMPLE XII**Construction of His-tagged *ema1* and *ema1* Homologs to Facilitate Enzyme Purification**

In order to purify the P450_{Ema1} enzyme and the P450 enzymes encoded by the *ema1* homologs from other biocatalysis strains, each of the P450 genes was cloned into the *E. coli* expression plasmid pET-28b(+) (commercially available from Novagen, Madison, WI). The pET-28 plasmids are designed to facilitate His-tag fusions at either the N-, or C-terminus and to provide strong expression of the genes in *E. coli* from the T7 phage promoter. In many cases, the coding sequence of the *ema* genes begins with the sequence ATGT. These genes were amplified by PCR such that the primers on the 5' end incorporated a PciI recognition site (5' ATATGT 3') at the 5' terminus. The last four bases of the PciI site correspond to the ATGT at the beginning of the *ema* gene coding sequence.

PCR primers at the 3' end of the genes were designed to remove the translation stop codon at the end of the *ema* gene coding sequence and to add an XhoI recognition site to the 3' terminus. The resulting PCR fragments were restricted with PciI and XhoI to generate PciI ends at the 5' termini and XhoI ends at the 3' termini, thereby facilitating cloning of the fragments into pET-28b(+) previously restricted with NcoI and XhoI. Since PciI and NcoI ends are compatible, the fragments were cloned into pET-28b(+) in the proper orientation to the T7 promoter and ribosome binding site in the plasmid to provide expression of the genes.

At the 3' end of each *ema* gene, the coding sequence was fused in frame at the XhoI site to the His-tag sequence followed by a translation stop codon. This results in the production of an Ema enzyme with six histidine residues added to the C-terminus to facilitate purification on nickel columns.

In the case of *ema* genes in which the ATG translation initiation codon is not followed by a T nucleotide, the *ema* genes were amplified by PCR using a different strategy for the 5' end. The primers at the 5' end were designed to incorporate a C immediately preceding the ATG translation initiation codon and the primers at the 3' end were the same as described above. The PCR fragments that were amplified were restricted with XhoI to create an XhoI end at the 3'-terminus and the 5' end was left as a blunt end. These fragments were cloned into pET-28b(+) that had been restricted with NcoI, but the NcoI ends were made blunt-ended by treatment with mung bean exonuclease, and restricted with XhoI.

In this manner, the *ema* genes were cloned into pET-28b(+) to create a functional fusion with the T7 promoter and the His-tag at the C-terminus as described previously. All His-tagged *ema* genes were sequenced to ensure that no errors were introduced by PCR.

Large amounts of the P450_{Ema1} and P450_{Ema2} enzymes were isolated and purified by standard protocols. *E. coli* strain BL21 DE3 (commercially available from Invitrogen; Carlsbad, CA) containing the T7 RNA polymerase gene under the control of the inducible *tac* promoter and the appropriate pET-28/ema plasmid was cultured and the cells were harvested and lysed. The lysates were applied to Ni-NTA columns (commercially available from Qiagen Inc., Valencia, CA) and the protein were purified according to the procedure recommended by the manufacturer.

Purified His-tagged P450_{Ema1} and P450_{Ema2} were highly active in *in vitro* activity assays as evidenced by a high rate of conversion of avermectin to 4"-keto-avemectin.

EXAMPLE XIII

Expression of ema1 in *Pseudomonas*

The ema1 gene constructs were next introduced into *P. putida* (wildtype *P. putida* commercially available from the American Type Culture Collection, Manassas, Virginia; ATCC Nos. 700801 and 17453). The *ema1* and *ema1/fd233* gene fragments were cloned as PacI/PmeI fragments into the plasmid pUK21 (Viera and Messing, *Gene* **100**:189-194, 1991). The fragments were cloned into a position located between the tac promoter (P_{tac}) and terminator (T_{tac}) on pUK21 in the proper orientation for expression from the tac promoter. The P_{tac} -*ema1*- T_{tac} and P_{tac} -*ema1/fd233*- T_{tac} gene fragments were removed from pUK21 as BglII fragments and these were cloned into the broad host-range, transmissible plasmid, pRK290 (Ditta *et al.*, *Proc. Natl. Acad. Sci. USA* **77**:7347-7351, 1980) to create plasmids pRK-*ema1* and pRK-*ema1/fd233* (Figure 3). These plasmids were introduced into *P. putida* strains ATCC 700801 and ATCC 17453 by conjugal transfer from *E. coli* hosts by standard methodology (Ditta *et al.*, *Proc. Natl. Acad. Sci. USA* **77**:7347-7351, 1980).

P. putida ATCC 700801 and ATCC 17453 containing plasmids pRK-*ema1* or pRK-*ema1/fd233* were tested for the ability to catalyze the oxidation of avermectin. The results shown in Table 3 demonstrate that these strains are able to catalyze this reaction.

EXAMPLE XIV

Identification of Genes Encoding Ferredoxins That Are Active With the
P450_{Ema1} Monooxygenase

P450 monooxygenases require two electrons for each hydroxylation reaction catalyzed (Mueller *et al.*, “Twenty-five years of P450_{cam} research: Mechanistic Insights into Oxygenase Catalysis.” Cytochrome P450, 2nd Edition, P.R. Ortiz de Montellano (ed.), pp. 83-124; Plenum Press, NY 1995). These electrons are transferred to the P450 monooxygenase one at a time by a ferredoxin. The electrons are ultimately derived from NAD(P)H and are passed to the ferredoxin by a ferredoxin reductase. Specific P-450 monooxygenase enzymes have a higher activity when they interact with a specific ferredoxin. In many cases, the gene encoding a ferredoxin that interacts specifically with a given P450 monooxygenase is located adjacent to the gene encoding the P450 enzyme.

As described above, in addition to the *ema1* gene, four P450 genes from strain R-922 and seven P450 genes from strain I-1529 (*see Example VI*) were isolated and sequenced. In some of these, there was sufficient sequence information about the DNA flanking the P-450 genes to look for the presence of associated ferredoxin genes. By this approach, two unique ferredoxin genes were identified from each of the two strains. Ferredoxin genes *fd229* and *fd230* were identified from strain R-922, and *fd233* and *fdEA* were identified from strain I-1529. In addition, a ferredoxin reductase gene was found to reside adjacent to the *fdEA* gene from strain I-1529.

In order to test the biological activity of each of these ferredoxins in combination with P450_{Ema1}, each individual ferredoxin gene was amplified by PCR to produce a gene fragment that included a blunt 5'-end, the native ribosome-binding site and ferredoxin gene coding sequence, and a PmeI restriction site on the 3'-end. Each such ferredoxin gene fragment was cloned into the PmeI site located 3' to the *ema1* gene in plasmid pTUA-*ema1*. In this way, artificial operons consisting of the *ema1* gene and one of the ferredoxin genes operably linked to a functional promoter were created.

In the case of the *fdEA* ferredoxin gene in which a ferredoxin reductase gene, *freEA*, was found to be located adjacent to the *fdEA* gene, a DNA fragment containing both the *fdEA* and *freEA* genes was generated by a similar PCR strategy. This gene fragment was also cloned in the PmeI site of plasmid pTUA-*ema1* as described for the other ferredoxin genes.

Each *ema1*-ferredoxin gene combination was tested for biological activity by introduction of the individual *ema1*-ferredoxin gene plasmids into *S. lividans* strain ZX7. The biocatalysis activity derived from each plasmid in *S. lividans* was determined. Of the four different constructs, only the ferredoxin gene *fd233* derived from strain I-1529 provided increased activity when compared to the expression of *ema1* alone in the same plasmid and host background (see Table 3). The pTUA-*ema1/fd233* plasmid in *S. lividans* provided approximately 1.5 to 3- fold higher activity compared to the pTUA-*ema1* plasmid. The other three plasmids containing the other ferredoxin genes gave results essentially the same as the plasmid with only the *ema1* gene. Likewise, the pTUA-*ema1/fdEA/freEA* plasmid did not yield results different from those of pTUA-*ema1*. The nucleotide and deduced amino acid sequences of the *fd233* gene are shown in SEQ ID NOs:35 and 36, respectively.

A BLAST analysis of the nucleotide and amino acid sequences of *fd233* revealed that the closest matches were to ferredoxins from *S. coelicolor* (GenBank Accession AL445945) and *S. lividans* (GenBank Accession AF072709). At the nucleotide level, *fd233* shares 80 and 79.8 % identity with the ferredoxin genes from *S. coelicolor* and *S. lividans*, respectively. At the peptide level, *fd233* shares 79.4 and 77.8% identity with the ferredoxins from *S. coelicolor* and *S. lividans*, respectively.

Since *fd233* is derived from strain I-1529 and *ema1* is from strain R-922, the proteins encoded by the two genes cannot interact with each other in nature. In an approach designed to identify a ferredoxin gene from strain R-922 that is homologous to the *fd233* gene and that might encode a ferredoxin that interacts optimally with the P450_{Ema1}, the *fd233* gene was used as a hybridization probe to a gene library of DNA from strain R-922. A strongly hybridizing cosmid, pPEH232, was identified and the hybridizing DNA was cloned and sequenced. Comparison of the deduced amino acid sequences from *fd233* and the ferredoxin gene on cosmid pPEH232, *fd232*, revealed that they differed in only a single amino acid.

In a similar manner, plasmid pTUA-*ema1-fd232* was constructed and tested in *S. lividans* ZX7. This plasmid gave similar results as those obtained with plasmid pTUA-*ema1-fd233* (see Table 3). The nucleotide and deduced amino acid sequences of *fd232* are shown in SEQ ID NOs:37 and 38, respectively.

The *ema1-fd233* operon was also subcloned, as a PacI-PmeI fragment, into pTBBKA and pEAA that had been digested with the same restriction enzymes. *S. lividans*

ZX7::pTBBKA-ema1-fd233, and *S. lividans ZX7::pEAA-ema1-fd233* were tested in the avermectin conversion assay and found to have higher activities than the strains harboring the *ema1* gene alone in the comparable plasmids (see Table 3).

EXAMPLE XV

Heterologous Expression of P450_{Ema1} and P450_{Ema2} in Other Cells

The expression constructs pRK-*ema1* (Example XIII) and pRK-*ema2* (created in a way analogous to that described in Example XIII for pRK-*ema1*) were mobilized by conjugation into three fluorescent soil *Pseudomonas* strains. Conjugation was performed according to standard methods (Ditta *et al.*, *Proc. Natl. Acad. Sci. USA* 77:7347-7351, 1980). The strains were: *P. fluorescens* MOCG134, *P. fluorescens* Pf-5, and *P. fluorescens* CHAO. Standard resting cell assays for the conversion of avermectin to 4"-ketoavermectin were conducted for each of the transconjugants. For strains Pf-5 and CHAO, the levels of conversion were below the detection limit. Strain MOCG134 yielded 3% conversion for *ema1* and 5% for *ema2*.

In addition, the constructs listed in the Table 5 were introduced into *Streptomyces avermitilis* MOS-0001 by protoplast-mediated transformation (Kieser, T.; Bibb, M.J.; Buttner, M.J.; Chater, K.F.; Hopwood, D.A. (eds.): *Practical Streptomyces Genetics*. The John Innes Foundation, Norwich (England), 2000), (Stutzman-Engwall, K. *et al.* (1999) *Streptomyces avermitilis* gene directing the ratio of B₂:B₁ avermectins, WO 99/41389).

Table 5

Construct	% Conversion of avermectin, 16 hrs
None	0
pTBBKA- <i>ema1</i>	10.90 +/- 3.48
pTUA- <i>ema1</i>	5.326 +/- 2.19
pEAA- <i>ema1</i>	6.74 +/- 0.08
pTBBKA- <i>ema1A/fd233</i>	28.50 +/- 0.20
pTUA- <i>ema1A/fd233</i>	23.97 +/- 5.95

Wild-type *Str. avermitilis* MOS-0001 was tested and found to be incapable of the oxidation of avermectin to 4"-ketoavermectin.

Transformed *S. avermitilis* strains MOS-0001::pTBBKA-*ema1*, MOS-0001 (pTUA-*ema1*), MOS-0001::pEAA-*ema1*, MOS-0001::pTBBKA-*ema1A/fd233*, and MOS-0001 (pTUA-*ema1A/fd233*) were each tested for their ability to oxidize avermectin to 4"-keto-avermectin using resting cells. To do this, the whole cell biocatalysis assay described above (including analysis method) was performed. Note that for the whole cell biocatalysis assay, transformed *Streptomyces avermitilis*, like strain R-922, was grown in PHG medium and, again like strain R-922, had a reaction time of 16 hours (*i.e.*, during which time the 500 mg transformed *Streptomyces avermitilis* wet cells in 10 ml of 50 mM potassium phosphate buffer, pH 7.0, were shaken at 160 rpm at 28°C in the presence of 15 µl of a solution of avermectin in isopropanol (30 mg/ml)).

As shown in Table 5, in the presence of the inducer, thiostrepton (5 µg/ml), the *ema1*- or *ema1A/fd233*-containing strains MOS-0001::pTBBKA-*ema1*, MOS-0001::pTBBKA-*ema1A/fd233*, MOS-0001 (pTUA-*ema1*), MOS-0001 (pTUA-*ema1A/fd233*) were found to oxidize avermectin to 4"-keto-avermectin as evidenced by the appearance of the oxidized 4"-keto-avermectin compound. Note that the *S. avermitilis* strain MOS-0001::pEAA-*ema1* demonstrated this oxidation activity in the absence of thiostrepton since in this strain the *ema1* gene is expressed from the *ermE* promoter that does not require induction.

Thus, expression of the *ema1* P450 monooxygenase gene in various *Streptomyces* and *Pseudomonas* strains provided recombinant cells that were able to convert avermectin to 4"-ketoavermectin in resting cell assays.

Next, expression and activity of P450_{Ema1} monooxygenase was tested in *E. coli*. To do this, the *ema1* gene was cloned into the *E. coli* expression plasmid pET-28b(+) (commercially available from Novagen, Madison, WI) as described previously. *E. coli* strain BL21 DE3 (commercially available from Invitrogen; Carlsbad, CA) that contains the T7 RNA polymerase gene under control of the inducible *tac* promoter and the pET-28/*ema1* plasmid was cultured in 50 ml LB medium containing 5 mg/l kanamycin in a 250-ml flask with one baffle, for 16 hours at 37°C, with shaking at 130 rpm. 0.5 ml of this culture was used to inoculate 500 ml LB medium with 5 mg/l kanamycin in a 2-liter flask with one baffle, and the

culture was incubated for 4 hours at 37°C followed by 4 hours and 30°C, with shaking at 130 rpm throughout. The cells were harvested by centrifugation, washed in 50 mM potassium phosphate buffer, and centrifuged again.

For the resting cell assays, 90 mg wet cells were weighed into deep-well plates in triplicate and resuspended in 0.5 ml 50 mM potassium phosphate buffer. For cell-free extracts, 4 grams wet cells in 8 ml disruption buffer were disrupted in French press.

For the resting cell assays, 5 µl of substrate (2.5 mg/ml in 2-propanol) was added to the cell suspension. The plate was sealed with air permeable foil, and the reaction was incubated on an orbital shaker at 1000 rpm at 28°C for 22 hours. No conversion of avermectin to 4"-ketoavermectin was detected.

For the cell-free assays, 100 µl cell free extract, 1µl substrate solution (20 mg/ml) in 2-propanol, 5 µl 100 mM NADPH, 10 µl ferredoxin, 10 µl ferredoxin reductase, and 374 µl potassium phosphate buffer pH 7.0 were added as described in Example III, and the assay was incubated at 30°C with shaking at 600 rpm for 20 hours. 9.2% +/- 0.3% of avermectin was converted to 4"-ketoavermectin.

Thus, expression of the *ema1* gene in *E. coli* resulted in the production of the active Ema1 P450 monooxygenase enzyme which, when purified from the cells, was able to convert avermectin to 4"-ketoavermectin.

EXAMPLE XVI

Identification and Cloning of Genes Encoding Ferredoxin Reductases that Support Increased Activity of the P450_{Ema1} Monooxygenase

The electron transport pathway that supports the activity of P450 monooxygenases also includes ferredoxin reductases. These proteins donate electrons to the ferredoxin and, as is the case with ferredoxins and P450 monooxygenases, specific ferredoxin reductases are known to be better electron donors for certain ferredoxins than others.

According, a number of ferredoxin reductase genes from *Streptomyces* strains were cloned and were evaluated for their impacts on the biocatalysis reaction. To do this, numerous bacterial ferredoxin reductase (Fre) protein sequences were retrieved from NCBI and aligned with the program Pretty from the GCG package. Two conserved regions,

approximately 266 amino acid residues apart, were used to make degenerate oligonucleotides for PCR. The forward primer (CGSCCSCCSCTSWSSAAS (SEQ ID NO:96; where "S" is C or G; and "W" is A or G)) and the reverse primer (SASSGCSTTSBCCCARTGYTC (SEQ ID NO:97; where "S" is C or G; "B" is C, G, or T; "R" is A or G; and "Y" is C or T)) were used to amplify 800 bp products from the biocatalytically active *Streptomyces* strains R-922 and I-1529. These pools of products were cloned into TOPO TA cloning vectors (commercially available from Invitrogen Inc., Carlsbad, CA), and 20 clones each from R922 and I-1529 were sequenced according to standard methods (see, e.g., *Current Protocols in Molecular Biology*, eds. Ausubel *et al.*, John Wiley & Sons, Inc. 2000). Sequencing revealed that 4 unique *fre* gene fragments were isolated from the strains: three from R922 (*fre3*, *fre12*, *fre14*) and one from I-1529 (*fre16*). The *fre3*, *fre12*, *fre14*, and *fre16* gene fragments were used as probes to identify full-length ferredoxin reductases from genomic clone banks of *Streptomyces* strains R922 and I-1529. By this approach, the complete coding sequence of each of the 4 different *fre* genes was cloned and sequenced. The nucleic acid and amino acid sequences are provided as follows: *fre3* (SEQ ID NOs:98 and 99); *fre12* (SEQ ID NOs:100 and 101); *fre14* (SEQ ID NOs:102 and 103); and *fre16* (SEQ ID NOs:104 and 105).

In order to assess the biological activity of each *fre* gene in relation to the activity of Ema1, each gene was inserted into the *ema1/fd233* operon described above, 3' to the *fd233* gene. This resulted in the formation of artificial operons consisting of the *ema1*, *fd233*, and individual *fre* genes that were expressed from the same promoter. The *ema1/fd233/fre* operons were cloned into the *Pseudomonas* plasmid pRK290 and introduced into 3 different *P. putida* strains. These strains were then analysed for Ema1 biocatalysis activity using the whole cell assay and one of the genes, the *fre* gene *fre16* from strain I-1529, was found to increase the activity of P450_{Ema1} monooxygenase by approximately 2-fold. This effect was strain specific, as it was seen only in one of the *P. putida* strains, ATCC Deposit No. 17453, and not in the other two. In *P. putida* strain ATCC 17453, the presence of *fre* gene *fre16* resulted in 44% conversion of avermectin to 4"-keto-avermectin, as compared to 23% without this gene. The other *fre* genes had no impact on the biocatalysis activity in any of the *P. putida* strains tested.

In a similar approach, each of the *ema1/fd233/fre* operons were cloned into the *Streptomyces* plasmids pTUA, pTBBKA, and pEAA, and introduced into *S. lividans* strain

ZX7. In each case there was no impact in *S. lividans* by any of the *fre* genes on biocatalysis activity.

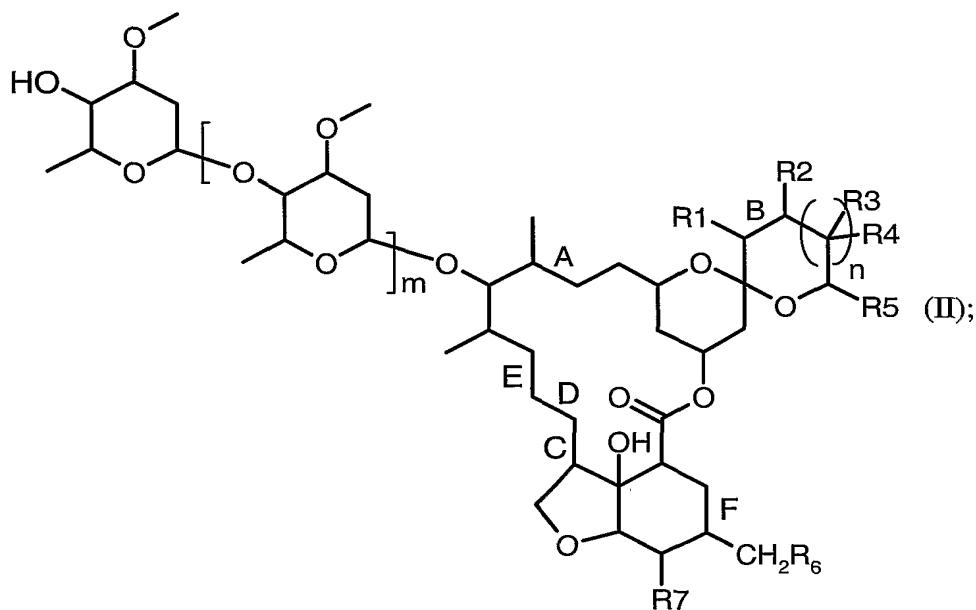
EQUIVALENTS

Those skilled in the art will recognize, or be able to ascertain, using no more than routine experimentation, numerous equivalents to the specific substances and procedures described herein. Such equivalents are considered to be within the scope of this invention.

The patent and scientific literature referred to herein establishes knowledge that is available to those with skill in the art. The issued patents, applications, and references, including GenBank database sequences, that are cited herein are hereby incorporated by reference to the same extent as if each was specifically and individually indicated to be incorporated by reference. Any conflict between any reference cited herein and the specific teachings of this specification shall be resolved in favor of the latter. Likewise, any conflict between an art-understood definition of a word or phrase and a definition of the word or phrase as specifically taught in this specification shall be resolved in favor of the latter.

What is claimed is:

1. A purified nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II)



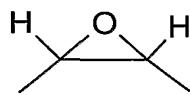
wherein

10 R₁-R₇ represent, independently of each other hydrogen or a substituent;

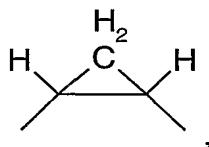
 m is 0, 1 or 2;

 n is 0, 1, 2 or 3; and

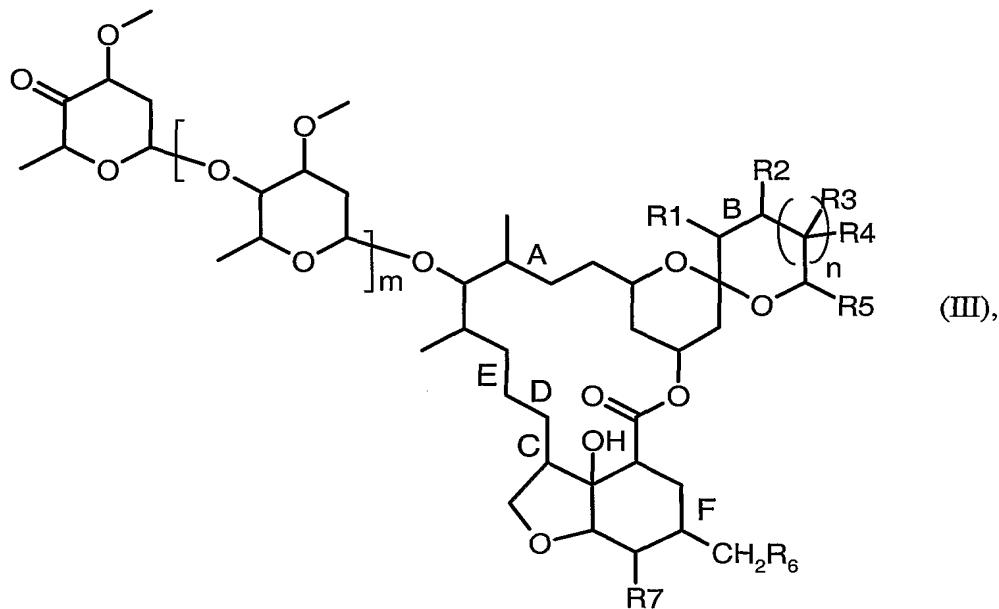
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
in order to produce a compound of the formula (III)



wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (II) above.

- 10
2. The nucleic acid molecule of claim 1, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"ketovermectin.
 - 15
 3. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase, which polypeptide is substantially similar, and has between at least 50%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2.

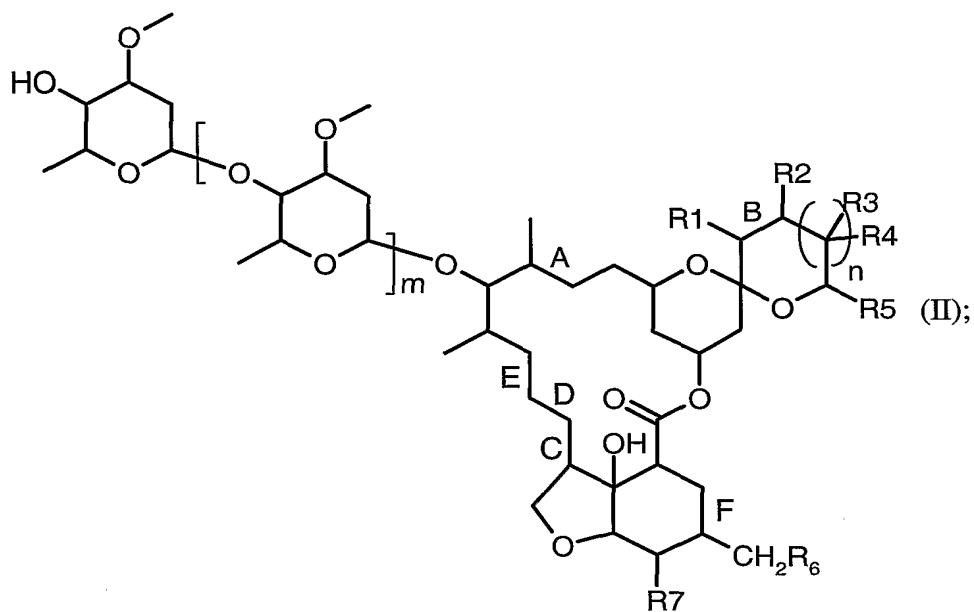
4. The nucleic acid molecule of claim 3 comprising a nucleotide sequence
 - a) as given in SEQ ID NO:1;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - 5 d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
 - e) complementary to (a), (b) or (c);
 - f) which is the reverse complement of (a), (b) or (c); or
 - 10 g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin. .
- 15 5. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that is at least 66 % identical to SEQ ID NO:1.
- 20 6. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase, which polypeptide is substantially similar, and has at least between 60%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- 25 7. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18,

SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

8. The nucleic acid molecule of claims 1 or 2 comprising a nucleotide sequence
 - a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94;
 - b) having substantial similarity to (a);
 - c) capable of hybridizing to (a) or the complement thereof;
 - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;
 - e) complementary to (a), (b) or (c); and
 - f) which is the reverse complement of (a), (b) or (c).
 - g) . which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.
9. The nucleic acid molecule of claim 8, comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, and SEQ ID NO:94.
10. The nucleic acid molecule of anyone of claims 1 to 9, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain.

11. The nucleic acid molecule of anyone of claims 1 to 10 further comprising a nucleic acid sequence encoding a tag which is linked to the P450 monooxygenase via a covalent bond.

5 12. A polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II)



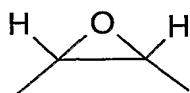
10 wherein

R₁-R₇ represent, independently of each other hydrogen or a substituent;

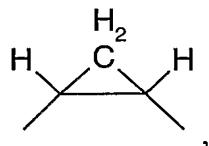
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

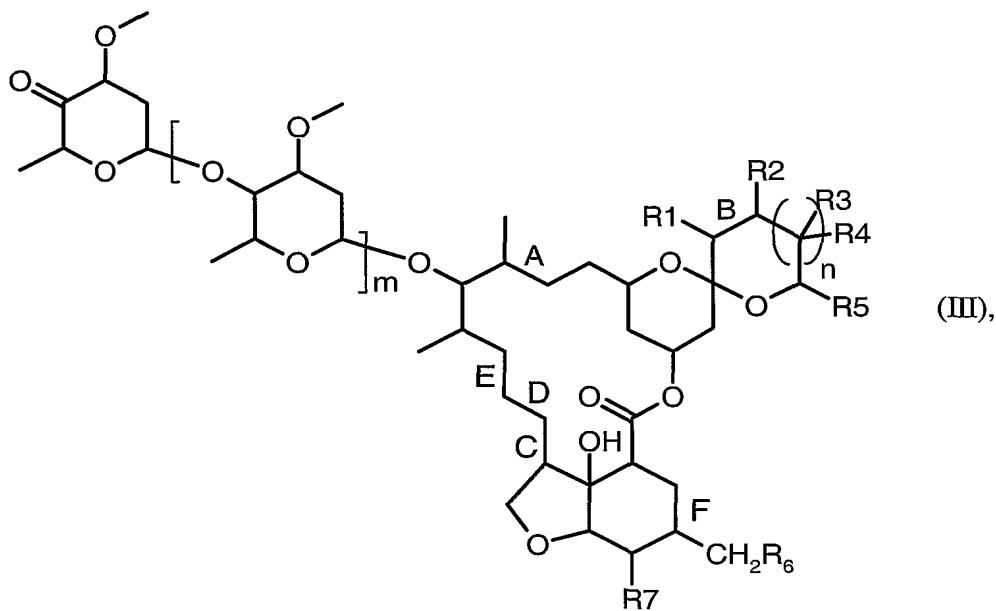
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
in order to produce a compound of the formula (III)



wherein

R_1 - R_7 , m , n , A , B , C , D , E and F have the same meanings as given for formula (II) above.

10 13. A polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"ketovermectin.

14. The polypeptide of claims 12 or 13 that comprises an amino acid sequence that is encoded by a nucleic acid molecule

- a) as given in SEQ ID NO:1 or the complement thereof;
- b) having substantial similarity to (a);
- c) capable of hybridizing to (a) or the complement thereof;

- d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
- e) complementary to (a), (b) or (c);
- 5 f) which is the reverse complement of (a), (b) or (c); or
- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.

10 15. The polypeptide of claims 12 to 14, comprising an amino acid sequence that is at least 50% identical to SEQ ID NO:2.

16. The polypeptide of claims 12 or 13 comprising an amino acid sequence that is encoded by a nucleic acid molecule

- 15 a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;
- b) having substantial similarity to (a);
- c) capable of hybridizing to (a) or the complement thereof;
- 20 d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof, or the complement thereof;
- e) complementary to (a), (b) or (c);
- f) which is the reverse complement of (a), (b) or (c); or

g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.

5 17. The polypeptide of claim 16, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, and SEQ ID NO:95.

10

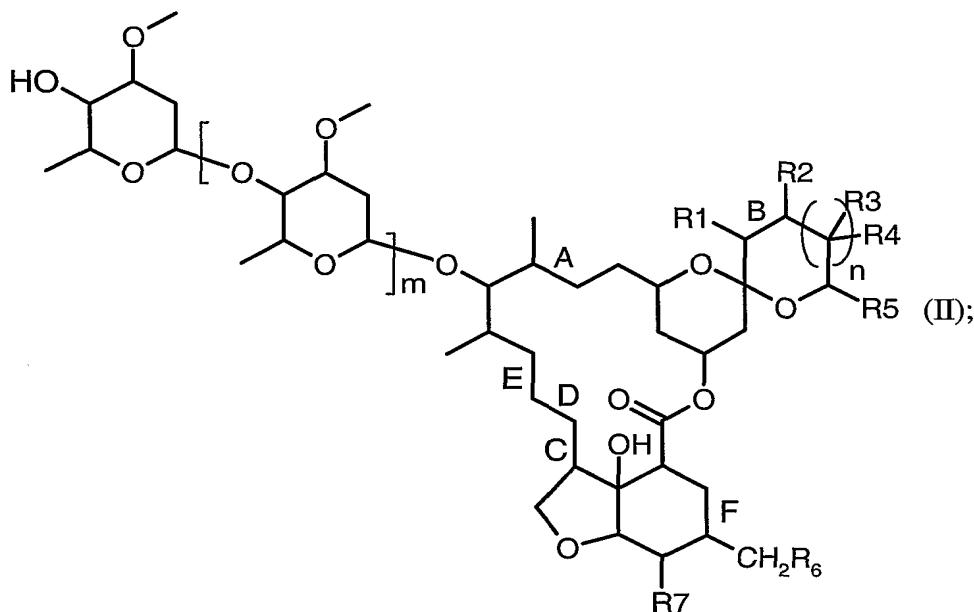
18. The polypeptide of anyone of claims 12 to 17, further comprising a tag.

19. A binding agent that specifically binds to the polypeptide of anyone of claims 12 to 18.

15 20. The binding agent of claim 20, wherein the binding agent is an antibody.

21. A family of polypeptides exhibiting an enzymatic activity of a P450 monooxygenase, wherein each member of the family is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II)

20



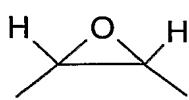
wherein

R_1-R_7 represent, independently of each other hydrogen or a substituent;

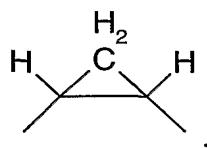
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

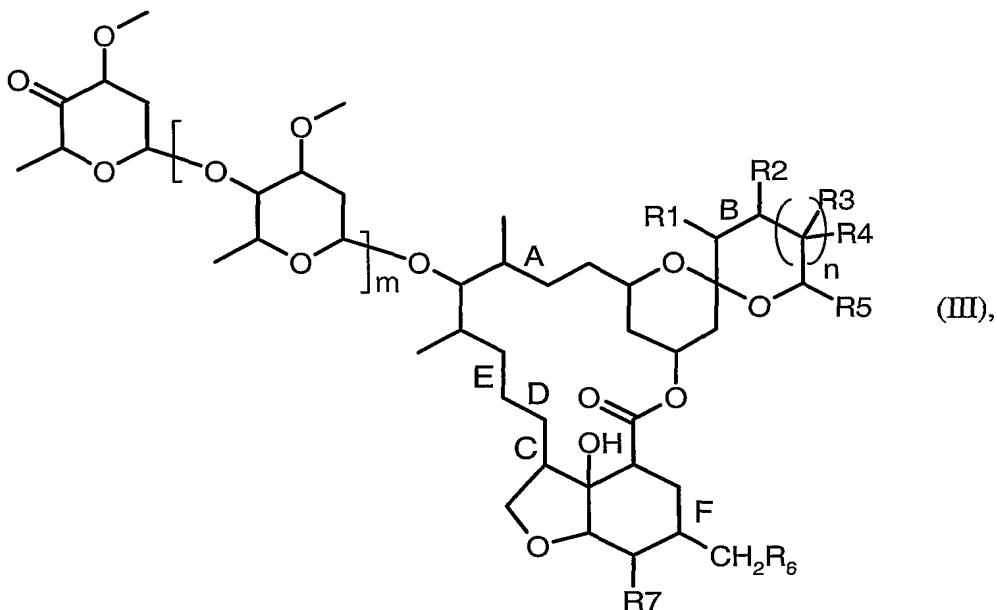
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
in order to produce a compound of the formula (III)



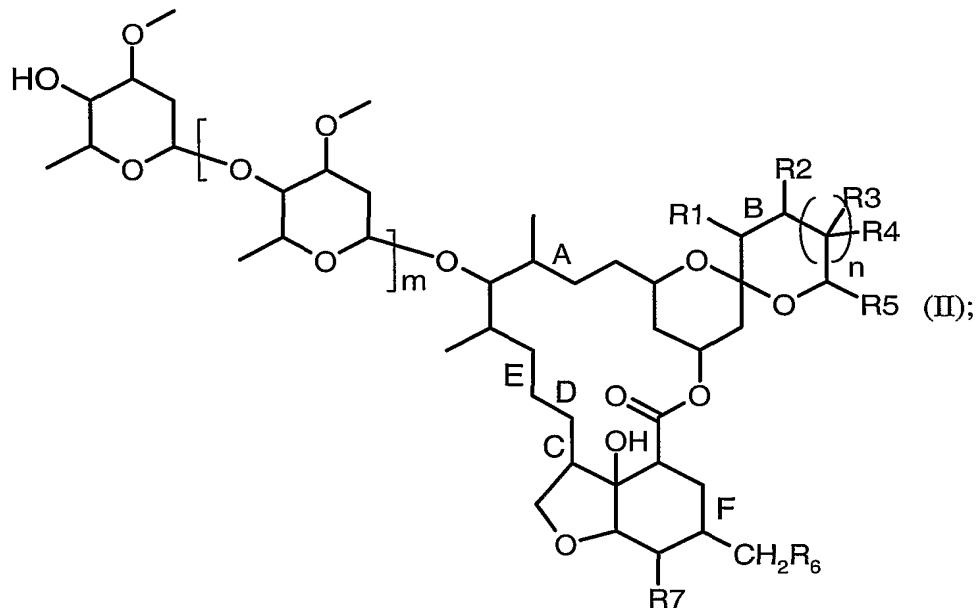
wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (II) above.

- 5 22. A family of polypeptides exhibiting an enzymatic activity of a P450 monooxygenase,
wherein each member of the family oxidizes avermectin to 4" keto-avermectin.

10 23. The family of claims 21 or 22, wherein each member of the family is comprises an amino
acid sequence that is at least 50% identical to SEQ ID NO:2.

15 24. A purified nucleic acid molecule comprising a nucleotide sequence encoding a
polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin
reductase,respectively, wherein the nucleic acid molecule is isolated from a *Streptomyces*
strain comprising a P450 monooxygenase that is capable of regioselectively oxidizing the
alcohol at position 4" of a compound of formular (II)



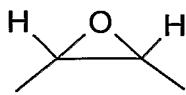
wherein

R_1 - R_7 represent, independently of each other hydrogen or a substituent;

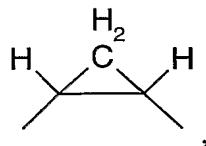
m is 0, 1 or 2;

5 n is 0, 1, 2 or 3; and

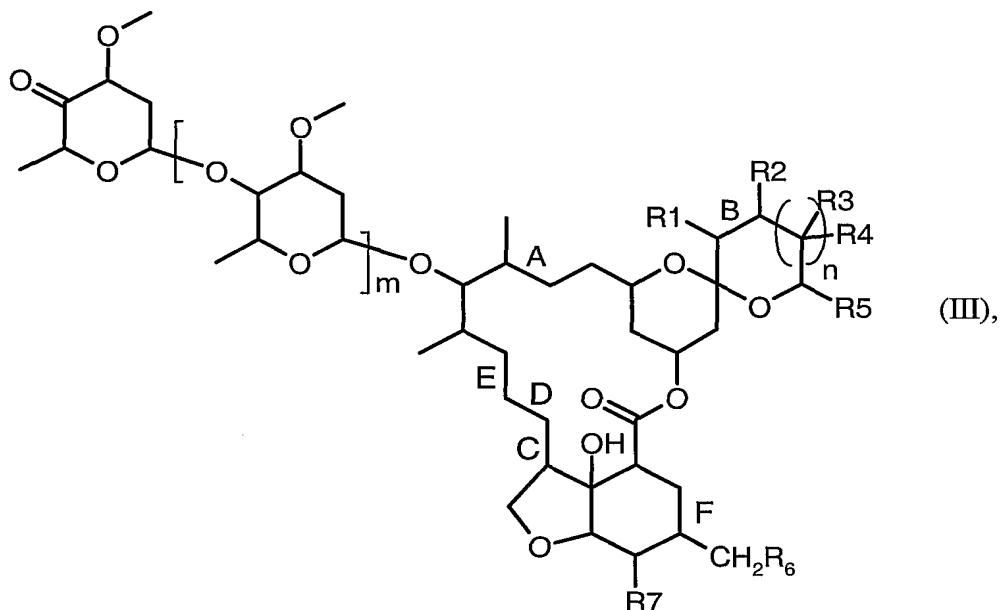
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
in order to produce a compound of the formula (III)

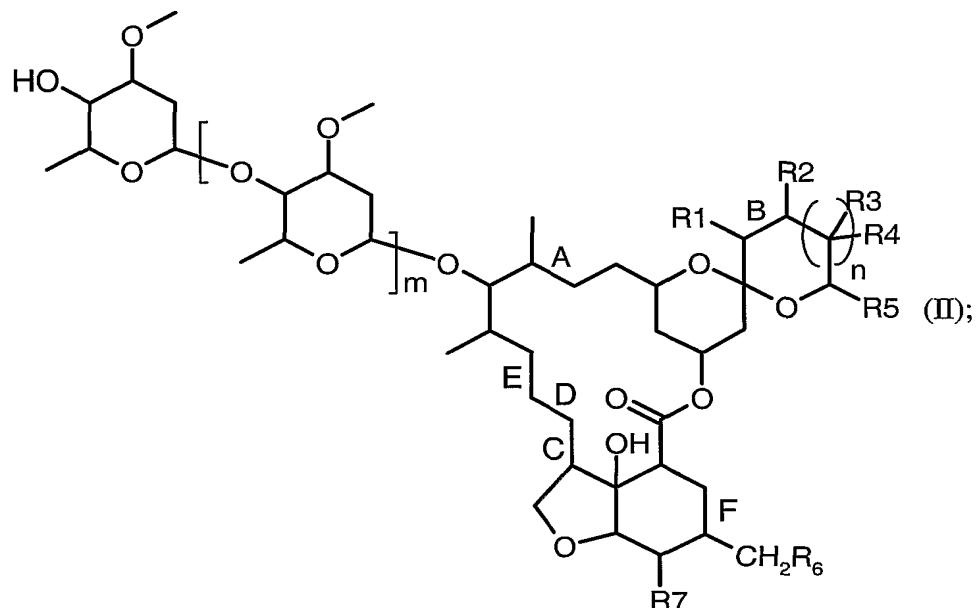


wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (II) above.

- 5 25. A purified nucleic acid molecule according to claim 24 comprising a nucleotide sequence encoding a polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"ketovermectin.
- 10 26. The nucleic acid molecule of claim 25, comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:35 and SEQ ID NO:37.
- 15 27. The nucleic acid molecule of claim 25, comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, and SEQ ID NO:104.
28. A polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the polypeptide is isolated from a *Streptomyces* strain comprising a

P450 monooxygenase that is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II)



5

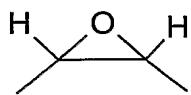
wherein

R₁-R₇ represent, independently of each other hydrogen or a substituent;

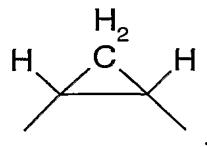
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

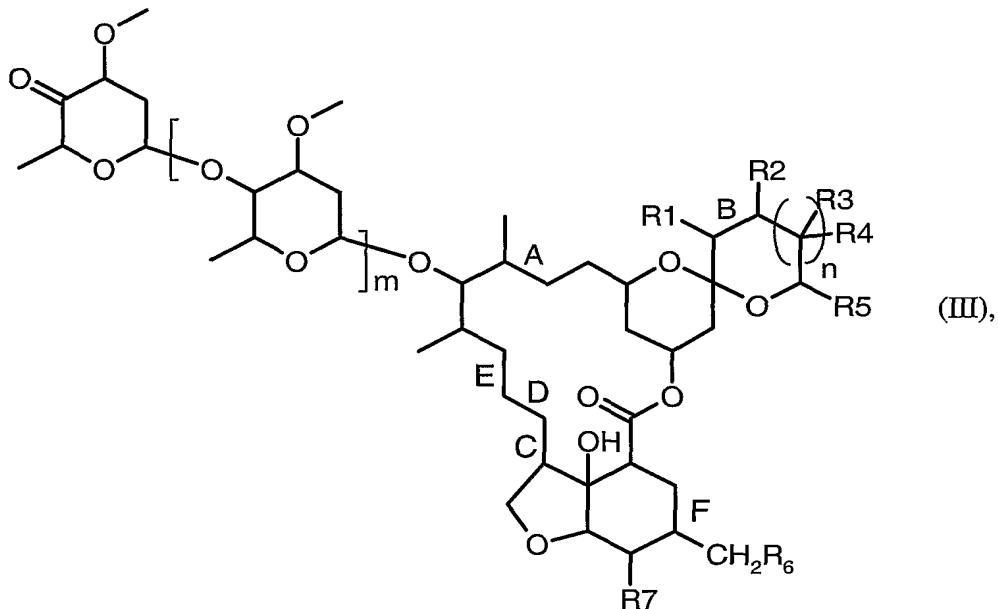
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
in order to produce a compound of the formula (III)



5

wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (II) above.

29. A polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the ferredoxin protein is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"ketovermectin.

10

30. The ferredoxin protein of claim 29, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:36 and SEQ ID NO:38.

15

31. The ferredoxin reductase protein of claim 29, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, and SEQ ID NO:105.

32. A cell genetically engineered to comprise a nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase according to anyone of claims 1 to 11.

5 33. The cell of claim 32 further comprising a nucleic acid molecule encoding a ferredoxin protein and a ferredoxin reductrase protein, respectively, or a combination thereof.

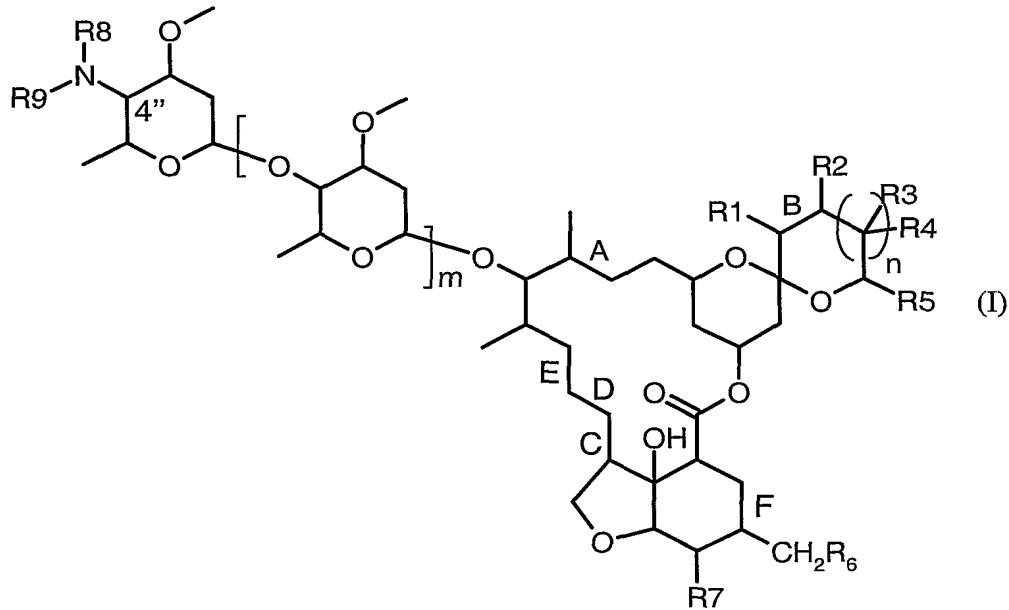
34. The cell of claims 32 or 33, wherein the nucleic acid molecule is positioned for expression in the cell.

10

35. The cell of anyone of claims 32 to 34, wherein the cell is a genetically engineered cell selected from the group consisting of a *Streptomyces* strain cell and a *Pseudomona* strain cell, and an *Escherichia coli* strain cell.

15 36. The cell of claim 35, wherein the cell has NRRL Designation No. B-30478 and NRRL Designation No.B-30479, respectively.

37. A method for the preparation a compound of the formula



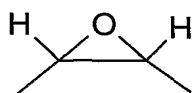
in which

R₁-R₉ represent, independently of each other hydrogen or a substituent;

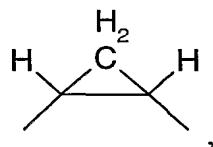
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula

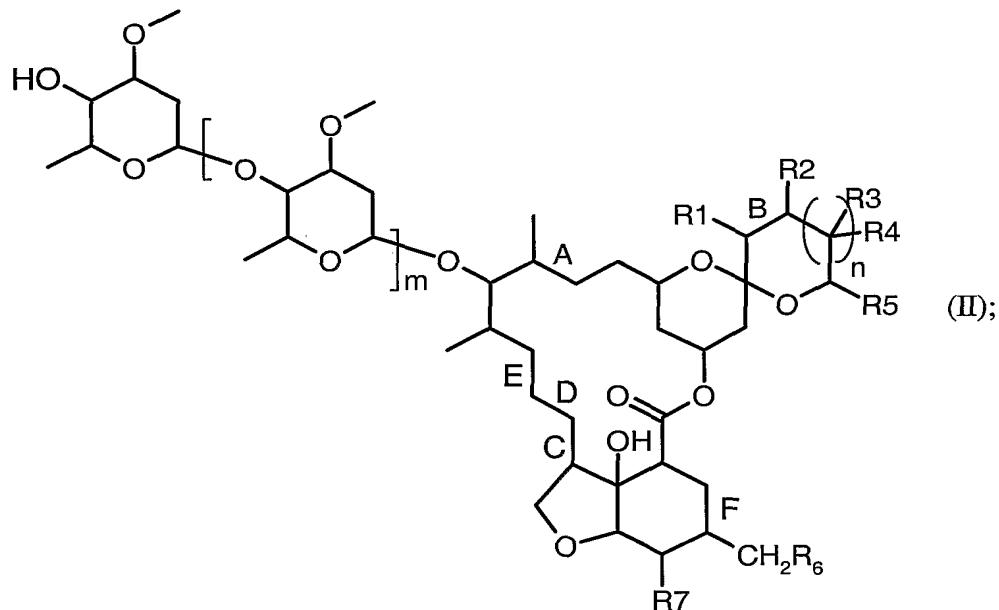


, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,
which process comprises

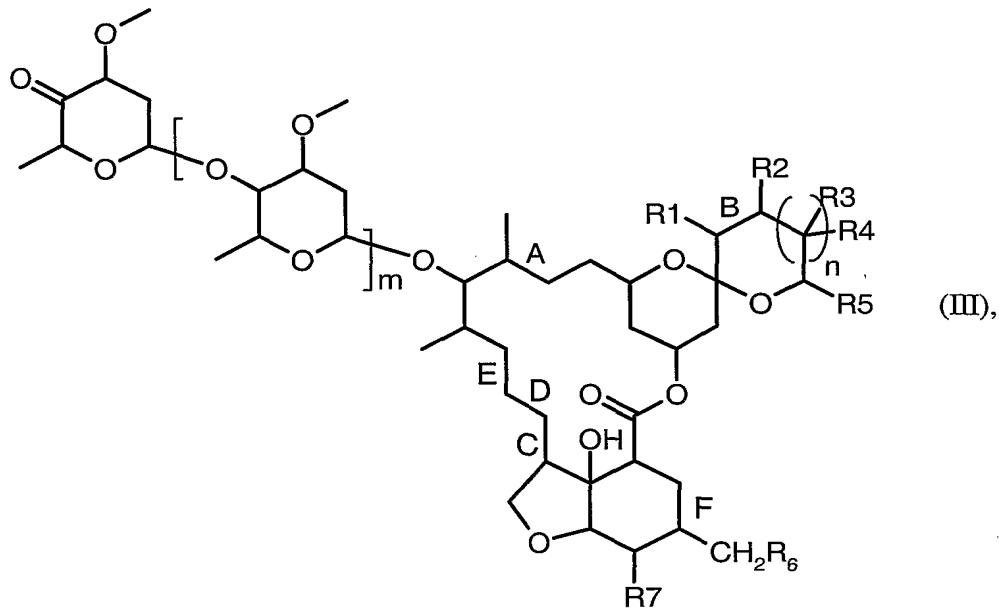
1) bringing a compound of the formula



wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regioselectively oxidising the alcohol at position 4" in order to form a compound of the formula

5



in which R₁, R₂, R₃, R₄, R₅, R₆, R₇, m, n, A, B, C, D, E and F have the meanings given for formula (I); and

2) reacting the compound of the formula (III) with an amine of the formula HN(R₈)R₉,

10

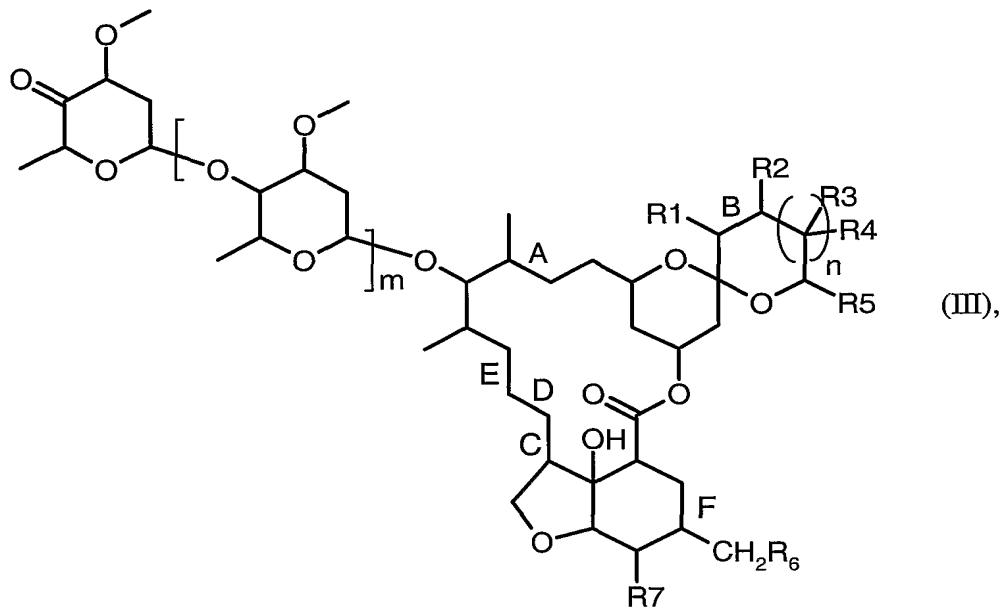
wherein R₈ and R₉ have the same meanings as given for formula (I), and which is known, in the presence of a reducing agent;

and, in each case, if desired, converting a compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, into a different compound of formula (I) or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, separating a mixture of E/Z isomers obtainable in accordance with the process and isolating the desired isomer, and/or converting a free compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, into a salt or converting a salt, obtainable in accordance with the process or by another

15

method, of a compound of formula (I) or of an E/Z isomer or tautomer thereof into the free compound of formula (I) or an E/Z isomer or tautomer thereof or into a different salt.

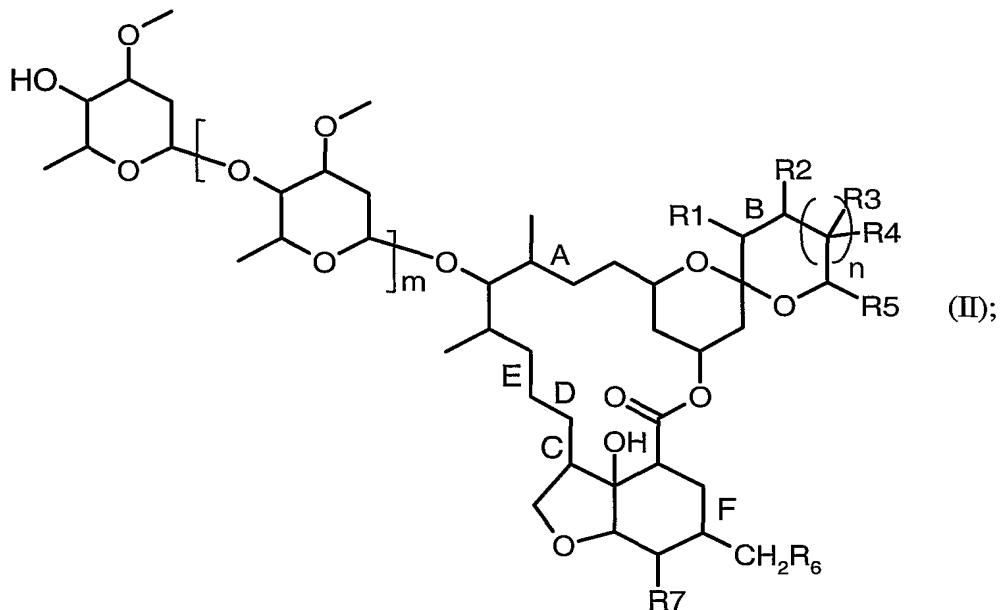
5 38. A method for the preparation of a compound of the formula



in which R₁, R₂, R₃, R₄, R₅, R₆, R₇, m, n, A, B, C, D, E and F have the meanings given for formula (III) of claim 37,

10 which process comprises

- 1) bringing a compound of the formula



wherein

R₁-R₇, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regioselectively oxidising the alcohol at position 4", maintaining said contact for a time sufficient for the oxidation reaction to occur and isolating and purifying the compound of formula (II).

- 10 39. A method according to anyone of claims 37 or 38 for making emamectin, comprising
adding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and
regioselectively oxidizes avermectin to 4"ketovermectin to a reaction mixture
comprising avermectin and incubating the reaction mixture under conditions that allow
the polypeptide to regioselectively oxidize avermectin to 4"ketovermectin.

15 40. The method of anyone of claims 37 to 39, wherein the reaction mixture further comprises
a ferredoxin protein.

41. The method of anyone of claims 37 to 40, wherein the reaction mixture further comprises
a ferredoxin reductase protein.

42. A formulation for making emamectin comprising a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4" keto-avermectin.

5

43. The formulation of claims 42 further comprising a ferredoxin protein.

44. The formulation of claim 42 or 43 further comprising a ferredoxin reductase protein.

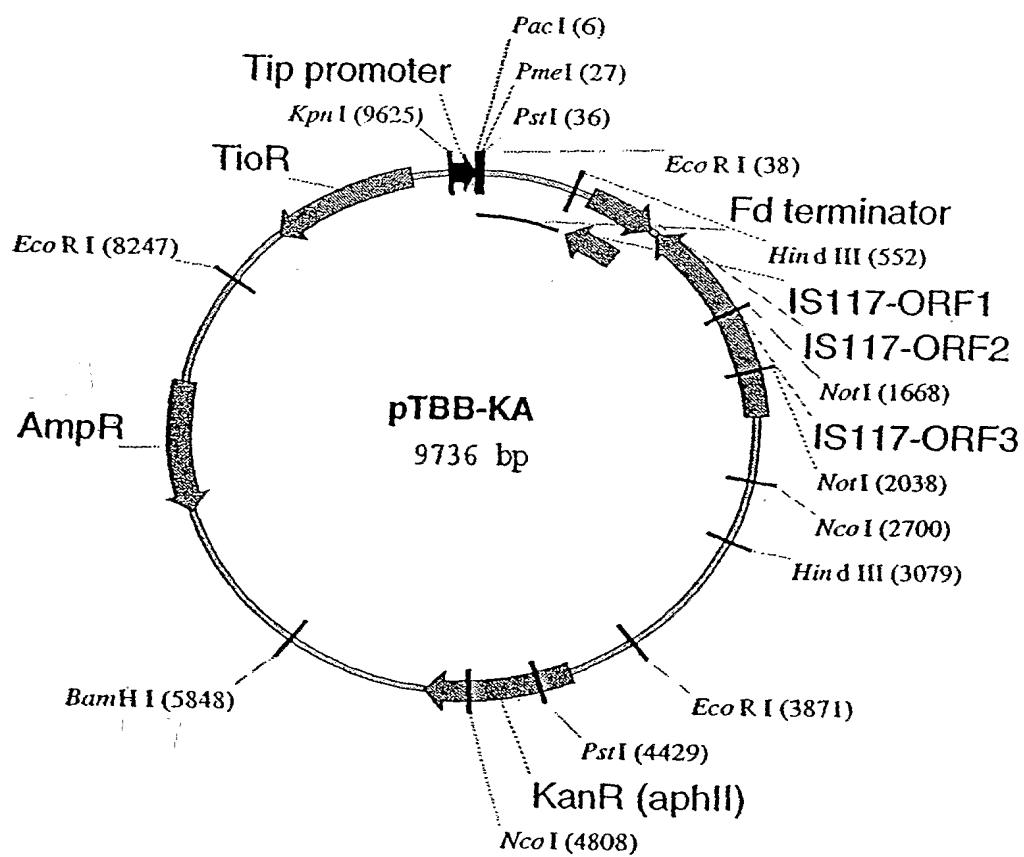


Figure 1

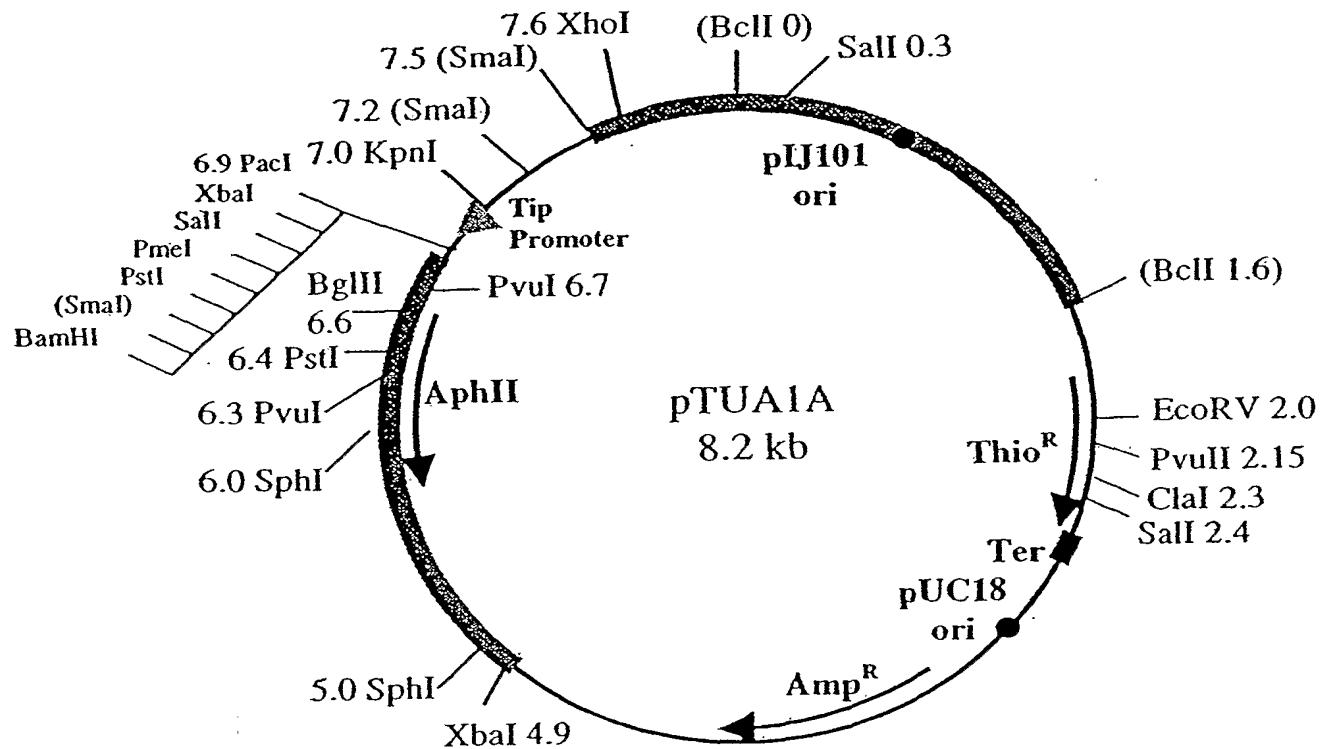


Figure 2

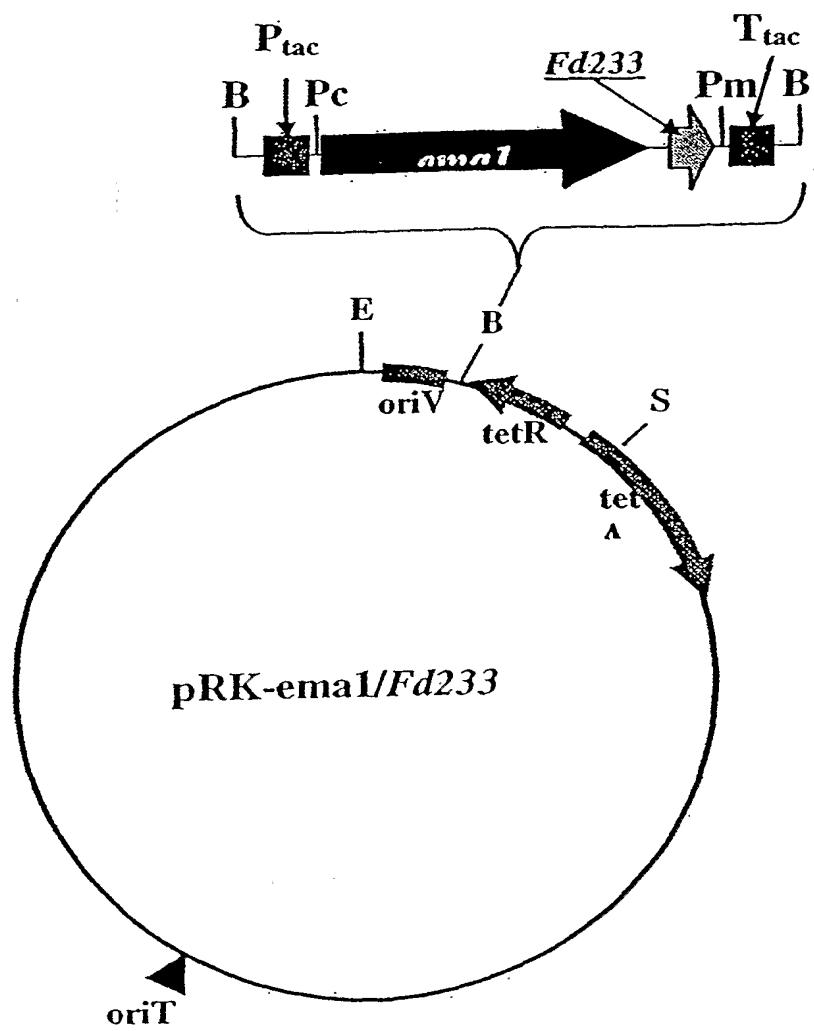


Figure 3

SEQUENCE LISTING

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<120> METHODS AND COMPOSITIONS FOR MAKING EMAMECTIN

<130> PB/5-60016A

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 35 40 45

Phe Met Asp Asp Ser Pro Val Trp Leu Val Thr Arg Phe Glu Glu Val
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Gln Ile Ala Asp Ala Leu Ala Arg Leu Pro Glu His Ala Glu Asp
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Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr
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 180 185 190

Arg Thr Trp Gly Ala Asp Leu Ile Ser Met Asp Pro Asp Arg Leu Gly
 195 200 205

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Arg Arg Glu Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr
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Ile Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
 260 265 270

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 340 345 350

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Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Thr His Tyr Pro
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425

430

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Pro	Leu	Ala	Pro	Ser	Ala	Glu	Glu	Asn	Pro	Leu	Thr	Arg	Leu	Met	Asp
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Met	Leu	Gly	Leu	Pro	Glu	His	Leu	Arg	Val	Tyr	Met	Leu	Gly	Ser	Ile
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Val Ile Cys Glu Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp		
180	185	190
Arg Lys Trp Gly Ala Asp Leu Ile Ser Met Asp Pro Asp Arg Leu Gly		
195	200	205
Ala Thr Phe Pro Ala Met Ile Glu His Ile His Glu Met Val Arg Glu		
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245	250	255
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Ser Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu		
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Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met		
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Asp Val Asp Leu Ala Gly Thr Arg Ile His Lys Gly Asp Ala Val Gln		
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Leu Leu Leu Val Ala Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro		
340	345	350
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His		
355	360	365
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala		
370	375	380
Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Ala His Tyr Pro		
385	390	395
Glu Met Ser Leu Gly Ile Glu Pro Glu Arg Leu Glu Arg Leu Pro Leu		
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Pro Gly Asn Trp Arg Leu Asn Ser Leu Pro Leu Arg Leu Gly		
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<210> 5
<211> 1413
<212> DNA
<213> Streptomyces rimosus

<400> 5

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cgcaccacgc	tccccctcta	cgtcgccctc	caccggggcg	agccgaacct	gatggaaaccg	180
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gaggtgatgc	gtgaccagcg	tttcgtcaac	aaccggaccc	tggtgccccgg	catcgccg	360
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<210> 6

<211> 470

<212> PRT

<213> Streptomyces rimosus

<400> 6

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20	25	30		
Ala Ala Ser Pro Asp Thr Thr Asp Arg Thr Thr Leu Pro Ser Tyr Val				
35	40	45		
Gly Leu His Pro Gly Glu Pro Asn Leu Met Glu Pro Glu Leu Leu Glu				
50	55	60		
Asn Pro Tyr Thr Gly Tyr Gly Thr Leu Arg Glu Gln Ala Pro Leu Val				
65	70	75	80	
Arg Ala Arg Phe Ile Asp Asp Ser Pro Ile Trp Leu Val Thr Arg Phe				
85	90	95		
Asp Val Val Arg Glu Val Met Arg Asp Gln Arg Phe Val Asn Asn Pro				
100	105	110		
Thr Leu Val Pro Gly Ile Gly Ala Asp Lys Asp Pro Arg Ala Arg Leu				
115	120	125		
Ile Glu Leu Phe Gly Ile Pro Glu Asp Leu Ala Pro Tyr Leu Thr Asp				
130	135	140		
Asn Ile Leu Thr Ser Asp Pro Pro Asp His Thr Arg Leu Arg Arg Leu				
145	150	155	160	
Val Ser Arg Ala Phe Thr Ala Arg Arg Ile Gln Asp Leu Arg Pro Arg				
165	170	175		
Val Glu Arg Ile Thr Asp Glu Leu Leu Glu Arg Leu Pro Asp His Ala				
180	185	190		
Glu Asp Gly Val Val Asp Leu Val Glu His Phe Ala Tyr Pro Leu Pro				
195	200	205		
Ile Thr Val Ile Cys Glu Leu Val Gly Ile Asp Glu Glu Asp Arg Ala				
210	215	220		
Leu Trp Arg Arg Phe Gly Ala Asp Leu Ala Ser Leu Asn Pro Lys Arg				

225	230	235	240												
Ile	Gly	Ala	Thr	Met	Pro	Glu	Met	Ile	Ser	His	Ile	His	Glu	Leu	Ile
245	250	255													
Asp	Glu	Arg	Arg	Ala	Ala	Leu	Arg	Asp	Asp	Leu	Leu	Ser	Gly	Leu	Ile
260	265	270													
Arg	Ala	Gln	Asp	Asp	Asp	Gly	Gly	Arg	Leu	Ser	Asp	Val	Glu	Met	Val
275	280	285													
Thr	Leu	Val	Leu	Thr	Leu	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His
290	295	300													
Leu	Ile	Ser	Asn	Gly	Thr	Leu	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Arg
305	310	315	320												
Arg	Leu	Ile	Asp	Glu	Asp	Pro	Ala	Leu	Leu	Pro	Arg	Ala	Val	His	Glu
325	330	335													
Leu	Met	Arg	Trp	Cys	Gly	Pro	Ile	Gln	Ala	Thr	Gln	Leu	Arg	Tyr	Ala
340	345	350													
Leu	Glu	Asp	Thr	Glu	Val	Ala	Gly	Val	Gln	Val	Arg	Gln	Gly	Glu	Ala
355	360	365													
Leu	Met	Phe	Ser	Leu	Val	Ala	Ala	Asn	His	Asp	Pro	Arg	His	Tyr	Thr
370	375	380													
Gly	Pro	Glu	Arg	Leu	Asp	Leu	Thr	Arg	Gln	Pro	Ala	Gly	Arg	Ala	Glu
385	390	395	400												
Asp	His	Val	Gly	Phe	Gly	His	Gly	Met	His	Tyr	Cys	Leu	Gly	Ala	Ser
405	410	415													
Leu	Ala	Arg	Gln	Glu	Ala	Glu	Val	Ala	Tyr	Gly	Lys	Leu	Leu	Thr	Arg
420	425	430													
Tyr	Pro	Asp	Leu	Ala	Leu	Ala	Leu	Thr	Pro	Glu	Gln	Leu	Glu	Asp	Gln
435	440	445													
Glu	Arg	Leu	Arg	Gln	Pro	Gly	Thr	Trp	Arg	Leu	Arg	Arg	Leu	Pro	Leu
450	455	460													
Arg	Leu	His	Ala	Gln	Ser										
465	470														

<210> 7
 <211> 1293
 <212> DNA
 <213> Streptomyces lydicus

<400> 7

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cagggccccc	tcgtgcgggg	gcggttcatg	gacgactccc	ccgtgtggtt	cgtgaccggc	180
ttcgaggagg	tccgcgaggt	cctgcgtgac	ccgcggttcc	ggaacaatcc	ggtctccgcg	240
gccccggccg	cggcccccga	ggacaccccg	ctgtcccgcc	tatggacat	gatgggtttc	300
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acccggctgc	gcccgcgtgt	ctcccgcc	ttcacccgc	ggaagatcac	cgatctgcgg	420
ccgcgcgtca	cacagatagc	cgacgagctg	ctggcccgcc	tgccggagca	cgccgaggac	480
ggcgtcgctg	acctgatcca	gcacttcgccc	tatccccctgc	cgatcaccgt	catctgcgaa	540
ctggtcggca	tccccggagga	ggaccggcccg	cagtggcgc	cctggggcgc	cgacctgtgc	600
tcgctgcagc	cggaccggat	gagccggtcc	ttcccgccgca	tgatcgacca	catccacgag	660
ctgatcgccg	cgcggcgccg	ggcgctcacc	gacgatctgc	tcagcgagct	gatccggacc	720
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gtcctggccg	gccacgagac	caccgcgcac	ctcatcgcc	acggcacccggc	ggccctgtc	840
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cacgagttga	tgcgtggtg	cggcccggtg	cacatgaccc	agctgcgcta	cggccggcag	960
gacgtcgagc	tggcgggcgt	ccggatccgc	acgggggacg	ccgtccagct	cattctggtg	1020
tccggcaacc	gcgaccggcg	ccactacacc	gaccccgacc	ggctggacct	gacccggcac	1080
cctgcccggcc	acgcggagaa	ccatgtgggg	ttcggccacg	gggcgcaacta	ctgtctgggc	1140
gccacgctcg	ccaagcagga	gggcgaggtc	gccctcgccg	ccctgtcgtcag	gcacttcccc	1200
gagctgtcgc	tggccgtcgc	gccggaggcc	ctggagcgc	caccggtacc	gggcagctgg	1260
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<210> 8
<211> 430
<212> PRT
<213> Streptomyces lydicus

<400> 8
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Ala Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg
35 40 45
Phe Met Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val
50 55 60
Arg Glu Val Leu Arg Asp Pro Arg Phe Arg Asn Asn Pro Val Ser Ala
65 70 75 80
Ala Pro Gly Ala Ala Pro Glu Asp Thr Pro Leu Ser Arg Leu Met Asp
85 90 95
Met Met Gly Phe Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile
100 105 110
Leu Asn Asn Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
115 120 125
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Thr
130 135 140
Gln Ile Ala Asp Glu Leu Ala Arg Leu Pro Glu His Ala Glu Asp
145 150 155 160
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr
165 170 175
Val Ile Cys Glu Leu Val Gly Ile Pro Glu Glu Asp Arg Pro Gln Trp
180 185 190
Arg Thr Trp Gly Ala Asp Leu Val Ser Leu Gln Pro Asp Arg Met Ser
195 200 205
Arg Ser Phe Pro Ala Met Ile Asp His Ile His Glu Leu Ile Ala Ala
210 215 220
Arg Arg Arg Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr
225 230 235 240
His Asp Asp Asp Gly Ser Arg Leu Ser Asp Val Glu Met Val Thr Met
245 250 255
Val Leu Thr Val Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
260 265 270
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu
275 280 285
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met
290 295 300
Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Tyr Ala Ala Glu

305	310	315	320
Asp Val Glu Leu Ala Gly Val Arg Ile Arg Thr Gly Asp Ala Val Gln			
325	330	335	
Leu Ile Leu Val Ser Ala Asn Arg Asp Pro Arg His Tyr Thr Asp Pro			
340	345	350	
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His			
355	360	365	
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala			
370	375	380	
Lys Gln Glu Gly Glu Val Ala Leu Gly Ala Leu Leu Arg His Phe Pro			
385	390	395	400
Glu Leu Ser Leu Ala Val Ala Pro Glu Ala Leu Glu Arg Thr Pro Val			
405	410	415	
Pro Gly Ser Trp Arg Leu Asn Ala Leu Pro Leu Arg Leu Arg			
420	425	430	

<210> 9
<211> 1299
<212> DNA
<213> Streptomyces sp.

<400> 9

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ttcgaggagg tccgccaagt cctgcgcgac cagcggttcg tgaacaatcc ggcgtcgccg	240
ctcctggcca gtcaggtcga ggagatgccc atggtaaagc tgctggagca gatgggcotc	300
cccgagcacc ttccgggtcta tctgctcggaa tcgatcctca acagtgcacgc ccccgatcac	360
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gtcttcgccc gtcacgagac caccggccac ctcataggca acggcacggc ggcgtcgctc	840
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cccgcaggcc acgcccggaaa ccacgtggc ttccggccacg ggatgcacta ctgcttggc	1140
gccgcgcgtcg ccaggcagga aggcgaatg gcttcggca aactgctcgc gcactacccg	1200
gacgtagcgc tggcgctcgaa accggaaagcc ctggagcggg tgccgatgcc cggcagttgg	1260
cggctgaatt cttggccgtc gcggttggcg aagcgctaa	1299

<210> 10
<211> 432
<212> PRT
<213> Streptomyces sp.

<400> 10
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Asn	Val	Met	Glu
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Gly	Gly	Tyr	Gly
Ala	Leu	Arg	Glu
35	40	45	
Phe	Met	Asp	Asp
Ser	Pro	Val	Trp
50	55	60	
Phe	Val	Thr	Arg
Arg	Gln	Asn	Asn
65	70	75	80
Leu	Leu	Gly	Ser
85	90	95	
Gln	Met	Gly	Leu
Leu	Pro	Glu	His
100	105	110	
Leu	Asn	Ser	Asp
115	120	125	
Arg	Ala	Phe	Thr
130	135	140	
Gln	Ile	Ala	Asp
145	150	155	160
Gly	Val	Val	Asp
165	170	175	
Val	Ile	Cys	Glu
180	185	190	
Arg	Ala	Trp	Gly
195	200	205	
Thr	Ser	Phe	Pro
210	215	220	
Arg	Arg	Gly	Ala
225	230	235	240
His	Asp	Asp	Asp
245	250	255	
Val	Phe	Ala	Leu
260	265	270	
Gly	Asn	Gly	Thr
275	280	285	
Leu	Lys	Asp	Asp
290	295	300	
Arg	Trp	Cys	Gly
305	310	315	320
Asp	Ile	Asp	Leu
325	330	335	
Leu	Ile	Leu	Val
340	345	350	
Asp	Arg	Leu	Asp
355	360	365	
Val	Gly	Phe	Gly
370	375	380	
Arg	Gln	Glu	Gly
385	390	395	400
Asp	Val	Ala	Leu
405	410	415	
Pro	Gly	Ser	Trp
420	425	430	
Arg	Leu	Asn	Ser
Leu	Pro	Leu	Arg
Ala	Lys	Leu	Arg

<210> 11
<211> 1293
<212> DNA
<213> Streptomyces chattanoogenensis

<400>	11					
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gcgcggggcg	cggccccccg	ggacaccccg	ctgtccggc	tatggacat	gatgggttcc	300
cccgagcacc	tgcgcgtcta	tctgctcgcc	tcgatcctca	acaacgacgc	ccccgaccac	360
acccggctgc	gccgcctgg	ctccccggcc	ttcaccgcgc	ggaagatcac	cgatctgcgg	420
ccgcgcgtca	cacagatagc	cgacgagctg	ctggccggc	tgccggagca	cgccgaggac	480
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gacgtcgagc	tggcgggcgt	ccggatccgc	acggggacg	ccgtccagct	catccctggtg	1020
tcggcgaacc	gcgaccccgcg	ccactacacc	gaccccgacc	gtctggacct	gacccggcac	1080
cccgccggc	acgcggagaa	ccatgtgggg	ttcggccacg	gggcgcacta	ctgtctggc	1140
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gagctgtcgc	tggcgtcgc	gcccggacgc	ctggagcga	caccgggtacc	gggcagctgg	1260
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<210> 12
<211> 430
<212> PRT
<213> Streptomyces chattanoogenensis

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35          40          45
Phe Met Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val
50          55          60
Arg Glu Val Leu Arg Asp Pro Arg Phe Arg Asn Asn Pro Val Ser Ala
65          70          75          80
Ala Pro Gly Ala Ala Pro Glu Asp Thr Pro Leu Ser Arg Leu Met Asp
85          90          95
Met Met Gly Phe Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile
100         105         110
Leu Asn Asn Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
115         120         125
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Thr

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130	135	140
Gln Ile Ala Asp Glu Leu	Leu Ala Arg Leu Pro	Glu His Ala Glu Asp
145	150	155
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro	Leu Pro Ile Thr	160
165	170	175
Val Ile Cys Glu Leu Val Gly Ile Pro	Glu Asp Arg Pro Gln Trp	
180	185	190
Arg Thr Trp Gly Ala Asp Leu Val Ser	Leu Gln Pro Asp Arg Met Ser	
195	200	205
Arg Ser Phe Pro Ala Met Ile Asp His Ile His	Glu Leu Ile Ala Ala	
210	215	220
Arg Arg Arg Ala Leu Thr Asp Asp Leu Leu Ser	Glu Leu Ile Arg Thr	
225	230	235
240		
His Asp Asp Asp Gly Ser Arg Leu Ser Asp	Val Glu Met Val Thr Met	
245	250	255
Val Leu Thr Val Val Leu Ala Gly His	Glu Thr Thr Ala His Leu Ile	
260	265	270
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp	Gln Leu Arg Leu	
275	280	285
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val	His Glu Leu Met	
290	295	300
Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Tyr	Ala Ala Glu	
305	310	315
320		
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325	330	335
Leu Ile Leu Val Ser Ala Asn Arg Asp Pro Arg His Tyr	Thr Asp Pro	
340	345	350
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His	Ala Glu Asn His	
355	360	365
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly	Ala Thr Leu Ala	
370	375	380
Lys Gln Glu Gly Glu Val Ala Leu Gly Ala	Leu Leu Arg His Phe Pro	
385	390	395
400		
Glu Leu Ser Leu Ala Val Ala Pro Asp Ala Leu Glu Arg	Thr Pro Val	
405	410	415
Pro Gly Ser Trp Arg Leu Asn Ala Leu Pro Leu Arg Leu	Gly	
420	425	430

<210> 13
<211> 1290
<212> DNA
<213> Streptomyces sp.

<400> 13

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gagcactacc gggtgtacct cgccaaacacc atcctcacca	tggacgcccc cgaccacacc	360
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cgggtggagg acatcgcgga cgatctgctg aggccggtcg	ccgagcacgc cgaggacggc	480
gtcgctcgacc tcatcaagca ctacgcctat ccgctgcccc	taacggtcat ctgcgaaactg	540

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gtcgagggtg	ccgggggtgc	ggtcaagcag	ggcgaagcgg	tgctggccat	gctgggtcg	1020
gcgaaccacg	accggcccca	cttcggcgc	cccgccccgg	tcgacccatc	ccgcccagccg	1080
gcggggccgg	ccgagaacca	cgtcggttc	ggccacggca	tgcaactact	cctggggcgc	1140
agcctggccc	gccaggaggg	cgaggtcg	ttcgggaacc	tgctcg	ctacccggac	1200
gtgtcgctgg	cggtggaaacc	ggacggcc	cagcgggtcc	cgctggcgg	caactggcgg	1260
ctggccgcac	tgccgg	tgccgtccg	gctgcgctga			1290

<210> 14
<211> 429
<212> PRT
<213> Streptomyces sp.

<400> 14

Met	Thr	Glu	Leu	Ala	Asp	Ser	Pro	Phe	Ser	Glu	His	Val	Gly	Lys	His
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Pro	Gly	Glu	Pro	Asn	Val	Met	Glu	Pro	Ala	Leu	Leu	Thr	Asp	Pro	Phe
															20
Thr	Gly	Tyr	Gly	Glu	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg
															35
Phe	Ala	Asp	Asp	Thr	Pro	Val	Trp	Phe	Ile	Thr	Arg	Phe	Glu	Glu	Ala
															50
Arg	Glu	Val	Leu	Arg	Asp	His	Arg	Phe	Ala	Asn	Ala	Pro	Ala	Phe	Ala
															65
Ala	Gly	Gly	Gly	Ser	Gly	Asp	Thr	Pro	Ser	Asn	Arg	Leu	Met	Glu	Ile
															85
Met	Gly	Leu	Pro	Glu	His	Tyr	Arg	Val	Tyr	Leu	Ala	Asn	Thr	Ile	Leu
															100
Thr	Met	Asp	Ala	Pro	Asp	His	Thr	Arg	Ile	Arg	Arg	Leu	Val	Ser	Arg
															115
Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu	Asp
															130
Ile	Ala	Asp	Asp	Leu	Leu	Arg	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp	Gly
															145
Val	Val	Asp	Leu	Ile	Lys	His	Tyr	Ala	Tyr	Pro	Leu	Pro	Ile	Thr	Val
															165
Ile	Cys	Glu	Leu	Val	Gly	Ile	Pro	Glu	Glu	Asp	Arg	Leu	Gln	Trp	Arg
															180
Asp	Trp	Gly	Ser	Ala	Phe	Val	Ser	Leu	Gln	Pro	Asp	Arg	Leu	Ser	Lys
															195
Ala	Phe	Pro	Ala	Met	Ile	Glu	His	Ile	His	Ala	Leu	Ile	Arg	Glu	Arg
															210
Arg	Ala	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Val	His
															225
Asp	Asp	Asp	Gly	Gly	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met	Val
															245
Leu	Thr	Leu	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile	Gly

	260		265		270										
Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	His	Leu	Leu
							275		280				285		
Lys	Ser	Asp	Pro	Glu	Leu	Leu	Pro	Arg	Ala	Val	His	Glu	Leu	Met	Arg
							290		295			300			
Trp	Cys	Gly	Pro	Val	Gln	Met	Thr	Gln	Leu	Arg	Tyr	Ala	Thr	Glu	Asp
							305		310			315		320	
Val	Glu	Val	Ala	Gly	Val	Gln	Val	Lys	Gln	Gly	Glu	Ala	Val	Leu	Ala
							325			330				335	
Met	Leu	Val	Ala	Ala	Asn	His	Asp	Pro	Arg	His	Phe	Ala	Asp	Pro	Ala
							340			345				350	
Arg	Leu	Asp	Leu	Thr	Arg	Gln	Pro	Ala	Gly	Arg	Ala	Glu	Asn	His	Val
							355			360			365		
Gly	Phe	Gly	His	Gly	Met	His	Tyr	Cys	Leu	Gly	Ala	Ser	Leu	Ala	Arg
							370			375			380		
Gln	Glu	Gly	Glu	Val	Ala	Phe	Gly	Asn	Leu	Leu	Ala	His	Tyr	Pro	Asp
							385			390			395		400
Val	Ser	Leu	Ala	Val	Glu	Pro	Asp	Ala	Leu	Gln	Arg	Val	Pro	Leu	Pro
							405			410				415	
Gly	Asn	Trp	Arg	Leu	Ala	Ala	Leu	Pro	Val	Arg	Leu	Arg			
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<210> 15
<211> 1428
<212> DNA
<213> Streptomyces albofaciens

<400> 15
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cccgccacca ccgaccgcac cacgctccccc tcctacgtcg gcctccaccc gggcgagccg 180
aacctgtatgg aaccggagat gctggacaac ccgtacacccg gctacggcac gctgcgcgag 240
caggcgcgcg tcgtccgcgc ccggttcatc gacgactcgc ccatctggct ggtgacccgc 300
ttcgacgtgg tgcgcgaggt gatgcgcgac cagcggttcg tcaacaaccc gaccctggtg 360
cccgccatcg gtgcggacca ggaccgcgc gcccggctga tcgagctgtt cggcatcccc 420
gaggacctgg ccccgtacct caccgacacc atcctcacca gcgacccgccc ggaccacacc 480
cggtctgcgc gcttgttctc ccgtgccttc accgcacgcc gtagccagga cctgccccgc 540
cgcgctcgagc ggtatcaccga cgagctgtcg gcgccggctgc cggaccatgc cgaggacggc 600
gtcgctcgacc tcgtcgagca ttgcgcctac ccgtgcctca tcacggtcat ctgcgaactg 660
gtcgccatcg acgaggagga ccgggcgcgt tggcggcggt tcggccgcga cctgcctcg 720
ctgaacccca agcgcatcg cgccaccatg ccggagatga tcgcccacat ccacgagggtg 780
atcgacgagc ggcgtgcggg a cctgcgggac gacctgtca gcgggctcat ccggccgcag 840
gacgacgacg gcccggcggt gagcgacgtc gagatggtca cgctggtgct gaccctggtg 900
ctggccggtc acgagaccac cgccaccatc atcagcaacg gcaccctcgc cctgctcacc 960
caccggacc agcggcggt gatcgacgag gaccggcgca tgctgccccg cgcggtccac 1020
gagctgtatgc gctgggtgcgg gccgatccag gccaccccgac tgccgtacgc catggaggac 1080
accgagggtgg ccggtgtcca ggtccggccag ggcgaggcccc tgatgttca gctcgctcg 1140
gccaaccacg acccgcccca ctacaccggc ccggagccgc tcgacccgtac gcggcagccg 1200
gcggcccgcg ccgaggacca cgtccggctc gggcacggga tgcaactactg cctgggtgccc 1260
tcactggccc ggcaggaggc cgaggtggcg tacggcaagc tgctcaccctg ctacccggac 1320
ctggcgctcg cgctcaccctg ggaacagctg gaggaccagg aacgcctgcg gcagccggc 1380
acctggccgc tgcgacggct gccgctgagg ttgcacgcggg agagctga 1428

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<210> 16
<211> 475
<212> PRT
<213> Streptomyces albofaciens

<400> 16
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Thr Ala Ser Pro Ser Thr Ala Ala Pro Ala Thr Thr Pro Ser Ala
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Ala Ala Ser Pro Asp Thr Thr Ser Pro Ala Thr Thr Asp Arg Thr Thr
35 40 45
Leu Pro Ser Tyr Val Gly Leu His Pro Gly Glu Pro Asn Leu Met Glu
50 55 60
Pro Glu Leu Leu Asp Asn Pro Tyr Thr Gly Tyr Gly Thr Leu Arg Glu
65 70 75 80
Gln Ala Pro Leu Val Arg Ala Arg Phe Ile Asp Asp Ser Pro Ile Trp
85 90 95
Leu Val Thr Arg Phe Asp Val Val Arg Glu Val Met Arg Asp Gln Arg
100 105 110
Phe Val Asn Asn Pro Thr Leu Val Pro Gly Ile Gly Ala Asp Gln Asp
115 120 125
Pro Arg Ala Arg Leu Ile Glu Leu Phe Gly Ile Pro Glu Asp Leu Ala
130 135 140
Pro Tyr Leu Thr Asp Thr Ile Leu Thr Ser Asp Pro Pro Asp His Thr
145 150 155 160
Arg Leu Arg Arg Leu Val Ser Arg Ala Phe Thr Ala Arg Arg Ile Gln
165 170 175
Asp Leu Arg Pro Arg Val Glu Arg Ile Thr Asp Glu Leu Leu Ala Arg
180 185 190
Leu Pro Asp His Ala Glu Asp Gly Val Val Asp Leu Val Glu His Phe
195 200 205
Ala Tyr Pro Leu Pro Ile Thr Val Ile Cys Glu Leu Val Gly Ile Asp
210 215 220
Glu Glu Asp Arg Ala Leu Trp Arg Arg Phe Gly Ala Asp Leu Ala Ser
225 230 235 240
Leu Asn Pro Lys Arg Ile Gly Ala Thr Met Pro Glu Met Ile Ala His
245 250 255
Ile His Glu Val Ile Asp Glu Arg Arg Ala Asp Leu Arg Asp Asp Leu
260 265 270
Leu Ser Gly Leu Ile Arg Ala Gln Asp Asp Asp Gly Gly Arg Leu Ser
275 280 285
Asp Val Glu Met Val Thr Leu Val Leu Thr Leu Val Leu Ala Gly His
290 295 300
Glu Thr Thr Ala His Leu Ile Ser Asn Gly Thr Leu Ala Leu Leu Thr
305 310 315 320
His Pro Asp Gln Arg Arg Leu Ile Asp Glu Asp Pro Ala Leu Leu Pro
325 330 335
Arg Ala Val His Glu Leu Met Arg Trp Cys Gly Pro Ile Gln Ala Thr
340 345 350
Gln Leu Arg Tyr Ala Met Glu Asp Thr Glu Val Ala Gly Val Gln Val
355 360 365
Arg Gln Gly Glu Ala Leu Met Phe Ser Leu Val Ala Ala Asn His Asp

370	375	380
Pro Arg His Tyr Thr Gly Pro Glu Arg Leu Asp Leu Thr Arg Gln Pro		
385	390	395
Ala Gly Arg Ala Glu Asp His Val Gly Phe Gly His Gly Met His Tyr		
	405	410
Cys Leu Gly Ala Ser Leu Ala Arg Gln Glu Ala Glu Val Ala Tyr Gly		
	420	425
Lys Leu Leu Thr Arg Tyr Pro Asp Leu Ala Leu Ala Leu Thr Pro Glu		
	435	440
Gln Leu Glu Asp Gln Glu Arg Leu Arg Gln Pro Gly Thr Trp Arg Leu		
	450	455
Arg Arg Leu Pro Leu Arg Leu His Ala Glu Ser		
	465	470
		475

<210> 17

<211> 1293

<212> DNA

<213> Streptomyces platensis

<400> 17

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cagggcccg tcgtccgcgg ccgcttcgtg gacgactcac ccgtctggct ggtacgcga	180
ttcgaggagg tccgccaagt cctgcgcgac cagcggttcg tgaacaaccc ggccggcccc	240
tccctgggcc acgcggccga ggacaacccg ctcaccaggc tgatggacat gctggccctc	300

cccgagcacc tccgcccccta cctcctcgga tcgattctca attacgacgc ccccgaccac	360
accggcgtgc gcccctgggt gtcgcgggcc ttccaccgccc gcaagatcac cgacctgcgg	420
ccgcgggtcg agcagatcgc cgacgcctg ctggcccgcc tgcccgagca cgccgaggac	480
ggcgtcgtcg atctcatccg gcacttcgccc taccgcgtgc cgatcacgt catctgcgaa	540
ctggtcggca taccgcaga ggaccgcgg cagtggcgga cgtggggcgc cgacctcgctc	600
tcgatggagc cggacggctcaccgcgtccg ttcccggca tgatcgagca catccaccgg	660
atggtccggg agccgcgcgg cgcgctcacc ggcgatctgc tcagcggagct gatccgtgcc	720
catgacgacg acggcggccg gtcagcgcac gtcgagatgg tcaccttgat ctcacgctc	780
gtcctcgccg gtacacgagac caccgcgtcac ctcacatcgaca acggcacggc ggccgtcgtc	840
accaccccg accaactcgc cctgctccag gacgaccggg ccctgctccc ccgtgcgtc	900
cacgagctga tgcgctgggt cggccgggtg cagatgaccc agctgcgtta cgccgcggcc	960
gacgtcgtacc tggccggcac cacgatccac cggggcgtacg ccgtccaact catcctggtg	1020
tcggcgaact tcgaccccccgc ccactacacc gaccccgacc gcctcgatct gacccgcac	1080
cccgccggac atgcggagaa ccatgtgggt ttcggccatg gggcgacta ctgcctgggc	1140
gccacactcg ccaagcagga gggcgaagtc gccttcggca aactgctcgc gcactacccg	1200
gagatggcgt tggcgctcgc accggagcgc ctggagcggg cgcggctggcc gggcaactgg	1260
cggctgaacg cgctgcgggt gcgggttgggg tga	1293

<210> 18

<211> 430

<212> PRT

<213> Streptomyces platensis

<400> 18

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		15

Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Thr Asp Pro Phe
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 Thr Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg
 35 40 45
 Phe Val Asp Asp Ser Pro Val Trp Leu Val Thr Arg Phe Glu Glu Val
 50 55 60
 Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ala Pro
 65 70 75 80
 Ser Leu Gly His Ala Ala Glu Asp Asn Pro Leu Thr Arg Leu Met Asp
 85 90 95
 Met Leu Gly Leu Pro Glu His Leu Arg Pro Tyr Leu Leu Gly Ser Ile
 100 105 110
 Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
 115 120 125
 Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu
 130 135 140
 Gln Ile Ala Asp Ala Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp
 145 150 155 160
 Gly Val Val Asp Leu Ile Arg His Phe Ala Tyr Pro Leu Pro Ile Thr
 165 170 175
 Val Ile Cys Glu Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp
 180 185 190
 Arg Thr Trp Gly Ala Asp Leu Val Ser Met Glu Pro Asp Arg Leu Thr
 195 200 205
 Ala Ser Phe Pro Pro Met Ile Glu His Ile His Arg Met Val Arg Glu
 210 215 220
 Arg Arg Gly Ala Leu Thr Gly Asp Leu Leu Ser Glu Leu Ile Arg Ala
 225 230 235 240
 His Asp Asp Asp Gly Gly Arg Leu Ser Asp Val Glu Met Val Thr Leu
 245 250 255
 Ile Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
 260 265 270
 Ser Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu
 275 280 285
 Leu Gln Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met
 290 295 300
 Arg Trp Cys Gly Pro Val Gln Met Thr Gln Leu Arg Tyr Ala Ala Ala
 305 310 315 320
 Asp Val Asp Leu Ala Gly Thr Thr Ile His Arg Gly Asp Ala Val Gln
 325 330 335
 Leu Ile Leu Val Ser Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro
 340 345 350
 Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His
 355 360 365
 Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala
 370 375 380
 Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Ala His Tyr Pro
 385 390 395 400
 Glu Met Ala Leu Gly Val Ala Pro Glu Arg Leu Glu Arg Thr Pro Leu
 405 410 415
 Pro Gly Asn Trp Arg Leu Asn Ala Leu Pro Val Arg Leu Gly
 420 425 430

<210> 19
<211> 1293
<212> DNA
<213> Streptomyces kasugaensis

<400> 19

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cagggcccg	tctgtcgaaa	gccccgtcatg	gacgactccc	ccgtgtggtt	cgtgacccgc	180
ttcgaggagg	tccgcgaggt	cctgcgtgac	ccgcgggttc	ggaacaatcc	ggtctccgcg	240
gcgcggggcg	cgccccccga	ggacaccccg	ctgtcccccgc	tgtatggacat	gatgggtttc	300
cccgagcacc	tgcgctcta	tctgtcgcc	tcgatcctca	acaacgacgc	ccccgaccac	360
accggcgtgc	gccgcctgg	ctcccggtcc	ttcaccgcgc	ggaagatcac	cgatctgcgg	420
ccgcgcgtca	cacagatagc	cgacgagctg	ctggcccgcc	tgccggagca	cgccgaggac	480
ggcgtcgtcg	acctgatcca	gcacttcgccc	tatccccctgc	cgatcaccgt	catctgcgaa	540
ctggtcggca	tccccgagga	ggaccgcgg	cagtggcgca	cctggggcg	cgacctggc	600
tcgctgcagc	cggaccggat	gagccgggtcc	ttcccggtca	tgtatcgacca	catccacgag	660
ctgatcgcgg	cgcgccgcgg	ggcgctcacc	gacgatctgc	tcagcgagct	gatccggacc	720
catgacgacg	acggcagccg	gctcagcgcac	gtcgagatgg	tcaccatgg	cctcaccgtc	780
gtcctggccg	gccacgagac	caccgcgcac	ctcatcgca	acggcacggc	ggccctgctc	840
accaccccg	accagctgcg	gctgctcaag	gacgacccgg	cgctgctgccc	gcgcgcgggt	900
cacgagttga	tgcgctgg	cgcccggtg	cacatgaccc	agctgcgcta	cgccgcccgg	960
gacgtcgagc	tggcgccgt	ccgatccgc	acgggggacg	ccgtccagct	catcctgg	1020
tcggcgaacc	gcgaccccg	ccactacacc	gaccccgacc	ggctggaccc	gacccggcac	1080
cctgcccggcc	acgcccggaa	ccatgtgggg	ttcggccacg	ggcgccacta	ctgtctggc	1140
gccacgctcg	ccaaggcagga	gggcgagggtc	gccctcgccg	ccctgctcag	gcacttcccc	1200
gagctgtcg	tggccgtcgc	gccggaggcc	ctggagcgc	cacgggtacc	gggcagctgg	1260
cgctgaacg	cgctgcccgt	gcgtctgcgc	tga			1293

<210> 20
<211> 430
<212> PRT
<213> Streptomyces kasugaensis

<400> 20

Met	Ser	Ala	Ser	Pro	Ser	Asn	Thr	Phe	Thr	Glu	His	Val	Gly	Lys	His
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Pro	Gly	Glu	Pro	Asn	Val	Met	Asp	Pro	Ala	Leu	Ile	Gly	Asp	Pro	Phe
								20		25				30	
Ala	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg
							35		40				45		
Phe	Met	Asp	Asp	Ser	Pro	Val	Trp	Phe	Val	Thr	Arg	Phe	Glu	Glu	Val
							50		55				60		
Arg	Glu	Val	Leu	Arg	Asp	Pro	Arg	Phe	Arg	Asn	Asn	Pro	Val	Ser	Ala
							65		70				75		80
Ala	Pro	Gly	Ala	Ala	Pro	Glu	Asp	Thr	Pro	Leu	Ser	Arg	Leu	Met	Asp
							85		90				95		
Met	Met	Gly	Phe	Pro	Glu	His	Leu	Arg	Val	Tyr	Leu	Leu	Gly	Ser	Ile
							100		105				110		
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
							115		120				125		
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Thr
							130		135				140		

Gln Ile Ala Asp Glu Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp
 145 150 155 160
 Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr
 165 170 175
 Val Ile Cys Glu Leu Val Gly Ile Pro Glu Glu Asp Arg Pro Gln Trp
 180 185 190
 Arg Thr Trp Gly Ala Asp Leu Val Ser Leu Gln Pro Asp Arg Met Ser
 195 200 205
 Arg Ser Phe Pro Ala Met Ile Asp His Ile His Glu Leu Ile Ala Ala
 210 215 220
 Arg Arg Arg Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr
 225 230 235 240
 His Asp Asp Asp Gly Ser Arg Leu Ser Asp Val Glu Met Val Thr Met
 245 250 255
 Val Leu Thr Val Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
 260 265 270
 Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu
 275 280 285
 Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met
 290 295 300
 Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Tyr Ala Ala Glu
 305 310 315 320
 Asp Val Glu Leu Ala Gly Val Arg Ile Arg Thr Gly Asp Ala Val Gln
 325 330 335
 Leu Ile Leu Val Ser Ala Asn Arg Asp Pro Arg His Tyr Thr Asp Pro
 340 345 350
 Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His
 355 360 365
 Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala
 370 375 380
 Lys Gln Glu Gly Glu Val Ala Leu Gly Ala Leu Leu Arg His Phe Pro
 385 390 395 400
 Glu Leu Ser Leu Ala Val Ala Pro Glu Ala Leu Glu Arg Thr Pro Val
 405 410 415
 Pro Gly Ser Trp Arg Leu Asn Ala Leu Pro Leu Arg Leu Arg
 420 425 430

<210> 21
 <211> 1428
 <212> DNA
 <213> Streptomyces rimosus

<400> 21

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cccgccacca	ccgaccgcac	cacgctcccc	tcc tacgtcg	gcctccaccc	gggcgagccg	180
aacctgatgg	aaccggagct	gctggacaac	ccgtacacccg	gctacggcac	gctgcgcgag	240
caggccccgc	tcgtccgtgc	ccggttcatc	gacgactcgc	ccatctggct	ggtgaccgc	300
ttcgacgtgg	tgcgcgaggt	gatgcgcgac	cagcggttcg	tcaacaaccc	gaccctggtg	360
cccgccatcg	gtgcggacaa	ggacccgcgc	gcccggtctga	tcgagctgtt	cggcatcccc	420
gaggacctga	ccccgtacct	cgccgacacc	atccctcacca	gcgacccgccc	ggaccacacc	480
cggtcgcc	gcctggtctc	ccgtgccttc	accgcgcgc	gcatccagga	cctgcggccg	540
cgcgtcgagc	agatcaccga	cgcgctgctg	gagcgactgc	cgaggacatgc	cgaggacggc	600

gtcgtcgacc	tcgtcgagca	cttcgcctac	ccgctgcca	tcacggcat	ctgcgagctg	660
gtcggcatcg	acgaggagga	ccggacgctg	tggcggcggt	tcggcgccga	cctcgcccta	720
ctgaacccca	agcgcatcg	cgccaccatg	ccggagatga	tcgcgcacat	ccacgaggtg	780
atcgacgagc	ggcgcgcggc	cctgcgggac	gacctgctca	gcgggctcat	ccgggcgcag	840
gacgacgacg	gcggccggct	gagcgacgtc	gagatggtca	ccctggctct	gaccctggtg	900
ctggccggtc	acgagaccac	cgcccacctc	atcagcaacg	gcacccctcg	cctgctcacc	960
cacccccgacc	agcggccggct	gatcgacgag	gaccggcac	tgctgcccgc	cgcggtccac	1020
gagctgatgc	gctgggtgcgg	gccgatccag	gccacccagc	tgcggtacgc	catggaggac	1080
accgaggtcg	ccgggtgtcca	ggtccgcccag	ggcgaggccc	tgatgttcag	cctcgccgcg	1140
gccaaccacg	accggcgcca	ctacaccggg	ccggagcggc	tcgacactgac	gcggcagccg	1200
gccggccgcg	ccgaggacca	cgtcggttc	gggcacggga	tgcaactactg	cctgggtgcc	1260
tcactcgccc	ggcaggaggc	cgaggtggcc	tacggaaagc	tgctcacccg	ctacccggac	1320
ctggagctcg	ctctcacacc	ggaacagctg	gaggaccagg	aacgcctgcg	gcagcccgcc	1380
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<210> 22

<211> 475

<212> PRT

<213> Streptomyces rimosus

<400> 22

Met	Thr	Thr	Ser	Pro	Thr	Glu	Ser	Arg	Ala	Ala	Thr	Pro	Thr	Gly	Ser
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					20				25						30
Ala	Thr	Ser	Ser	Asp	Thr	Thr	Tyr	Pro	Ala	Thr	Thr	Asp	Arg	Thr	Thr
					35				40						45
Leu	Pro	Ser	Tyr	Val	Gly	Leu	His	Pro	Gly	Glu	Pro	Asn	Leu	Met	Glu
				50			55			60					
Pro	Glu	Leu	Leu	Asp	Asn	Pro	Tyr	Thr	Gly	Tyr	Gly	Thr	Leu	Arg	Glu
				65			70			75					80
Gln	Ala	Pro	Leu	Val	Arg	Ala	Arg	Phe	Ile	Asp	Asp	Ser	Pro	Ile	Trp
				85			90								95
Leu	Val	Thr	Arg	Phe	Asp	Val	Val	Arg	Glu	Val	Met	Arg	Asp	Gln	Arg
				100			105								110
Phe	Val	Asn	Asn	Pro	Thr	Leu	Val	Pro	Gly	Ile	Gly	Ala	Asp	Lys	Asp
				115			120								125
Pro	Arg	Ala	Arg	Leu	Ile	Glu	Leu	Phe	Gly	Ile	Pro	Glu	Asp	Leu	Thr
				130			135								140
Pro	Tyr	Leu	Ala	Asp	Thr	Ile	Leu	Thr	Ser	Asp	Pro	Pro	Asp	His	Thr
				145			150								160
Arg	Leu	Arg	Arg	Leu	Val	Ser	Arg	Ala	Phe	Thr	Ala	Arg	Arg	Ile	Gln
				165			170								175
Asp	Leu	Arg	Pro	Arg	Val	Glu	Gln	Ile	Thr	Asp	Ala	Leu	Glu	Arg	
				180			185								190
Leu	Pro	Asp	His	Ala	Glu	Asp	Gly	Val	Val	Asp	Leu	Val	Glu	His	Phe
				195			200								205
Ala	Tyr	Pro	Leu	Pro	Ile	Thr	Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Asp
				210			215								220
Glu	Glu	Asp	Arg	Thr	Leu	Trp	Arg	Arg	Phe	Gly	Ala	Asp	Leu	Ala	Ser
				225			230								240
Leu	Asn	Pro	Lys	Arg	Ile	Gly	Ala	Thr	Met	Pro	Glu	Met	Ile	Ala	His
				245			250								255

Ile	His	Glu	Val	Ile	Asp	Glu	Arg	Arg	Ala	Ala	Leu	Arg	Asp	Asp	Leu
				260		265						270			
Leu	Ser	Gly	Leu	Ile	Arg	Ala	Gln	Asp	Asp	Asp	Gly	Gly	Arg	Leu	Ser
				275		280					285				
Asp	Val	Glu	Met	Val	Thr	Leu	Val	Leu	Thr	Leu	Val	Leu	Ala	Gly	His
				290		295			300						
Glu	Thr	Thr	Ala	His	Leu	Ile	Ser	Asn	Gly	Thr	Leu	Ala	Leu	Leu	Thr
				305		310		315			320				
His	Pro	Asp	Gln	Arg	Arg	Leu	Ile	Asp	Glu	Asp	Pro	Ala	Leu	Leu	Pro
				325		330		335				335			
Arg	Ala	Val	His	Glu	Leu	Met	Arg	Trp	Cys	Gly	Pro	Ile	Gln	Ala	Thr
				340		345			345		350				
Gln	Leu	Arg	Tyr	Ala	Met	Glu	Asp	Thr	Glu	Val	Ala	Gly	Val	Gln	Val
				355		360		365							
Arg	Gln	Gly	Glu	Ala	Leu	Met	Phe	Ser	Leu	Val	Ala	Ala	Asn	His	Asp
				370		375		380							
Pro	Arg	His	Tyr	Thr	Gly	Pro	Glu	Arg	Leu	Asp	Leu	Thr	Arg	Gln	Pro
				385		390		395			400				
Ala	Gly	Arg	Ala	Glu	Asp	His	Val	Gly	Phe	Gly	His	Gly	Met	His	Tyr
				405		410		415							
Cys	Leu	Gly	Ala	Ser	Leu	Ala	Arg	Gln	Glu	Ala	Glu	Val	Ala	Tyr	Gly
				420		425		430							
Lys	Leu	Leu	Thr	Arg	Tyr	Pro	Asp	Leu	Glu	Leu	Ala	Leu	Thr	Pro	Glu
				435		440		445							
Gln	Leu	Glu	Asp	Gln	Glu	Arg	Leu	Arg	Gln	Pro	Gly	Thr	Trp	Arg	Leu
				450		455		460							
Arg	Arg	Leu	Pro	Leu	Lys	Leu	His	Ala	Arg	Ser					
				465		470		475							

<210> 23

<211> 1293

<212> DNA

<213> Streptomyces tubercidicus

<400> 23

atgtcgccat	tatccaactc	cccgctcgcc	gcacatgtcg	ggaaacaccc	tggcgagccg	60
aatgtatgg	accggcgct	gatcaccgac	ccgttcggcg	gctacggcgc	actgcgcgag	120
caaggcccg	tcgtacgggg	ccggttcatg	gacgactcgc	ccgtctggct	ggtgacgcgc	180
ttcgaagagg	tccgccaagt	cctgcgcgat	cagcggttcg	tgaacaaccc	ggccgcacccg	240
tccctgggac	gctcgatcga	cgaaagcccc	gcccgtcagac	ttttggaaat	gttggggttg	300
cccgaccatt	tccggccgta	tctgctcggg	tcgatcctca	actacgacgc	acccgaccac	360
acccggctcc	gccgacttgtt	ctcgcgccgc	ttcacggcac	gcaagatcac	cgacctgcgg	420
ccgcgggtcg	agcagatcac	cgacgacctg	ctgacccggc	ttcccgagca	cgccgaggac	480
ggtgtggtcg	acctcatcca	gcacttcgccc	tacccctgc	cgatcaccgt	gatctgcgaa	540

ctgggtcgcc	tccggcgaagc	ggaccgccc	caatggcgga	agtggggagc	cgatctcgtc	600
tcgctggagc	cggggcgct	gagcaccgc	ttcccgccga	tggtcgagca	catccatgag	660
ctgatccgc	agcggcgccg	cgcgctcacc	gacgatctgc	ttagcgagct	gatccgcacc	720
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gtcctggccg	gccacgagac	caccgcccac	ctcataggca	acggcacggc	ggcgctgctc	840
acccaccccg	accagctgcg	cctactcaag	gacgatccgg	cgctgctgcc	gcccgcgcgtc	900
cacgagctga	tgcgctggtg	cgggccgggt	cacatgaccc	agctgcggtt	cgcgccgag	960
gacgtcgagg	tcgcccggac	accgatccac	aagggcgacg	ccgtacaact	catcctggta	1020

tcggcgaact	tcgacccccc	ccactacacc	gaccccgacc	gtctcgacct	gacccgcccac	1080
cccgccgccc	acggcgagaa	ccatgtggc	tgcggccacg	aatgcacta	ctgcctgggt	1140
gccaccctcg	ccaaacacagga	aggcgaagtc	gccttctccc	gcctttcac	gcactacccg	1200
gaactgtccc	tggcgctcgc	ggcggaccag	ctggcgcgaa	cacaggtacc	cggcagctgg	1260
cggctggaca	ccctgcccct	gcgactgggg	tga			1293

<210> 24
<211> 430
<212> PRT
<213> Streptomyces tubercidicus

<400> 24						
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Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Thr Asp Pro Phe						
20	25	30				
Gly Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg						
35	40	45				
Phe Met Asp Asp Ser Pro Val Trp Leu Val Thr Arg Phe Glu Glu Val						
50	55	60				
Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ala Pro						
65	70	75	80			
Ser Leu Gly Arg Ser Ile Asp Glu Ser Pro Ala Val Arg Leu Leu Glu						
85	90	95				
Met Leu Gly Leu Pro Asp His Phe Arg Pro Tyr Leu Leu Gly Ser Ile						
100	105	110				
Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser						
115	120	125				
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu						
130	135	140				
Gln Ile Thr Asp Asp Leu Leu Thr Arg Leu Pro Glu His Ala Glu Asp						
145	150	155	160			
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr						
165	170	175				
Val Ile Cys Glu Leu Val Gly Ile Ala Glu Ala Asp Arg Pro Gln Trp						
180	185	190				
Arg Lys Trp Gly Ala Asp Leu Val Ser Leu Glu Pro Gly Arg Leu Ser						
195	200	205				
Thr Ala Phe Pro Ala Met Val Glu His Ile His Glu Leu Ile Arg Glu						
210	215	220				
Arg Arg Gly Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr						
225	230	235	240			
His Asp Asp Asp Gly Gly Arg Leu Ser Asp Ile Glu Met Val Thr Met						
245	250	255				
Ile Leu Thr Ile Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile						
260	265	270				
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu						
275	280	285				
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met						
290	295	300				
Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Phe Ala Ser Glu						
305	310	315	320			
Asp Val Glu Val Ala Gly Thr Pro Ile His Lys Gly Asp Ala Val Gln						

	325	330	335
Leu Ile Leu Val Ser Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro			
340	345	350	
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His			
355	360	365	
Val Gly Phe Gly His Gly Met His Tyr Cys Leu Gly Ala Thr Leu Ala			
370	375	380	
Lys Gln Glu Gly Glu Val Ala Phe Ser Arg Leu Phe Thr His Tyr Pro			
385	390	395	400
Glu Leu Ser Leu Gly Val Ala Ala Asp Gln Leu Ala Arg Thr Gln Val			
405	410	415	
Pro Gly Ser Trp Arg Leu Asp Thr Leu Pro Leu Arg Leu Gly			
420	425	430	

<210> 25

<211> 1293

<212> DNA

<213> Streptomyces platensis

<400> 25

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caaggggccgg tcgtacgggg ccggttcatg gacgactcac ccgtctggct cgtacgcgc	180
ttcgaggaag tccgccaagt cctgcgcac cagcggttcc tgaacgatcc gacggcccc	240
tccctggggc gtcattcga cgacagcccc acggccaggc tgctggagat gatgggactg	300
cccgagcatt tccggccgta tctgctcggc tcgattctga acaacgacgc ccccgaccac	360
accggctgc gccgtctggc ttacacggcac gcaagatcac cgacactgcgg	420
ccgcgggtcg agcagatcgc cgacgagctg ctgacccggc ttcccgagta cgccgaggac	480
ggcgtgtcg acctcatcaa gcacttcgcc tacccccgtc cgatcgccgt catctgcgaa	540
ctggtcggca tagccgaagc ggatcgccg cagtggcga agtggggtgc cgacactcgtc	600
tcgctgcagc cggaccggct cagcaccccg ttcccgcgta tgatcgagca catccatgag	660
ctgatccgcg agcggcgccg ggcgtcacg gacgatctgc tcagcgagct gatccgtgcc	720
catgacgacg acggccggccg gtcagcgcac gtccgagatgg tcaccatgat cctcacggtg	780
gtgctcgccg gccacgagac caccgcgcac ctataggca acggcacggc ggcgtgc	840
acccaccccg accagctgcg gctgctcagg gacgacccgg ctctgtttcc ccgtgcccgc	900
cacgagctgt tgcgctggc cggccggc cacatgaccc agatgcggtt tgcgccgag	960
gatgtcgaca tcgccccggac gaagatccgt aaggggcgacg ccgtacaact gatccctggta	1020
tcggccaact tcgacccccc ccactacacc gaccccgAAC gtctcgaccc gacccgtcac	1080
cccgccggcc acgcccggaa ccatgtggc ttccggccacg ggatgcacta ctgcctggc	1140
gccacccctcg ccaaacagga gggcgaagtc gcgttcgaga agctttcgc gcactacccg	1200
gaggtgtcgcc tggcgctcgc accggaacaa ctgaaaagga caccactgcc cggcagctgg	1260
cggctcgatt ccctgcccgtc gcggttcgaa taa	1293

<210> 26

<211> 430

<212> PRT

<213> Streptomyces platensis

<400> 26

Met Ser Ala Leu Ser Ser Ser Pro Phe Ala Ala His Val Gly Lys His			
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Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Ala Asp Pro Phe			

	20	25	30													
Gly	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg	
	35				40							45				
Phe	Met	Asp	Asp	Ser	Pro	Val	Trp	Leu	Val	Thr	Arg	Phe	Glu	Glu	Val	
	50				55							60				
Arg	Gln	Val	Leu	Arg	Asp	Gln	Arg	Phe	Leu	Asn	Asp	Pro	Thr	Ala	Pro	
	65				70				75				80			
Ser	Leu	Gly	Arg	Ser	Phe	Asp	Asp	Ser	Pro	Thr	Ala	Arg	Leu	Leu	Glu	
	85					90							95			
Met	Met	Gly	Leu	Pro	Glu	His	Phe	Arg	Pro	Tyr	Leu	Leu	Gly	Ser	Ile	
	100					105							110			
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser	
	115					120							125			
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu	
	130					135							140			
Gln	Ile	Ala	Asp	Glu	Leu	Leu	Thr	Arg	Leu	Pro	Glu	Tyr	Ala	Glu	Asp	
	145					150					155				160	
Gly	Val	Val	Asp	Leu	Ile	Lys	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Ala	
	165					170								175		
Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Ala	Glu	Ala	Asp	Arg	Pro	Gln	Trp	
	180					185							190			
Arg	Lys	Trp	Gly	Ala	Asp	Leu	Val	Ser	Leu	Gln	Pro	Asp	Arg	Leu	Ser	
	195					200							205			
Thr	Ser	Phe	Pro	Ala	Met	Ile	Glu	His	Ile	His	Glu	Leu	Ile	Arg	Glu	
	210					215							220			
Arg	Arg	Gly	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Ala	
	225					230					235				240	
His	Asp	Asp	Asp	Gly	Gly	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met	
	245					250								255		
Ile	Leu	Thr	Val	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile	
	260					265							270			
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu	
	275					280							285			
Leu	Arg	Asp	Asp	Pro	Ala	Leu	Phe	Pro	Arg	Ala	Val	His	Glu	Leu	Leu	
	290					295							300			
Arg	Trp	Cys	Gly	Pro	Val	His	Met	Thr	Gln	Met	Arg	Phe	Ala	Ser	Glu	
	305					310					315				320	
Asp	Val	Asp	Ile	Ala	Gly	Thr	Lys	Ile	Arg	Lys	Gly	Asp	Ala	Val	Gln	
	325					330							335			
Leu	Ile	Leu	Val	Ser	Ala	Asn	Phe	Asp	Pro	Arg	His	Tyr	Thr	Asp	Pro	
	340					345							350			
Glu	Arg	Leu	Asp	Leu	Thr	Arg	His	Pro	Ala	Gly	His	Ala	Glu	Asn	His	
	355					360							365			
Val	Gly	Phe	Gly	His	Gly	Met	His	Tyr	Cys	Leu	Gly	Ala	Thr	Leu	Ala	
	370					375							380			
Lys	Gln	Glu	Gly	Glu	Val	Ala	Phe	Glu	Lys	Leu	Phe	Ala	His	Tyr	Pro	
	385					390					395				400	
Glu	Val	Ser	Leu	Gly	Val	Ala	Pro	Glu	Gln	Leu	Glu	Arg	Thr	Pro	Leu	
	405					410							415			
Pro	Gly	Ser	Trp	Arg	Leu	Asp	Ser	Leu	Pro	Leu	Arg	Leu	Arg			
	420					425							430			

<211> 1293

<212> DNA

<213> Streptomyces platensis

<400> 27

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caggccccgg	tcgtacgggg	ccggttcggt	gacgactcac	cggtctggtt	cgtgacgcgc	180
ttcgaggagg	tccgccaagt	cctgcgcgac	cagcggttcg	tgaacaatcc	ggccgcgcgg	240
cccctggccc	catcggccga	ggagaacccg	ctgaccagac	tgatggacat	gctgggcctc	300
cccgagcacc	tccgcgtcta	catgctcggg	tcgattctca	actacgacgc	ccccgaccac	360
acccggctgc	gccgtctggt	gtcgcgcgcg	ttcacggcgc	ggaagatcac	cgatctgcga	420
ccgcgtgtcg	agcagatcgc	cgacgagctg	ctggccccc	tcccccagta	cgccgaggac	480
ggcgtcgctcg	acctcatcca	gcatttcgcc	tacccgctgc	cgatcaccgt	catctgcgag	540
ctggtcggca	tacccgaagc	ggaccgcgg	cagtggcga	agtggggcgc	cgacctcata	600
tcgatggacc	cggaccggct	cggcgcgaacg	ttcccgagca	tgatcgagca	catccatgag	660
atggtccggg	agccgcgcgc	ggcgctcacc	gatgatctgc	tcagcgagct	gatccgtacc	720
catgacgacg	atggcggccg	gctcagcgcac	gtcgagatgg	tcaccatgtat	cctcacgctc	780
gtcctcgccg	gtcacgagac	caccgcaccc	ctcatcagaca	acggcacggc	ggcgctgctc	840
acccaccccg	accagctgcg	cctgctcaag	gacgacccgg	ccctgctccc	ccgggcccgtc	900
catgagctga	tgcgctgggt	cgggccgggt	cagatgacgc	agctgcgcta	cgccgcccgc	960
gacgtcgacc	tcgcccgtac	gcccgttccac	aaggcgcacg	ccgtacaact	cctcctgggt	1020
gccccgcgaa	tcgacccccc	ccactacacc	gacccgcacc	gtctcgatct	gacgcgtcac	1080
cccgccggcc	acggcgagaa	ccatgtgggt	ttcggccacg	gtgcgcatta	ctgcctgggt	1140
gccaccctcg	ccaagcagga	gggcgaagtc	gcgttcggca	agctgcgcgc	gcactacccg	1200
gagatgtccc	tggcatcga	accggAACGT	ctggagcgtat	tgccgctgccc	tggcaactgg	1260
cggtgaatt	ccctggccgtt	cgggctgggg	tga			1293

<210> 28

<211> 430

<212> PRT

<213> Streptomyces platensis

<400> 28

Met	Ser	Ala	Leu	Ser	Ser	Ser	Pro	Phe	Ala	Ala	His	Val	Gly	Lys	His
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Pro	Gly	Glu	Pro	Asn	Val	Met	Glu	Pro	Ala	Leu	Leu	Thr	Asp	Pro	Phe
						20				25			30		
Ala	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Ala	Pro	Val	Val	Arg	Gly	Arg
						35				40			45		
Phe	Val	Asp	Asp	Ser	Pro	Val	Trp	Phe	Val	Thr	Arg	Phe	Glu	Glu	Val
						50			55			60			
Arg	Gln	Val	Leu	Arg	Asp	Gln	Arg	Phe	Val	Asn	Asn	Pro	Ala	Ala	Pro
						65			70			75			80
Pro	Leu	Ala	Pro	Ser	Ala	Glu	Glu	Asn	Pro	Leu	Thr	Arg	Leu	Met	Asp
						85			90			95			
Met	Leu	Gly	Leu	Pro	Glu	His	Leu	Arg	Val	Tyr	Met	Leu	Gly	Ser	Ile
						100			105			110			
Leu	Asn	Tyr	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
						115			120			125			
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu
						130			135			140			
Gln	Ile	Ala	Asp	Glu	Leu	Leu	Ala	Arg	Leu	Pro	Glu	Tyr	Ala	Glu	Asp

145	150	155	160
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr			
165	170	175	
Val Ile Cys Glu Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp			
180	185	190	
Arg Lys Trp Gly Ala Asp Leu Ile Ser Met Asp Pro Asp Arg Leu Gly			
195	200	205	
Ala Thr Phe Pro Ala Met Ile Glu His Ile His Glu Met Val Arg Glu			
210	215	220	
Arg Arg Ala Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr			
225	230	235	240
His Asp Asp Asp Gly Gly Arg Leu Ser Asp Val Glu Met Val Thr Met			
245	250	255	
Ile Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile			
260	265	270	
Ser Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu			
275	280	285	
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met			
290	295	300	
Arg Trp Cys Gly Pro Val Gln Met Thr Gln Leu Arg Tyr Ala Ala Ala			
305	310	315	320
Asp Val Asp Leu Ala Gly Thr Arg Ile His Lys Gly Asp Ala Val Gln			
325	330	335	
Leu Leu Leu Val Ala Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro			
340	345	350	
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His			
355	360	365	
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala			
370	375	380	
Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Ala His Tyr Pro			
385	390	395	400
Glu Met Ser Leu Gly Ile Glu Pro Glu Arg Leu Glu Arg Leu Pro Leu			
405	410	415	
Pro Gly Asn Trp Arg Leu Asn Ser Leu Pro Leu Arg Leu Gly			
420	425	430	

<210> 29
<211> 1293
<212> DNA
<213> Streptomyces lydicus

<400> 29

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aacgtgatgg atccggcgct gatcggtat ccgttcgcgc gttacggcgc gctgcgcgag	120
caggcccgg tcgtcgaaaa gcggttcgtg gagactccc ccgtgtgggt cgtgaccggc	180
tccgaggagg tccgcgaggt cctgcgggac cagcgggttcc ggaacaatcc ggtctcctcg	240
gcgcgggacg cggacccccga ggacaccccg ctgtccccgc tgatggacat gatgggtttc	300
cccgagcacc tgcgctcta tctgctcgcc tcgatcctca acaacgacgc ccccgaccac	360
acccggctgc gccgcctgggt ctccgggcc ttccacgcgc ggaagatcac cgatctgcgg	420
cccgacgtcg cacagatagc cgacgagctg ctggccggc tgccggagca cgccgaggac	480
ggcgtcgtag acctgatcca gcacttcgccc tatccccctgc cgatcaccgt catctgcgaa	540
ctgggtcgaa tccccgagga ggaccggcccg cagtggcgca cctggggcgc cgacctggtc	600
tcgctgcagc cggaccggat gagccggatcc ttcccgccga tgatcgacca catccacgag	660

ctgatcgccg	cgcggcgccg	ggcgctcacc	gacgacctgc	tca	gagct	gatccgaacc	720
catgacgacg	acggcagccg	gctcagcgcac	gtcgagatgg	tcaccatgg	cctcaccgtc	780	
gtcctggccg	gccacgagac	caccgcgcac	ctcatcgca	acggcacggc	ggccctgctc	840	
accccaccccg	accagctg	cg	gctcaag	gacgacc	cgctg	gcccgg	900
cac	gagttg	tg	gcgtgg	cg	ccgg	ccgag	960
gac	gtcgagc	tg	gggggcgt	ccggatccgc	aagggg	gacg	1020
tcggc	gaacc	g	gatccgc	ccactacacc	gaacccg	gac	1080
ccc	ccgg	ac	gagaa	ccatgtgg	tccggccac	gggcg	1140
g	ccacgctcg	cc	agcag	gggcgaggtc	gc	ccctg	1200
g	gctcg	tg	ggccgtcg	gcccgg	cc	ctgtctgg	1260
g	gtaatg	cg	ctgccc	ctggag	gttacc	ggcag	1293
cg	gtgaat	cg	ctgccc	gttacc	ggcag	ctgg	

<210> 30

<211> 430

<212> PRT

<213> Streptomyces lydicus

<400> 30

Met	Ser	Ala	Leu	Pro	Ser	Asn	Thr	Phe	Thr	Glu	His	Val	Gly	Lys	His
1								5		10				15	
Pro	Gly	Pro	Asn	Val	Met	Asp	Pro	Ala	Leu	Ile	Gly	Asp	Pro	Phe	
									20	25				30	
Ala	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg
								35	40				45		
Phe	Val	Asp	Asp	Ser	Pro	Val	Trp	Phe	Val	Thr	Arg	Phe	Glu	Val	
								50	55				60		
Arg	Glu	Val	Leu	Arg	Asp	Gln	Arg	Phe	Arg	Asn	Asn	Pro	Val	Ser	Ser
								65	70				75		80
Ala	Pro	Asp	Ala	Asp	Pro	Glu	Asp	Thr	Pro	Leu	Ser	Arg	Leu	Met	Asp
								85		90				95	
Met	Met	Gly	Phe	Pro	Glu	His	Leu	Arg	Val	Tyr	Leu	Gly	Ser	Ile	
								100		105			110		
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
								115		120			125		
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Ala
								130		135			140		
Gln	Ile	Ala	Asp	Glu	Leu	Leu	Ala	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp
								145		150			155		160
Gly	Val	Val	Asp	Leu	Ile	Gln	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Thr
								165		170			175		
Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Pro	Glu	Glu	Asp	Arg	Pro	Gln	Trp
								180		185			190		
Arg	Thr	Trp	Gly	Ala	Asp	Leu	Val	Ser	Leu	Gln	Pro	Asp	Arg	Met	Ser
								195		200			205		
Arg	Ser	Phe	Pro	Ala	Met	Ile	Asp	His	Ile	His	Glu	Leu	Ile	Ala	Ala
								210		215			220		
Arg	Arg	Arg	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Thr
								225		230			235		240
His	Asp	Asp	Asp	Gly	Ser	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met
								245		250			255		
Val	Leu	Thr	Val	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile
								260		265			270		
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu

275	280	285
Leu Lys Asp Asp Pro Ala Leu	Leu Pro Arg Ala Val His	Glu Leu Met
290	295	300
Arg Trp Cys Gly Pro Val His	Met Thr Gln Leu Arg Tyr	Ala Ala Glu
305	310	315
Asp Val Glu Leu Ala Gly Val	Arg Ile Arg Lys Gly Asp	Ala Val Gln
325	330	335
Leu Ile Leu Val Ser Ala Asn	Arg Asp Pro Arg His Tyr	Thr Glu Pro
340	345	350
Asp Arg Leu Asp Leu Thr Arg	His Pro Ala Gly His Ala	Glu Asn His
355	360	365
Val Gly Phe Gly His Gly Ala	His Tyr Cys Leu Gly Ala	Thr Leu Ala
370	375	380
Lys Gln Glu Gly Glu Val Ala	Leu Gly Ala Leu Leu Arg His	Phe Pro
385	390	395
Glu Leu Ser Leu Ala Val Ala	Pro Asp Ala Leu Glu Arg Thr	Pro Val
405	410	415
Pro Gly Ser Trp Arg Leu Asn Ala	Leu Pro Leu Arg Leu Arg	
420	425	430

<210> 31

<211> 1293

<212> DNA

<213> Streptomyces lydicus

<400> 31

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atgtcgccat cgaccagctc tcccctcagc gcccacgtcg gcaagcaccc gggcgaaccc      60
catgtatgg atccggcgct gatcagcgat ccgttcggcg gctacgggtgc cctgcgcgag      120
cagggacccg tcgtccgcgg acggttcttc gacgactcgc ctttgtggtt agtgaccgc      180
ttcgaggaag tccgccaggt cctgcgcgac cagcggttcg tgaacaaccc cgccgacccg      240
gcgctcgccg tcgcgccccg ggactccccg cagctgcgcg cgctggcgat gctgggcattc      300
cccgagcacc tgcacggcta tctgctcaac tcgatcctca actacgacgc ccccgaccac      360
acccggctgc gcccctggt ctcccgccgc ttccacggccc gcaagatcac cgatcttcgg      420
cccggggtgg cgagataaac cgccgagctg ctggaccgac tccggagca cgccgaggac      480
ggcgtggtcg acctgatcga gcacttcgcc taccgcgtgc cgatcacgggt gatctgcgaa      540
cttgcggca tcgcccggca ggaccggccc cagtggcggtt cctggggcgc cgacctggtc      600
tcggtgacc ccgaccggct cggccggacc ttcccgccga tgatcgacca catccacgcg      660
ctgatcgcc accggccggc cgcgctcacc gacgacctgc tcagcgagct gatccggacc      720
catgacgacg acgcagccg gtcagcgac gtcgagatgg tcaccctggt cctcaccctc      780
gtgctggccg gccacgagac caccgcacac ctcatggca acggcaccgc ggcctgctc      840
acccaccccg accagctgcg gtcgctcaag gacgacccgg cgctgctgccc ggcgcggcgc      900
cacgactga tgcgctggtg cggccggtg cacgtcaccc agctgcggta cgccgcccgg      960
gacgtcgacc tcgcccggcac cggatccgc aggggcgacg ccgtgcaggc cgtcctggtc      1020
tcggcgaacc acgaccggcg ccactacacc gacccggaaac gcctggaccc gacccggcag      1080
cccgccggcc gcccggagaa ccacgtgggc ttccggcactg gggccgacta ctgcctggc      1140
gccagcctcg ccaggcaggaa gggtgagggtc gcccctggcg ccctgttgcg cgcgtaccc      1200
gacctggcgc tggcggtggc gcccggaggag ctggagcgc ccccggtgcc cggtacctgg      1260
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<210> 32

<211> 430

<212> PRT

<213> Streptomyces lydicus

<400> 32

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Pro	Gly	Glu	Pro	His	Val	Met	Asp	Pro	Ala	Leu	Ile	Ser	Asp	Pro	Phe
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Gly	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg
															35
Phe	Phe	Asp	Asp	Ser	Pro	Leu	Trp	Leu	Val	Thr	Arg	Phe	Glu	Glu	Val
															50
Arg	Gln	Val	Leu	Arg	Asp	Gln	Arg	Phe	Val	Asn	Asn	Pro	Ala	Asp	Pro
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Ala	Leu	Gly	Val	Ala	Pro	Glu	Asp	Ser	Pro	Gln	Leu	Arg	Ala	Leu	Ala
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Met	Leu	Gly	Ile	Pro	Glu	His	Leu	His	Gly	Tyr	Leu	Leu	Asn	Ser	Ile
															100
Leu	Asn	Tyr	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
															115
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Ala
															130
Gln	Ile	Thr	Ala	Glu	Leu	Leu	Asp	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp
															145
Gly	Val	Val	Asp	Leu	Ile	Glu	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Thr
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Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Ala	Ala	Glu	Asp	Arg	Pro	Gln	Trp
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Arg	Ser	Trp	Gly	Ala	Asp	Leu	Val	Ser	Val	Asp	Pro	Asp	Arg	Leu	Gly
															195
Arg	Thr	Phe	Pro	Ala	Met	Ile	Asp	His	Ile	His	Ala	Leu	Ile	Gly	Gln
															210
Arg	Arg	Ala	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Thr
															225
His	Asp	Asp	Asp	Gly	Ser	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Leu
															245
Val	Leu	Thr	Leu	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile
															260
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu
															275
Leu	Lys	Asp	Asp	Pro	Ala	Leu	Leu	Pro	Arg	Ala	Val	His	Glu	Leu	Met
															290
Arg	Trp	Cys	Gly	Pro	Val	His	Val	Thr	Gln	Leu	Arg	Tyr	Ala	Ala	Glu
															305
Asp	Val	Asp	Leu	Ala	Gly	Thr	Arg	Ile	Arg	Arg	Gly	Asp	Ala	Val	Gln
															325
Ala	Val	Leu	Val	Ser	Ala	Asn	His	Asp	Pro	Arg	His	Tyr	Thr	Asp	Pro
															340
Glu	Arg	Leu	Asp	Leu	Thr	Arg	Gln	Pro	Ala	Gly	Arg	Ala	Glu	Asn	His
															355
Val	Gly	Phe	Gly	His	Gly	Ala	His	Tyr	Cys	Leu	Gly	Ala	Ser	Leu	Ala
															370
Arg	Gln	Glu	Gly	Glu	Val	Ala	Leu	Gly	Ala	Leu	Phe	Asp	Arg	Tyr	Pro
															385
Asp	Leu	Ala	Leu	Ala	Val	Ala	Pro	Glu	Glu	Leu	Glu	Arg	Thr	Pro	Val
															405
															410
															415

Pro Gly Thr Trp Arg Leu Thr Ser Leu Pro Val Arg Leu Gly
 420 425 430

<210> 33

<211> 1281

<212> DNA

<213> Streptomyces tubercidicus

<400> 33

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gtacggggcc ggttcatgga cgactcgccc gtctggctgg tgacgcgggtt cgaggaggtc	180
cggcaggtcc tgcgcgacca gcggttcgtg aacaatccgg cctcgccgtc cctgaactac	240
gcccggagg acaacccgct gacccggctg atggagatgc tgggcctcccc cgagcaccc	300
cgcgtctacc tgctcgatc gatccctaac tacgacgccc ccgaccacac ccggctgcgc	360
cgtctggtgt cgccggcggtt cacggccgc aagatcaccg acctgcggcc ccgggtcgag	420
cagatcgccg acgcgctgct ggccggctg cccgagcacg ccgaggacgg cgtcgctgac	480
ctcatccagc acttcgccta cccctggcg atcaccgtca tctgcgaact ggtcgccata	540
cccgaaaggcc accggccgca gtggcgaacg tggggcgccg acctcatctc gatggatccg	600
gaccggctcg ggcctcggtt cccggcgatg atcagacaca tccatcgat ggtccggaa	660
cggcgcgagg cgctcaccga cgacctgctc agcgaactga tccgcaccca tgacgacgac	720
ggcggggcgcc tcagcgacgt cgagatggtc accatgatcc tcacgctcgat ctcgcggc	780
cacgagacca cggcccaccc catcagcaac ggcacggcg ccgtgctcac ccaccccgac	840
cagctcgctc tggtaagga cgatccggcc ctccctcccc gtgcgtcca cgagctgatg	900
 cgctgggtgcg ggccgggtgca catgacccag ctgcgtacg ccaccgcccga cgtcgaccc	960
gccggcacac cgatccgca gggcgatcc gttcaactca tcctggtatac ggccaacttc	1020
gaccggcgatc actacaccga ccccgaccgc ctgcgtatca cccggcaccgc cgccggccac	1080
gcccggaaacc atgtgggtt cggccatgga ggcactact gcctgggcgc cacactcgcc	1140
aaacaggaag gtgaagtgc cttcgaaaa ctgctcacgc actacccgga catatcgctg	1200
ggcatcgccc cgaaacaccc ggagcggaca ccgtgcccgg gcaactggcg gctgaactcg	1260
ctgcccggatc ggttggggatc a	1281

<210> 34

<211> 426

<212> PRT

<213> Streptomyces tubercidicus

<400> 34

Met Asn Ser Pro Phe Ala Ala His Val Gly Lys His Pro Gly Glu Pro	
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20 25 30	
Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg Phe Met Asp Asp	
35 40 45	
Ser Pro Val Trp Leu Val Thr Arg Phe Glu Glu Val Arg Gln Val Leu	
50 55 60	
Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ser Pro Ser Leu Asn Tyr	
65 70 75 80	
Ala Pro Glu Asp Asn Pro Leu Thr Arg Leu Met Glu Met Leu Gly Leu	
85 90 95	
Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile Leu Asn Tyr Asp	

	100	105	110
Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser Arg Ala Phe Thr			
115	120	125	
Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu Gln Ile Ala Asp			
130	135	140	
Ala Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp Gly Val Val Asp			
145	150	155	160
Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr Val Ile Cys Glu			
165	170	175	
Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp Arg Thr Trp Gly			
180	185	190	
Ala Asp Leu Ile Ser Met Asp Pro Asp Arg Leu Gly Ala Ser Phe Pro			
195	200	205	
Ala Met Ile Glu His Ile His Gln Met Val Arg Glu Arg Arg Glu Ala			
210	215	220	
Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr His Asp Asp Asp			
225	230	235	240
Gly Gly Arg Leu Ser Asp Val Glu Met Val Thr Met Ile Leu Thr Leu			
245	250	255	
Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile Ser Asn Gly Thr			
260	265	270	
Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu Val Lys Asp Asp			
275	280	285	
Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met Arg Trp Cys Gly			
290	295	300	
Pro Val His Met Thr Gln Leu Arg Tyr Ala Thr Ala Asp Val Asp Leu			
305	310	315	320
Ala Gly Thr Pro Ile Arg Gln Gly Asp Ala Val Gln Leu Ile Leu Val			
325	330	335	
Ser Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro Asp Arg Leu Asp			
340	345	350	
Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His Val Gly Phe Gly			
355	360	365	
His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala Lys Gln Glu Gly			
370	375	380	
Glu Val Ala Phe Gly Lys Leu Leu Thr His Tyr Pro Asp Ile Ser Leu			
385	390	395	400
Gly Ile Ala Pro Glu His Leu Glu Arg Thr Pro Leu Pro Gly Asn Trp			
405	410	415	
Arg Leu Asn Ser Leu Pro Val Arg Leu Gly			
420	425		

<210> 35
<211> 195
<212> DNA
<213> Streptomyces tubercidicus

<400> 35
atgcggatca cgatcgacac cgacatctgt atcggcgccg gccagtgcgc cctgaccgcg 60
ccgggagtgt tcacccagga cgacgacggc ttcatgcgcc tgctgcccgg ccgcgaggac 120
ggtgccggcg acccgctggc gcgggaggcc gccccgcct gccccgtgca ggccatcacg 180
gtcacggacg actga 195

<210> 36
<211> 64
<212> PRT
<213> Streptomyces tubercidicus

<400> 36
Met Arg Ile Thr Ile Asp Thr Asp Ile Cys Ile Gly Ala Gly Gln Cys
1 5 10 15
Ala Leu Thr Ala Pro Gly Val Phe Thr Gln Asp Asp Asp Gly Phe Ser
 20 25 30
Ala Leu Leu Pro Gly Arg Glu Asp Gly Ala Gly Asp Pro Leu Val Arg
 35 40 45
Glu Ala Ala Arg Ala Cys Pro Val Gln Ala Ile Thr Val Thr Asp Asp
 50 55 60

<210> 37
<211> 195
<212> DNA
<213> Streptomyces tubercidicus

<400> 37
atgcggatca ccatcgacac cgacatctgc atcggcgccg gccagtgcgc cctgaccgcg 60
ccggaggtct tcacccagga cgacgacggt ttcagcgccc tgctgcccgg ccgcgaggac
ggcgcggcgcg acccgctggt gcgcgaggcc gccccgcct gccccgtgca ggccatttcg 120
gtcacggacg actga 180
 195

<210> 38
<211> 64
<212> PRT
<213> Streptomyces tubercidicus

<400> 38
Met Arg Ile Thr Ile Asp Thr Asp Ile Cys Ile Gly Ala Gly Gln Cys
1 5 10 15
Ala Leu Thr Ala Pro Gly Val Phe Thr Gln Asp Asp Asp Gly Phe Ser
 20 25 30
Ala Leu Leu Pro Gly Arg Glu Asp Gly Ala Gly Asp Pro Leu Val Arg
 35 40 45
Glu Ala Ala Arg Ala Cys Pro Val Gln Ala Ile Ser Val Thr Asp Asp
 50 55 60

<210> 39
<211> 9
<212> PRT
<213> Artificial Sequence
<220>
<223> Synthetic peptide.

<400> 39

Tyr Pro Tyr Asp Val Pro Asp Tyr Ala
1 5

<210> 40
<211> 10
<212> PRT
<213> Artificial Sequence
<220>
<223> Synthetic peptide.

<400> 40
Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu
1 5 10

<210> 41
<211> 11
<212> PRT
<213> Artificial Sequence
<220>
<223> Synthetic peptide.

<400> 41
Gln Pro Glu Leu Ala Pro Glu Asp Pro Glu Asp
1 5 10

<210> 42
<211> 11
<212> PRT
<213> Artificial Sequence
<220>
<223> Synthetic peptide.

<400> 42
Tyr Thr Asp Ile Glu Met Asn Arg Leu Gly Lys
1 5 10

<210> 43
<211> 7
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 43
Ile Ala Gly His Glu Thr Thr
1 5

<210> 44
<211> 20
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (6)..(18)
<223> Nucleotides 6, 9 and 18 are "s" wherein "s" = g or c.

<400> 44
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20

<210> 45
<211> 7
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 45
Val Ala Gly His Glu Thr Thr
1 5

<210> 46
<211> 20
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (3)..(18)
<223> Nucleotides 3, 6, 9, and 18 are "s" wherein "s" = g or c.

<400> 46
gtsgcsggsc acgagacsac

20

<210> 47
<211> 7
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 47
Leu Ala Gly His Glu Thr Thr
1 5

<210> 48
<211> 20
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (3)..(18)
<223> Nucleotides 3, 6, 9, and 18 are "s" wherein "s" = g or c.

<400> 48
ctsgcsggsc acgagacsa

20

<210> 49
<211> 9
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 49
Leu Leu Leu Ile Ala Gly His Glu Thr
1 5

<210> 50
<211> 25
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (2)..(17)
<223> Nucleotides 2, 5, 8, 14, and 17 are "s" wherein "s" = g or c.

<400> 50
tsctsctsat cgcsggscac gagac

25

<210> 51
<211> 9
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 51
His Gln Cys Leu Gly Gln Asn Leu Ala
1 5

<210> 52
<211> 26
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (12)..(24)
<223> Nucleotides 12, 15, and 24 are "s" wherein "s" = g or c.

<400> 52
gtggtcacgg asccstgctt ggascg

26

<210> 53
<211> 8
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 53
Phe Gly His Gly Val His Gln Cys
1 5

<210> 54
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (6)..(15)
<223> Nucleotides 6, 12, and 15 are "s" wherein "s" = g or c.

<400> 54
aagccsgtgtc cscasgtggc cacg

24

<210> 55
<211> 8
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 55

Phe Gly Phe Gly Val His Gln Cys
1 5

<210> 56
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (6)..(15)
<223> Nucleotides 6, 12, and 15 are "s" wherein "s" = g or c.

<400> 56
aaggcsaagc cscasgtggt cacg

24

<210> 57
<211> 8
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 57
Phe Gly His Gly Ile His Gln Cys
1 5

<210> 58
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (6)..(12)
<223> Nucleotides 6 and 12 are "s" wherein "s" = g or c.

<400> 58
aagccsgtg cstaggtggt cacg

24

<210> 59
<211> 8
<212> PRT
<213> Streptomyces
<220>
<221> misc_feature
<222>
<223> Streptomyces consensus sequence

<400> 59
Phe Gly His Gly Val His Phe Cys
1 5

<210> 60
<211> 24
<212> DNA
<213> Artificial Sequence

<220>
<221> misc_feature
<222> (6)..(15)
<223> Nucleotides 6, 12, and 15 are "s" wherein "s" = g or c.

<400> 60
aagccsgtg cscasgtgaa gacg

24

<210> 61
<211> 24
<212> PRT
<213> Streptomyces tubercidicus

<400> 61
His Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Thr Asp Pro
1 5 10 15
Phe Thr Gly Tyr Gly Ala Leu Arg
20

<210> 62
<211> 21
<212> PRT
<213> Streptomyces tubercidicus

<400> 62
Phe Val Asn Asn Pro Ala Ser Pro Ser Leu Asn Tyr Ala Pro Glu Asp
1 5 10 15
Asn Pro Leu Thr Arg
20

<210> 63
<211> 19
<212> PRT
<213> Streptomyces tubercidicus

<400> 63
Leu Leu Thr His Tyr Pro Asp Ile Ser Leu Gly Ile Ala Pro Glu His
1 5 10 15
Leu Glu Arg

<210> 64
<211> 17
<212> PRT
<213> Streptomyces tubercidicus

<400> 64
Val Tyr Leu Leu Gly Ser Ile Leu Asn Tyr Asp Ala Pro Asp His Thr
1 5 10 15
Arg

<210> 65
<211> 13
<212> PRT
<213> Streptomyces tubercidicus

<400> 65
Thr Trp Gly Ala Asp Leu Ile Ser Met Asp Pro Asp Arg
1 5 10

<210> 66
<211> 13
<212> PRT
<213> Streptomyces tubercidicus

<400> 66
Glu Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg
1 5 10

<210> 67
<211> 12
<212> PRT
<213> Streptomyces tubercidicus

<400> 67
Phe Met Asp Asp Ser Pro Val Trp Leu Val Thr Arg
1 5 10

<210> 68
<211> 12
<212> PRT
<213> Streptomyces tubercidicus

<400> 68
Leu Met Glu Met Leu Gly Leu Pro Glu His Leu Arg
1 5 10

<210> 69

<211> 11
<212> PRT
<213> Streptomyces tubercidicus

<400> 69
Val Glu Gln Ile Ala Asp Ala Leu Leu Ala Arg
1 5 10

<210> 70
<211> 11
<212> PRT
<213> Streptomyces tubercidicus

<400> 70
Leu Val Lys Asp Asp Pro Ala Leu Leu Pro Arg
1 5 10

<210> 71
<211> 8
<212> PRT
<213> Streptomyces tubercidicus

<400> 71
Asp Asp Pro Ala Leu Leu Pro Arg
1 5

<210> 72
<211> 8
<212> PRT
<213> Streptomyces tubercidicus

<400> 72
Thr Pro Leu Pro Gly Asn Trp Arg
1 5

<210> 73
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 73
Leu Asn Ser Leu Pro Val Arg
1 5

<210> 74
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 74
Ile Thr Asp Leu Arg Pro Arg
1 5

<210> 75
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 75
Glu Gln Gly Pro Val Val Arg
1 5

<210> 76
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 76
Ala Val His Glu Leu Met Arg
1 5

<210> 77
<211> 5
<212> PRT
<213> Streptomyces tubercidicus

<400> 77
Ala Phe Thr Ala Arg
1 5

<210> 78
<211> 5
<212> PRT
<213> Streptomyces tubercidicus

<400> 78
Phe Glu Glu Val Arg
1 5

<210> 79
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 79
Pro Gly Glu Asp Asn Val Met
1 5

<210> 80
<211> 21
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (3)..(18)
<223> Nucleotides 3, 6, 12, and 18 are "s" wherein "s" = c or g.
<220>
<221> misc_feature
<222> (9)..(9)
<223> Nucleotide 9 is "r" wherein "r" = a or g.
<220>
<221> misc_feature
<222> (15)..(15)
<223> Nucleotide 15 is "y" wherein "y" = c or t.

<400> 80
ccsgggarc csaaygtsat g

21

<210> 81
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 81
Ala Leu Ile Thr Asp Pro Phe
1 5
<210> 82
<211> 21
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (3)..(18)
<223> Nucleotides 3, 6, 12, and 18 are "s" wherein "s" = c or g.

<400> 82
gcsctsaty a csgacccstt c

21

<210> 83
<211> 8
<212> PRT
<213> Streptomyces tubercidicus

<400> 83
Phe Met Asp Asp Ser Pro Val Trp
1 5

<210> 84
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (13)..(13)
<223> Nucleotide 13 is "w" wherein "w" = a or t.
<220>
<221> misc_feature
<222> (14)..(21)
<223> Nucleotides 14, 15, 18, and 21 are "s" wherein "s" = c or g.

<400> 84
ttcatggacg acwssccsgt stgg

24

<210> 85
<211> 8
<212> PRT
<213> Streptomyces tubercidicus

<400> 85
Leu Asn Tyr Asp Ala Pro Asp His
1 5

<210> 86
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (3)..(18)
<223> Nucleotides 3, 15 and 18 are "s" wherein "s" = c or g.
<220>
<221> misc_feature
<222> (6)..(9)
<223> Nucleotides 6 and 9 are "y" wherein "y" = c or t.

<400> 86
ctsaaytayg acgcscsga ccac

24

<210> 87
<211> 8
<212> PRT
<213> Streptomyces tubercidicus

<400> 87
Val Glu Gln Ile Ala Asp Ala Leu
1 5

<210> 88
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (3)..(24)
<223> Nucleotides 3, 15, 21, and 24 are "s" wherein "s" = c or g.
<220>
<221> misc_feature
<222> (12)..(12)
<223> Nucleotide 12 is "y" wherein "y" = c or t.
<220>
<221> misc_feature
<222> (6)..(6)
<223> Nucleotide 6 is "r" wherein "r" = a or g.

<400> 88
gtsgarcaga tygcgacgc scts

24

<210> 89
<211> 8
<212> PRT
<213> Streptomyces tubercidicus

<400> 89
Asp Leu Ile Ser Met Asp Pro Asp
1 5

<210> 90
<211> 24
<212> DNA
<213> Artificial Sequence
<220>
<221> misc_feature
<222> (6)..(21)
<223> Nucleotides 6, 11, 12, and 21 are "s" wherein "s" = c or g.
<220>
<221> misc_feature
<222> (9)..(9)
<223> Nucleotide 9 is "r" wherein "r" = a or g.
<220>
<221> misc_feature
<222> (10)..(10)
<223> Nucleotide 10 is "w" wherein "w" = a or t.

<400> 90
ctggastarw sstacctggg sctg

24

<210> 91
<211> 36

<212> DNA
<213> Streptomyces tubercidicus

<400> 91
agattaat~~ta~~ atgtcggaat taatgaactg tccgtt

36

<210> 92
<211> 32
<212> DNA
<213> Streptomyces tubercidicus

<400> 92
aaactcaccc caaccgcacc ggcagc~~g~~agt tc

32

<210> 93
<211> 7
<212> PRT
<213> Streptomyces tubercidicus

<400> 93
Met Ser Glu Leu Met Asn Ser
1 5

<210> 94
<211> 1293
<212> DNA
<213> Streptomyces tubercidicus

<400> 94
atgtcg~~gg~~caa tatccagctc cccgttcg~~cc~~ gcacacgtcg gaaagcatcc cggcgaggccg 60
aatgtgatgg acccg~~gg~~cg~~ct~~ gatcac~~cc~~gac ccgttcggcg gctacggcg~~c~~ actgcgtgag 120
caaggccccg tcctacc~~gg~~gg ccggttcatg gacgactcac ccgtctggct cgtgacgcgc 180
ttcgaagagg tccgcca~~ag~~t cctgcgc~~at~~ cagcgg~~tt~~cc tgaacaaccc ggccgcgtcg 240
tcaccggggc attcgat~~cg~~a cgagagcccc acggccaggc tgctggacat gatggggatg 300
cccgaacatt tccggcgta tctgatgggg t~~c~~gatcctca acaacgacgc ccccgaccac 360
acccggctgc g~~cc~~gtctgg~~t~~ gtcacgcgc ttcacggcac gcaagatcac cgatctgcgg 420
ccgcgggtcg agcagctcg~~c~~ cgacgagctg ctggccggc ttcccgagca cgccgaggac 480
ggtgtgg~~tc~~g acctgatcaa gcacttcg~~cc~~ tatcccc~~t~~gc cgatcaccgt gatctgc~~ga~~a 540
ctgg~~t~~cg~~gg~~ca tcccggaa~~g~~cc ggaccggccg caatggcg~~ga~~ agtggggcgc cgac~~ct~~cg~~tt~~ 600
tcgctgc~~ag~~c cggagcgg~~ct~~ cagcac~~ct~~cg ttcccg~~gg~~ca tgatcgagca catccatgaa 660
ctgatcccg~~cg~~ agcggcgc~~gg~~ cg~~cg~~ctcacc gacgatctgc tcagc~~g~~ag~~ct~~ gatccgtacc 720
catgacgac~~g~~ acggcagccg g~~c~~tcagcgac gtcgagatgg tcaccatgg~~t~~ cctcaccg~~tc~~ 780
gtcctggccg g~~cc~~ac~~g~~agac caccgcccac ctgatagg~~ca~~ acggcac~~gg~~gc ggcgtgc~~tc~~ 840
acccaccccg accagctgc~~g~~ cctggta~~ag~~ gacgacccgg agctgcttcc gcgtccg~~tc~~ 900
cacgagctgc tgcgctgg~~tg~~ cggccggc~~t~~ cagatgaccc agctgc~~gg~~ta cgcctccgag 960
gatgtcgaga tcgcccggac g~~cc~~gatccgt aagg~~gg~~cgac~~g~~ ccgtacaact c~~at~~c~~c~~t~~gt~~ta 1020
tcggcgaact tcgaccccg ccactacacc gccccg~~aa~~c gcctcgac~~ct~~ gacccg~~cc~~ac 1080
cccgccggcc acggc~~g~~agaa ccatgtggc~~gc~~ ttccggccac~~g~~ gaatgcacta ctgcctggc 1140
gccaccctcg ccaa~~ac~~agga gggcga~~gt~~tc g~~c~~ttcggc~~ga~~ agctcttac~~c~~ g~~c~~actacccg 1200
gagctgtcgc tggccgtcgc accggac~~g~~ag ttggagc~~g~~aa c~~g~~ccgg~~gt~~gc~~c~~ cggcagctgg 1260

cggttggatt cgctgccggt gcggttgggg tga

1293

<210> 95

<211> 430

<212> PRT

<213> Streptomyces tubercidicus

<400> 95

Met	Ser	Ala	Ile	Ser	Ser	Ser	Pro	Phe	Ala	Ala	His	Val	Gly	Lys	His
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Pro	Gly	Glu	Pro	Asn	Val	Met	Asp	Pro	Ala	Leu	Ile	Thr	Asp	Pro	Phe
									20		25			30	
Gly	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Leu	Pro	Gly	Arg
								35		40			45		
Phe	Met	Asp	Asp	Ser	Pro	Val	Trp	Leu	Val	Thr	Arg	Phe	Glu	Glu	Val
	50							55			60				
Arg	Gln	Val	Leu	Arg	Asp	Gln	Arg	Phe	Leu	Asn	Asn	Pro	Ala	Ala	Ser
	65							70			75			80	
Ser	Pro	Gly	His	Ser	Ile	Asp	Glu	Ser	Pro	Thr	Ala	Arg	Leu	Leu	Asp
								85		90			95		
Met	Met	Gly	Met	Pro	Glu	His	Phe	Arg	Pro	Tyr	Leu	Met	Gly	Ser	Ile
								100		105			110		
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
								115		120			125		
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu
								130		135			140		
Gln	Leu	Ala	Asp	Glu	Leu	Leu	Ala	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp
	145							150			155			160	
Gly	Val	Val	Asp	Leu	Ile	Lys	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Thr
								165		170			175		
Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Pro	Glu	Ala	Asp	Arg	Pro	Gln	Trp
								180		185			190		
Arg	Lys	Trp	Gly	Ala	Asp	Leu	Val	Ser	Leu	Gln	Pro	Glu	Arg	Leu	Ser
								195		200			205		
Thr	Ser	Phe	Pro	Ala	Met	Ile	Glu	His	Ile	His	Glu	Leu	Ile	Arg	Glu
								210		215			220		
Arg	Arg	Gly	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Thr
	225							230			235			240	
His	Asp	Asp	Asp	Gly	Ser	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met
								245		250			255		
Val	Leu	Thr	Val	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile
								260		265			270		
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu
								275		280			285		
Val	Lys	Asp	Asp	Pro	Glu	Leu	Leu	Pro	Arg	Ala	Val	His	Glu	Leu	Leu
								290		295			300		
Arg	Trp	Cys	Gly	Pro	Val	Gln	Met	Thr	Gln	Leu	Arg	Tyr	Ala	Ser	Glu
	305							310			315			320	
Asp	Val	Glu	Ile	Ala	Gly	Thr	Pro	Ile	Arg	Lys	Gly	Asp	Ala	Val	Gln
								325		330			335		
Leu	Ile	Leu	Val	Ser	Ala	Asn	Phe	Asp	Pro	Arg	His	Tyr	Thr	Ala	Pro
								340		345			350		

Glu Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His
 355 360 365
 Val Gly Phe Gly His Gly Met His Tyr Cys Leu Gly Ala Thr Leu Ala
 370 375 380
 Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Phe Thr His Tyr Pro
 385 390 395 400
 Glu Leu Ser Leu Ala Val Ala Pro Asp Glu Leu Glu Arg Thr Pro Val
 405 410 415
 Pro Gly Ser Trp Arg Leu Asp Ser Leu Pro Val Arg Leu Gly
 420 425 430

<210> 96
 <211> 18
 <212> DNA
 <213> Artificial Sequence

<400> 96
 cgscscscsc tswssaas 18

<210> 97
 <211> 21
 <212> DNA
 <213> Artificial Sequence

<400> 97
 sassgcstts bcccartgtyt c 21

<210> 98
 <211> 1266
 <212> DNA
 <213> Streptomyces tubercidicus

<400> 98
 gtgggtcgacg cacaccagac gttcgtcatc gtcgggggtg gcctggccgg cgcaaaggcc 60
 gcggagactc tccgcgcgga ggggttccacc ggccgggtga tcctcatctg tgacgagcgc 120
 gaccaccgt acgagcgc(cc cccgctctcc aaggggttcc tgctcggcaa ggaagagcgc 180
 gacagcgtgt tcgtccatga gccccctgg taacgcccagg cacagatcga actgcacctg 240
 ggccagcccg ccgtccgcct cgaccccgag ggcaggaccc tccgcctcgg cgacggcacc 300
 ctgatcgcc acgacaagct gctgctggcc accggcgcgg aaccgcggcg cctggacatc 360
 cccggcaccg gcctggccgg cgtgcaccac ctgcgcggcc tcgcccacgc cgaacggctg 420
 cgcggcgtcc tggcctccct cggccgcgac aacggccatc tggtgatcgc cggagccggc 480
 tggatcgccc tggaggtcgcc cgccgcggcc cgctccatcg ggcggaggt gaccgtcg 540
 gaggccgcgg cgacccgcgt gcacggcatc ctggggcccg aactcggcg tctgttcacc 600
 gatctgcacc gcgagcacgg cgtccgttcc cacttcggcg cccgttccac cgagatcg 660
 gtagaggccg gcatgggtgct cgccgtgcgc accgacgacg gcgaggaaca cccgcaccc 720
 gatgtgctcg ccgcgatcgcc cgccgcggcc cgacccgcgc tcgccaaca ggccggctg 780
 gatctgcacc accccggagac cggcggccgg gtggccgtcg acgcggcgct ggcaccc 840
 gaccgtaca tctacgcgcg cggtgacgta gccgcgcgg accacccgct gctggacacc 900

cggctgcggg tcgaacactg ggccaaacgcc ctcaacggcg gccccggccgc cgccccggcc 960
 atgctcggcc aggacatcg ctacgaccgc atcccgtaact tcttctccga ccagtaacgc 1020
 gtcggcatgg agtactccgg ctacgccccg cccggctcggt acgcccagggt cgtctgccc 1080
 ggcgacgtcg ccaagcggga gttcatcgcc ttctggctgg cggcggacgg ccggctgctc 1140
 gcgggcattga acgtcaacgt ctgggacgtc gccgagtcca tccagcaact catccgctcc 1200
 ggggcgcctgt tggagcccg cgcaactggcc gatccgcagg ttccgctggc ggcactgctc 1260
 ccgttag 1266

<210> 99
 <211> 421
 <212> PRT
 <213> Streptomyces tubercidicus

<400> 99
 Val Val Asp Ala His Gln Thr Phe Val Ile Val Gly Gly Gly Leu Ala
 1 5 10 15
 Gly Ala Lys Ala Ala Glu Thr Leu Arg Ala Glu Gly Phe Thr Gly Arg
 20 25 30
 Val Ile Leu Ile Cys Asp Glu Arg Asp His Pro Tyr Glu Arg Pro Pro
 35 40 45
 Leu Ser Lys Gly Phe Leu Leu Gly Lys Glu Glu Arg Asp Ser Val Phe
 50 55 60
 Val His Glu Pro Ala Trp Tyr Ala Gln Ala Gln Ile Glu Leu His Leu
 65 70 75 80
 Gly Gln Pro Ala Val Arg Leu Asp Pro Glu Gly Arg Thr Val Arg Leu
 85 90 95
 Gly Asp Gly Thr Leu Ile Ala Tyr Asp Lys Leu Leu Leu Ala Thr Gly
 100 105 110
 Ala Glu Pro Arg Arg Leu Asp Ile Pro Gly Thr Gly Leu Ala Gly Val
 115 120 125
 His His Leu Arg Arg Leu His Ala Glu Arg Leu Arg Gly Val Leu
 130 135 140
 Ala Ser Leu Gly Arg Asp Asn Gly His Leu Val Ile Ala Gly Ala Gly
 145 150 155 160
 Trp Ile Gly Leu Glu Val Ala Ala Ala Ala Arg Ser Tyr Gly Ala Glu
 165 170 175
 Val Thr Val Val Glu Ala Ala Pro Thr Pro Leu His Gly Ile Leu Gly
 180 185 190
 Pro Glu Leu Gly Leu Phe Thr Asp Leu His Arg Glu His Gly Val
 195 200 205
 Arg Phe His Phe Gly Ala Arg Phe Thr Glu Ile Val Gly Glu Gly Gly
 210 215 220
 Met Val Leu Ala Val Arg Thr Asp Asp Gly Glu Glu His Pro Ala His
 225 230 235 240
 Asp Val Leu Ala Ala Ile Gly Ala Ala Pro Arg Thr Ala Leu Ala Glu
 245 250 255
 Gln Ala Gly Leu Asp Leu Ala Asp Pro Glu Thr Gly Gly Val Ala
 260 265 270
 Val Asp Ala Ala Leu Arg Thr Ser Asp Pro Tyr Ile Tyr Ala Ala Gly
 275 280 285
 Asp Val Ala Ala Ala Asp His Pro Leu Leu Asp Thr Arg Leu Arg Val
 290 295 300

Glu His Trp Ala Asn Ala Leu Asn Gly Gly Pro Ala Ala Ala Arg Ala
 305 310 315 320
 Met Leu Gly Gln Asp Ile Ser Tyr Asp Arg Ile Pro Tyr Phe Phe Ser
 325 330 335
 Asp Gln Tyr Asp Val Gly Met Glu Tyr Ser Gly Tyr Ala Pro Pro Gly
 340 345 350
 Ser Tyr Ala Gln Val Val Cys Arg Gly Asp Val Ala Lys Arg Glu Phe
 355 360 365
 Ile Ala Phe Trp Leu Ala Ala Asp Gly Arg Leu Leu Ala Gly Met Asn
 370 375 380
 Val Asn Val Trp Asp Val Ala Glu Ser Ile Gln Gln Leu Ile Arg Ser
 385 390 395 400
 Gly Ala Pro Leu Glu Pro Gly Ala Leu Ala Asp Pro Gln Val Pro Leu
 405 410 415
 Ala Ala Leu Leu Pro
 420

<210> 100
 <211> 1314
 <212> DNA
 <213> Streptomyces tubercidicus

<400> 100
 atgcccgcgt cacggccgcg cttcgaccc ccgcaccgga gcggcgaccc gcctgcccc 60
 cccggggcc gtgcgcaccc accccgtgac cgtccaggac ggcatgatct acgtccatca 120
 cgcgcggag gagggcaccg ccgcataaag tcggtcgtc tcatacgggc ctcgctggcg 180
 ggcctgtacg cgcgcggtc cctgcgttcc caggggttcg acggccgcct ggtgatgtc 240
 ggggacgagt gccaacggccc ctacgaccgg ccccccgtgt ccaaggactt ctcaccggc 300
 gccaacggacc cggggccact cgccctggcc gacgcccagg agatcgccga actcgacgcc 360
 gaatggctgc tgggcacccg ggccacccgg ctcgacacccg gggacgcac ggtgctgtc 420
 gatggcggcc ggtccctgac caccgacggc gtggtcctcg ccacccggcgc cgcccccggc 480
 ctgctcccg gaccggtgcc cgccggggtc cacaccctgc gcaccctcga cgacgcccag 540
 gcgctccgtc cggatctggc gccggcgccg gtccgggtcg tggtgatcgg cggcggttc 600
 atcgccgcgc aggtcgccctc gtccctgcgc gccttaggc atgacgtcac cgtggtcgag 660
 gccgcgcgc tccccctgt cccccaactc ggccacgcca tggccagat ctgcgcgcgc 720
 ctgcacatgcgg accacggcgt cacgctgctc accggaaaccc gtgtcgcccg gtcgcgcagc 780
 gagggcgcacg gcccgcgtc caccggcgac gagctgaccg acggccgcct gtcggccgc 840
 gacgtggtcg tcgtcgccat cggggtaacgc ccccgacccg cctggctcac ggactccgga 900
 ctgcgcgtcg acgacggtgt gctctgcgc acgggtgtc tcaccccgct gcccggcgtc 960
 gtggccgtcg gcgacgtcgc cagggtggac ggcggccgtg cggagcactg gaccagcgcc 1020
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 cggagcctgc cgtacttctg gtccgaccag tacggcgtcc gcatccagtt cgccggccac 1140
 cggctgcca cccgacacacc ggcgcgtcctc gaaggctccc cccgacgaccg cagttctc 1200
 gcctgttaacg aacgggacgg acgcaccacc gcgggtgtcg ccctcaaccg gccccggcccc 1260
 ttcatgcggc tccggcgca actcgcccgcc accggccctgt cggccaccac ctga 1314

<210> 101
 <211> 437
 <212> PRT
 <213> Streptomyces tubercidicus

<400> 101

Met	Pro	Ala	Ala	Arg	Arg	Arg	Leu	Arg	Pro	Pro	His	Arg	Ser	Gly	Asp
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				20				25			30				
Gly	Arg	His	Asp	Leu	Arg	Pro	Ser	Arg	Arg	Gly	Gly	His	Arg	Arg	
				35				40			45				
Met	Lys	Ser	Val	Ala	Val	Ile	Gly	Ala	Ser	Leu	Ala	Gly	Leu	Tyr	Ala
				50				55			60				
Ala	Arg	Ser	Leu	Arg	Ser	Gln	Gly	Phe	Asp	Gly	Arg	Leu	Val	Ile	Val
				65				70			75			80	
Gly	Asp	Glu	Cys	His	Gly	Pro	Tyr	Asp	Arg	Pro	Pro	Leu	Ser	Lys	Asp
				85				90			95				
Phe	Leu	Thr	Gly	Ala	Thr	Asp	Pro	Gly	Arg	Leu	Ala	Leu	Asp	Ala	
				100				105			110				
Glu	Glu	Ile	Ala	Glu	Leu	Asp	Ala	Glu	Trp	Leu	Leu	Gly	Thr	Arg	Ala
				115				120			125				
Thr	Gly	Leu	Asp	Thr	Gly	Gly	Arg	Thr	Val	Leu	Leu	Asp	Gly	Gly	Arg
				130				135			140				
Ser	Leu	Thr	Thr	Asp	Gly	Val	Val	Leu	Ala	Thr	Gly	Ala	Ala	Pro	Arg
				145				150			155			160	
Leu	Leu	Pro	Gly	Pro	Val	Pro	Ala	Gly	Val	His	Thr	Leu	Arg	Thr	Leu
				165				170			175				
Asp	Asp	Ala	Gln	Ala	Leu	Arg	Ala	Asp	Leu	Ala	Pro	Ala	Pro	Val	Arg
				180				185			190				
Val	Val	Val	Ile	Gly	Gly	Gly	Phe	Ile	Gly	Ala	Glu	Val	Ala	Ser	Ser
				195				200			205				
Cys	Ala	Ala	Leu	Gly	His	Asp	Val	Thr	Val	Val	Glu	Ala	Ala	Pro	Leu
				210				215			220				
Pro	Leu	Val	Pro	Gln	Leu	Gly	His	Ala	Met	Ala	Glu	Ile	Cys	Ala	Ala
				225				230			235			240	
Leu	His	Ala	Asp	His	Gly	Val	Thr	Leu	Leu	Thr	Gly	Thr	Gly	Val	Ala
				245				250			255				
Arg	Leu	Arg	Ser	Glu	Gly	Asp	Gly	Arg	Arg	Val	Thr	Gly	Val	Glu	Leu
				260				265			270				
Thr	Asp	Gly	Arg	Leu	Leu	Pro	Ala	Asp	Val	Val	Val	Val	Gly	Ile	Gly
				275				280			285				
Val	Arg	Pro	Arg	Thr	Ala	Trp	Leu	Thr	Asp	Ser	Gly	Leu	Pro	Leu	Asp
				290				295			300				
Asp	Gly	Val	Leu	Cys	Asp	Ala	Gly	Cys	Val	Thr	Pro	Leu	Pro	Ala	Val
				305				310			315			320	
Val	Ala	Val	Gly	Asp	Val	Ala	Arg	Val	Asp	Gly	Ala	Arg	Ala	Glu	His
				325				330			335				
Trp	Thr	Ser	Ala	Thr	Glu	Gln	Ala	Ala	Val	Ala	Ala	Arg	Asn	Leu	Leu
				340				345			350				
Ala	Gly	Ser	Thr	Val	Ala	Thr	His	Arg	Ser	Leu	Pro	Tyr	Phe	Trp	Ser
				355				360			365				
Asp	Gln	Tyr	Gly	Val	Arg	Ile	Gln	Phe	Ala	Gly	His	Arg	Leu	Pro	Thr
				370				375			380				
Asp	Thr	Pro	Arg	Val	Leu	Glu	Gly	Ser	Pro	Asp	Asp	Arg	Ser	Phe	Leu
				385				390			395			400	
Ala	Cys	Tyr	Glu	Arg	Asp	Gly	Arg	Thr	Thr	Ala	Val	Leu	Ala	Leu	Asn
				405				410			415				

Arg Pro Arg Pro Phe Met Arg Leu Arg Arg Glu Leu Ala Arg Thr Ala
 420 425 430
 Leu Ser Ala Thr Thr
 435

<210> 102
 <211> 1233
 <212> DNA
 <213> Streptomyces tubercidicus

<400> 102
 atggcccaaga acacggcatt catcatcgcg ggagcggggc tggccggggc gaaggccgcg 60
 gagacactgc gcgcggaggg cttcggccgc cccgtcctgc tgctggcga cgagcgcgag 120
 cgtccctacg agcggccgc gctgtccaag ggctaacctt tggcacctc cgagcgggag 180
 aaggcgtacg tccatccgc ccagtggta gccgagcaco acgtcgatct gcggctggc 240
 aacgcgtca ccgcctcgaa cccggccggc cacgaggtga ccctcgccga cggcagccgg 300
 ctgggctacg ccaagctgt gctggccacc ggctccactc cgccggcgt gcccgtgccc 360
 ggcgcgcacc tcgacgggtt ccacacgctg cgtaacctgg cggacagcga ccgcctcaag 420
 gacctttcc ggtccgcgtc ccggatcgtg gtatcgccg gcggctggat cggcctggag 480
 accacggccg ccgcgcgtgc ggccggggtc gaggtgaccc tgctggagtc ggcgcgcgt 540
 cccctgctgg gggtgctggg ccgcgaggta gcccaggat tgcggatct gcacaccgag 600
 cacggtgtcg cgctgcgtc cgacacccag gtcacggaga tcaccggcac gaacggccgc 660
 gtcgacgggg tacggctggc cgacggcacc cgatcgcgg ccgacgcgt gatcgctggc 720
 gtcgggatca ccccccaactc cgagacggcc gccgcggccg ggctcaaggt cgacaacggc 780
 gtcgtcggtt acgagcggct gtgctctcc caccggaca tctacgccc cggcgaatgc 840
 gccaacgcct accacccct cctggcaag cacctccgog tcgagcactg ggccaacgcc 900
 ctccaccagc cgaagaccgc ggcccgccg atgctggccg gggaggccgg ctacgaccgg 960
 ctgcccgtact tcttcaccga ccagtacgac ctgggcatgg agtacacggg gcatgtggag 1020
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 ttctggctct ccggcggccg ggtgctggcc gggatgaatg tgaacgtatg ggacgtcacc 1140
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<210> 103
 <211> 410
 <212> PRT
 <213> Streptomyces tubercidicus

<400> 103
 Met Ala Gln Asn Thr Ala Phe Ile Ile Ala Gly Ala Gly Leu Ala Gly
 1 5 10 15
 Ala Lys Ala Ala Glu Thr Leu Arg Ala Glu Gly Phe Gly Gly Pro Val
 20 25 30
 Leu Leu Leu Gly Asp Glu Arg Glu Arg Pro Tyr Glu Arg Pro Pro Leu
 35 40 45
 Ser Lys Gly Tyr Leu Leu Gly Thr Ser Glu Arg Glu Lys Ala Tyr Val
 50 55 60
 His Pro Pro Gln Trp Tyr Ala Glu His Asp Val Asp Leu Arg Leu Gly
 65 70 75 80
 Asn Ala Val Thr Ala Leu Asp Pro Ala Gly His Glu Val Thr Leu Ala

	85	90	95												
Asp	Gly	Ser	Arg	Leu	Gly	Tyr	Ala	Lys	Leu	Leu	Leu	Ala	Thr	Gly	Ser
				100				105					110		
Thr	Pro	Arg	Arg	Leu	Pro	Val	Pro	Gly	Ala	Asp	Leu	Asp	Gly	Val	His
				115				120					125		
Thr	Leu	Arg	Tyr	Leu	Ala	Asp	Ser	Asp	Arg	Leu	Lys	Asp	Leu	Phe	Arg
				130				135					140		
Ser	Ala	Ser	Arg	Ile	Val	Val	Ile	Gly	Gly	Trp	Ile	Gly	Leu	Glu	
				145				150			155		160		
Thr	Thr	Ala	Ala	Ala	Arg	Ala	Ala	Gly	Val	Glu	Val	Thr	Val	Leu	Glu
									165		170		175		
Ser	Ala	Pro	Leu	Pro	Leu	Leu	Gly	Val	Leu	Gly	Arg	Glu	Val	Ala	Gln
									180		185		190		
Val	Phe	Ala	Asp	Leu	His	Thr	Glu	His	Gly	Val	Ala	Leu	Arg	Cys	Asp
								195		200		205			
Thr	Gln	Val	Thr	Glu	Ile	Thr	Gly	Thr	Asn	Gly	Ala	Val	Asp	Gly	Val
								210		215		220			
Arg	Leu	Ala	Asp	Gly	Thr	Arg	Ile	Ala	Ala	Asp	Ala	Val	Ile	Val	Gly
								225		230		235		240	
Val	Gly	Ile	Thr	Pro	Asn	Ser	Glu	Thr	Ala	Ala	Ala	Gly	Leu	Lys	
								245		250		255			
Val	Asp	Asn	Gly	Val	Val	Asp	Glu	Arg	Leu	Cys	Ser	Ser	His	Pro	
								260		265		270			
Asp	Ile	Tyr	Ala	Ala	Gly	Asp	Val	Ala	Asn	Ala	Tyr	His	Pro	Leu	Leu
								275		280		285			
Gly	Lys	His	Leu	Arg	Val	Glu	His	Trp	Ala	Asn	Ala	Gly	Leu	Lys	
								290		295		300			
Lys	Thr	Ala	Ala	Arg	Ala	Met	Leu	Gly	Gly	Glu	Ala	Gly	Tyr	Asp	Arg
								305		310		315		320	
Leu	Pro	Tyr	Phe	Phe	Thr	Asp	Gln	Tyr	Asp	Leu	Gly	Met	Glu	Tyr	Thr
								325		330		335			
Gly	His	Val	Glu	Pro	Gly	Gly	Tyr	Asp	Arg	Val	Val	Phe	Arg	Gly	Asp
								340		345		350			
Thr	Gly	Ala	Arg	Glu	Phe	Ile	Ala	Phe	Trp	Leu	Ser	Gly	Gly	Arg	Val
								355		360		365			
Leu	Ala	Gly	Met	Asn	Val	Asn	Val	Trp	Asp	Val	Thr	Asp	Pro	Ile	Arg
								370		375		380			
Ala	Leu	Val	Ala	Ser	Gly	Arg	Ala	Val	Asp	Pro	Glu	Arg	Leu	Ala	Asp
								385		390		395		400	
Ala	Asp	Val	Pro	Leu	Ala	Asp	Leu	Val	Pro						
					405				410						

<210> 104

<211> 1266

<212> DNA

<213> Streptomyces tubercidicus

<400> 104

gtggtcgacg cacaccagac gttcgatc gtcgggggtg gcctggccgg cgcaaaggcc 60
 gcggagactc tccgcgcgga agggttcacc ggccgggtga tcctcatctg tgacgagcgc 120
 gaccaccgt acgagcgccc cccgctatcc aaggggttcc tgctcgcaaa ggaagagcgc 180
 gacagcggtt tcgtccacga acccgcttgg tacgcccagg cacagatcga actgcacctg 240

ggccagcccg ccgtccgcct cgaccccgag gcaagaccg tccgcctcgg cgacggcacc 300
 ctgatcgact acgacaagct gctgctggcc accggcgccg agccgcgcg cctggacatc 360
 cccggcaccg gcctggccgg cgtcaccac ctgcgcgcgc tcgcccacgc cgaacggctg 420
 cgcggcgtcc tggctccct cggcgggac aacgggcattc tggatcgatcgc cggcgcggc 480
 tggatcgccc tggaggtcgc cgccgcggcc cgctccctacg gcgcccgggt caccgtcg 540
 gaggccgccc cgacaccgct gcacggcatc ctggggcccg aactcggcgg cctgttca 600
 gaactgcacc ggcacacacgg cgtgcgcctc cacttcggcg cccgtttcac cgagatcg 660
 ggacaggacg gcatggtgcg cggcgtgcgc accgacgacg gcgaggagca ccccggcc 720
 gacgtgctcg cgcgatcg cggccgcggc cgccgcgcac tcgccaaca ggccggactc 780
 gacctcgccc acccgaggc cggcggcggc gtggccgtcg acgcgacgct ggcacactc 840
 gacccgtaca tctacgcgc cggcgcacgtg gccgcccgcg accacccct cctggacacc 900
 cggctgcgcg tcgaacactg ggccaacgca ctcacggcg gcccggccgc cgccgcgc 960
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 ggcgacgtcg ccaaacggga gttcatcgatc ttctggctcg gcgaggacgg acggctgctc 1140
 gcggggatga acgtcaacgt ctgggacgtc gccgaaacca tccagcaact catccgcggc 1200
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 ccgttag 1266

<210> 105

<211> 421

<212> PRT

<213> Streptomyces tubercidicus

<400> 105

Val	Val	Asp	Ala	His	Gln	Thr	Phe	Val	Ile	Val	Gly	Gly	Gly	Leu	Ala
1									10					15	
Gly	Ala	Lys	Ala	Ala	Glu	Thr	Leu	Arg	Ala	Glu	Gly	Phe	Thr	Gly	Arg
								20		25			30		
Val	Ile	Leu	Ile	Cys	Asp	Glu	Arg	Asp	His	Pro	Tyr	Glu	Arg	Pro	Pro
								35		40			45		
Leu	Ser	Lys	Gly	Phe	Leu	Leu	Gly	Lys	Glu	Glu	Arg	Asp	Ser	Val	Phe
								50		55			60		
Val	His	Glu	Pro	Ala	Trp	Tyr	Ala	Gln	Ala	Gln	Ile	Glu	Leu	His	Leu
								65		70		75		80	
Gly	Gln	Pro	Ala	Val	Arg	Leu	Asp	Pro	Glu	Ala	Lys	Thr	Val	Arg	Leu
								85		90			95		
Gly	Asp	Gly	Thr	Leu	Ile	Ala	Tyr	Asp	Lys	Leu	Leu	Leu	Ala	Thr	Gly
								100		105			110		
Ala	Glu	Pro	Arg	Arg	Leu	Asp	Ile	Pro	Gly	Thr	Gly	Leu	Ala	Gly	Val
								115		120			125		
His	His	Leu	Arg	Arg	Leu	Ala	His	Ala	Glu	Arg	Leu	Arg	Gly	Val	Leu
								130		135			140		
Ala	Ser	Leu	Gly	Arg	Asp	Asn	Gly	His	Leu	Val	Ile	Ala	Gly	Ala	Gly
								145		150			155		160
Trp	Ile	Gly	Leu	Glu	Val	Ala	Ala	Ala	Arg	Ser	Tyr	Gly	Ala	Glu	
								165		170			175		
Val	Thr	Val	Val	Glu	Ala	Ala	Pro	Thr	Pro	Leu	His	Gly	Ile	Leu	Gly
								180		185			190		
Pro	Glu	Leu	Gly	Gly	Leu	Phe	Thr	Glu	Leu	His	Arg	Ala	His	Gly	Val
								195		200			205		
Arg	Phe	His	Phe	Gly	Ala	Arg	Phe	Thr	Glu	Ile	Val	Gly	Gln	Asp	Gly

210	215	220
Met Val Leu Ala Val Arg Thr Asp Asp Gly Glu	Glu His Pro Ala His	
225	230	235
Asp Val Leu Ala Ala Ile Gly Ala Ala Pro Arg	Thr Ala Leu Ala Glu	240
245	250	255
Gln Ala Gly Leu Asp Leu Ala Asp Pro Glu Ala	Gly Gly Val Ala	
260	265	270
Val Asp Ala Thr Leu Arg Thr Ser Asp Pro Tyr	Ile Tyr Ala Ala Gly	
275	280	285
Asp Val Ala Ala Ala Asp His Pro Leu Leu Asp	Thr Arg Leu Arg Val	
290	295	300
Glu His Trp Ala Asn Ala Leu Asn Gly Gly	Pro Ala Ala Ala Arg Ala	
305	310	315
Met Leu Gly Gln Asp Ile Ser Tyr Asp Arg Val	Pro Tyr Phe Phe Ser	320
325	330	335
Asp Gln Tyr Asp Val Gly Met Glu Tyr Ser Gly	Tyr Ala Pro Pro Gly	
340	345	350
Ser Tyr Ala Gln Val Val Cys Arg Gly Asp Val	Ala Lys Arg Glu Phe	
355	360	365
Ile Ala Phe Trp Leu Gly Glu Asp Gly Arg Leu	Leu Ala Gly Met Asn	
370	375	380
Val Asn Val Trp Asp Val Ala Glu Thr Ile Gln	Gln Leu Ile Arg Gly	
385	390	395
Gly Val Arg Leu Glu Pro Gly Glu Leu Ala Asp	Pro Glu Val Pro Leu	400
405	410	415
Thr Ser Leu Leu Pro		
420		

BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT
OF MICROORGANISMS FOR THE PURPOSE OF PATENT PROCEDURES

INTERNATIONAL FORM

TO

Syngenta
3054 Cornwallis Road
Research Triangle Park, NC 27709

VIABILITY STATEMENT

issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITORY AUTHORITY
identified at the bottom of this page

NAME AND ADDRESS OF THE PARTY TO WHOM
THE VIABILITY STATEMENT IS ISSUED

I. DEPOSITOR	II. IDENTIFICATION OF THE MICROORGANISM
Name: Syngenta 3054 Cornwallis Road Address: Research Triangle Park, NC 27709	Depositor's taxonomic designation and accession number given by the INTERNATIONAL DEPOSITORY AUTHORITY: <i>Streptomyces lividans</i> NRRL 30478 Date of: May 8, 2001 <input checked="" type="checkbox"/> ² Original Deposit <input type="checkbox"/> ² New Deposit <input type="checkbox"/> ² Repropagation of Original Deposit
III. (a) VIABILITY STATEMENT	
Deposit was found: <input checked="" type="checkbox"/> Viable <input type="checkbox"/> Nonviable on May 10, 2001 (Date) International Depository Authority's preparation was found viable on May 10, 2001 (Date) ³	
III. (b) DEPOSITOR'S EQUIVALENCY DECLARATION	
Depositor determined the International Depository Authority's preparation was <input checked="" type="checkbox"/> ² Equivalent <input type="checkbox"/> ² Not equivalent to deposit on <u>7/23/01</u> (Date) Signature of Depositor <u>Steve Hill</u>	
IV. CONDITIONS UNDER WHICH THE VIABILITY TEST WAS PERFORMED (Depositors/Depository)⁴	
V. INTERNATIONAL DEPOSITORY AUTHORITY	
Name: Agricultural Research Culture Collection (NRRL) International Depository Authority Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.	Signature(s) of person(s) having the power to represent the International Depository Authority or of authorized official(s): <i>J. Syng</i> 6-26-01

¹ Indicate the date of the original deposit or when a new deposit has been made.

² Mark with a cross the applicable box.

³ In the cases referred to in Rule 10.2(a){ii} and {iii}, refer to the most recent viability test.

⁴ Fill in if the information has been requested.

BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT
OF MICROORGANISMS FOR THE PURPOSE OF PATENT PROCEDURES

INTERNATIONAL FORM

TO

Syngenta
3054 Cornwallis Road
Research Triangle Park, NC 27709

VIABILITY STATEMENT

issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITORY AUTHORITY
identified at the bottom of this page

NAME AND ADDRESS OF THE PARTY TO WHOM
THE VIABILITY STATEMENT IS ISSUED

I. DEPOSITOR	II. IDENTIFICATION OF THE MICROORGANISM
Name: Syngenta 3054 Cornwallis Road Address: Research Triangle Park, NC 27709	Depositor's taxonomic designation and accession number given by the INTERNATIONAL DEPOSITORY AUTHORITY: <i>Pseudomonas putida</i> NRRL B-30479
	Date of: May 8, 2001
	<input checked="" type="checkbox"/> ² Original Deposit
	<input type="checkbox"/> ² New Deposit
	<input type="checkbox"/> ² Repropagation of Original Deposit
III. (a) VIABILITY STATEMENT	
Deposit was found: <input checked="" type="checkbox"/> Viable <input type="checkbox"/> Nonviable on May 10, 2001 (Date)	
International Depository Authority's preparation was found viable on May 16 2001 (Date) ³	
III. (b) DEPOSITOR'S EQUIVALENCY DECLARATION	
Depositor determined the International Depository Authority's preparation was	
<input checked="" type="checkbox"/> ² Equivalent <input type="checkbox"/> ² Not equivalent to deposit on	<u>7/23/01</u> (Date)
Signature of Depositor <u>Steve Hill</u>	
IV. CONDITIONS UNDER WHICH THE VIABILITY TEST WAS PERFORMED (Depositor/Depository)	
V. INTERNATIONAL DEPOSITORY AUTHORITY	
Name: Agricultural Research Culture Collection (NRRL) International Depository Authority	Signature(s) of person(s) having the power to represent the International Depository Authority or of authorized official(s):
Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.	<u>J. Sauer</u> Date: <u>6.26.01</u>

¹ Indicate the date of the original deposit or when a new deposit has been made.

² Mark with a cross the applicable box.

³ In the cases referred to in Rule 10.2(a)(iii) and (iv), refer to the most recent viability test.

^{*} Fill in if the information has been requested.